

Fundamentals of H₂ Binding and Reactivity on Transition Metals Underlying Hydrogenase Function and H₂ Production and Storage

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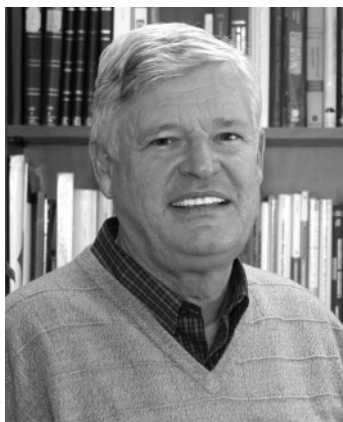
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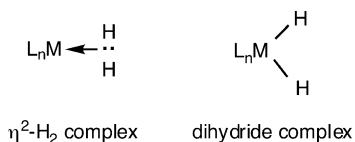
1. Introduction and Historical Perspective

Dihydrogen (H₂) is not only considered to be the fuel of the future but is also vital in chemical processes such as hydrogenation of organic compounds. Catalytic hydrogenations are the largest-volume human-made chemical reactions in the world, and all crude oil is treated with H₂ to remove sulfur and nitrogen by hydrodesulfurization and hydrodenitrogenation. Hundreds of million tons of ammonia fertilizer are produced annually from H₂ and N₂ by the Haber process which supports much of the world's population. The H₂ molecule is held together by a very strong two-electron H–H bond but is only useful chemically when the two H's are split apart in controlled fashion. To obtain proper perspective, one needs to be aware of how activation (the bond cleavage process) of H₂ occurs on metal complexes (e.g., industrial catalysts) and on enzymes in nature such as hydrogenases, which is one of the main focal points of this article. Remarkably, the detailed mechanism at the molecular level by which the H–H union splits to form for example a metal dihydride complex was not clearly established until only relatively recently in the history of H₂ activation. One of



Gregory Kubas received his B.S. from Case Institute of Technology in 1966 and his Ph.D. from Northwestern University with Duward Shriver in 1970. He performed postdoctoral studies at Princeton with Tom Spiro and moved on to Los Alamos initially as a postdoc and then as a staff member. He became a Laboratory Fellow in 1987 and more recently a Fellow of the American Association for the Advancement of Science. His discovery of metal complexes that bind dihydrogen molecules led to the 1993 American Chemical Society Award in Inorganic Chemistry and the 1994 E. O. Lawrence Award in Chemistry from the Department of Energy. His research on dihydrogen complexes led to new views of chemical bonding and hydrogen activation and opened new fields of chemical research on metal σ -bond complexes. Greg is author of the 2001 book considered to be the bible of this field, *Metal–Dihydrogen and σ -Bond Complexes*.

the reasons is that H₂ contains only a strongly bonded electron pair that was always assumed to be inert to further chemical interaction, except perhaps in a weak sense, e.g., physisorption. Thus, H₂ had never been caught in the act of chemically binding to a metal center or main group atom, usually the first step in breaking up a strong bond. The discovery by Kubas and co-workers in 1984 of coordination of a nearly intact H₂ molecule to a metal complex (L_nM; L = ligand) caught this in close detail and led to a new paradigm in chemistry.^{1–7}



The H₂ binds side-on to the metal center primarily via donation of its two σ electrons to a vacant d orbital and forms a *stable* dihydrogen complex. It is remarkable that these already strongly bonded electrons can donate to a metal center (empty d orbital) to form a nonclassical 2-electron, 3-center bond, as in other “electron-deficient” molecules such as diborane (B₂H₆) as well as the bonding in hydride-bridged⁸ M–H–M topologies. Such a complex can encompass interaction of any σ bond (C–H, Si–H, etc.) with a metal center and was termed a “ σ complex” by Crabtree.⁹

Our discovery of metal–H₂ complexes was totally unexpected. Metal dihydrides formed by oxidative addition of the H–H bond to a metal center had early on been known to be a part of well-established catalytic cycles,¹⁰ and a retrospective account of homogeneous hydrogenation was published in 1980 by a pioneer in the field, Jack Halpern.¹¹ Although some type of metal–H₂ interaction was assumed to be an intermediate in dihydride formation, it was not thought to be observable and certainly not isolable under ambient conditions. We were not seeking a dihydrogen

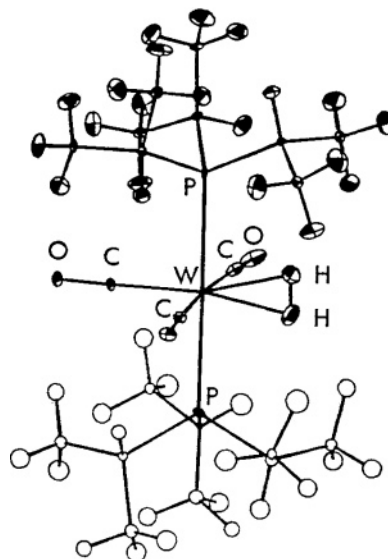
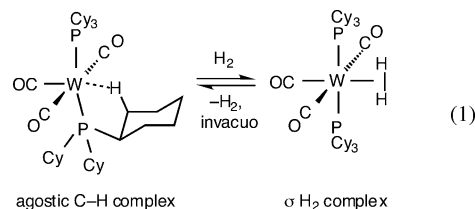


Figure 1. ORTEP drawing of the neutron structure of W(CO)₃-(PⁱPr₃)₂(H₂) at 30 K, showing the intact H–H bond elongated to 0.82(1) Å. The lower phosphine is disordered.

complex, and the first such complex, W(CO)₃(PR₃)₂(H₂) (Figure 1), was found serendipitously, an edifying saga detailed by this author.^{3,6} This stable crystalline complex was also notable in that it represented the first chemical compound isolable under ambient conditions containing a nearly intact H₂ molecule other than elemental hydrogen itself. The H–H bond length in W(CO)₃(PⁱPr₃)₂(H₂) (0.89 Å) is stretched about 20% over that in free H₂ (0.74 Å), showing that the H₂ is not physisorbed but rather chemisorbed, where the bond is “activated” toward breaking. This initially enigmatic interaction lies at the heart of all interactions of σ bonds X–Y with metals.^{5,6,9}

The serendipitous synthesis of an “unsaturated” 16-electron precursor, M(CO)₃(PCy₃)₂ (M = Mo, W; Cy = cyclohexyl), in 1979 led to the discovery of the H₂ complex.¹² This deep purple complex was a “5-coordinate” zerovalent group 6 complex, the first of its type. Importantly, the color changed instantly and reversibly to yellow on exposure to N₂ and H₂ both in solution and in the solid state, signifying adduct formation with the small molecules (eq 1). It was not until

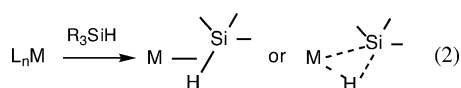


much later (1986) that a crystal structure of a tungsten analogue revealed a phosphine C–H bond weakly occupying the sixth binding site.¹³ This type of intramolecular interaction of a C–H bond had been known and has been popularly termed “agostic”.¹⁴ As here, it often serves to relieve electronic unsaturation in coordinatively unsaturated complexes that otherwise might not be stable and is entropically stabilized, i.e., a type of “chelate effect”. Importantly, H₂ was found to displace this C–H interaction in M(CO)₃-(PCy₃)₂ and could then be removed *reversibly* many times simply by exposure to vacuum or inert gas at ambient temperature to re-form the agostic complex. This property

was novel and is relevant to new materials for hydrogen storage, another subject of this article that will be discussed after the main subject, which is the relevance of H₂ complexes to hydrogen production and the function of hydrogenases.

Part of the reason that H₂ complexes were so well hidden was the stubborn notion that such complexes could not be stable versus classical dihydrides. At about the time of our finding, evidence for *unstable* M–H₂ interactions had been obtained spectroscopically by Turner, Sweany, and others via photolysis of Cr(CO)₆ in the presence of H₂ at low temperatures.^{17–20} Cr(CO)₅(H₂) was postulated based on IR CO stretching frequencies, but its molecular structure could not be determined and only recently has its proton NMR spectrum been observed, again at low temperature.^{21,22} Remarkably, even the theoretical basis for interaction of H₂ and σ bonds with a metal was still in its infancy this late in the history of inorganic chemistry. Theoretical analysis of the bonding of H₂ and CH₄ to metal fragments such as Cr(CO)₅ was published by Saillard and Hoffmann²³ in 1984, shortly after our publication of the W–H₂ complex, without mutual knowledge of our work. The interplay between theory and experiment has continued hand-in-hand to this day as one of the most valuable synergistic relations in all of chemistry.^{24,25} The apparent simplicity of H₂ was attractive, but the structure, bonding, and dynamics of complexes containing H₂ ligands proved to be unimaginably complex, resulting in abundant opportunities for study (>300 purely computational publications and dozens of others combining experiment with theory).

Initially, H₂ binding seemed unique to our M(CO)₃(PR₃)₂(H₂) complexes because the bulky phosphines (R = cyclohexyl or isopropyl) seemed to sterically inhibit formation of a classical 7-coordinate dihydride via oxidative addition. Kaesz viewed this as “arrested oxidative addition”, a term he used to describe the bonding in a silane complex, CpMn(CO)₂(η^2 -HSiPh₃).²⁶ Silane complexes^{27–29} were some of the first examples of σ -bond complexes but were initially unrecognized as such because the asymmetrically bound silane ligand lacked the superb clarity of the H₂ ligand, which has electrons only in the H–H bond. The hundreds of H₂



complexes that would be synthesized after our discovery were unimaginable to us, and it was difficult to even know where to search for new examples. It would take over a year before they were found by other researchers, most notably Morris, Crabtree, Chaudret, and Heinekey. This quartet has since performed elegant NMR and reactivity studies on H₂ and silane complexes^{9,30–35} and was later joined by well over a hundred other investigators worldwide. Remarkably, several complexes initially thought to be classical hydrides were revealed to be H₂ complexes by Crabtree beginning in 1989,^{9,36} using as criteria his finding that the H₂ ligand has very short proton NMR relaxation times ($T_1 < 100$ ms). The most interesting was RuH₂(H₂)(PPh₃)₃, originally reported in 1968 by Knoth,³⁷ which possessed unusual properties that elicited comments by Singleton in 1976 about the “dihydrogen-like nature” of the binding.³⁸ Ironically, attempts to obtain definitive proof for H₂ binding in this complex were difficult, even long after H₂ binding was established.³⁹

The variety and abundance of H₂ complexes is remarkable: about 500 H₂ complexes are known (most are stable) for nearly every transition metal and type of coligand. They are the focus of nearly 1500 publications, dozens of reviews, and three monographs.^{3,6,9,24,25,30–35,40–55} It is now clear that M–H₂ serves as the prototype for other metal σ -bond complexes^{6,9} that can be important in catalytic systems and perhaps other applied research as well. Two of the most frequently asked questions after the discovery of H₂ complexes were (1) are they relevant in catalysis, i.e., direct transfer of hydrogen from an H₂ ligand to a substrate, and (2) can methane bind to metal complexes? The answer to both is yes, although, so far, a *stable* methane complex has yet to be isolated (complexes containing higher alkanes have been reported). As will be shown, for all their apparent simplicity, M–H₂ (and other σ -bond interactions with metal centers) are arguably the most dynamic, complex, and enigmatic chemical topologies known from a structure/bonding/dynamics viewpoint. Only recently has the viewpoint on dihydrogen complexes shifted from its significance in basic science toward more practical aspects, most importantly hydrogen production and storage and the presumed intermediacy of metal–H₂ binding in biological systems such as hydrogenases. These will be the primary focal points of this article.

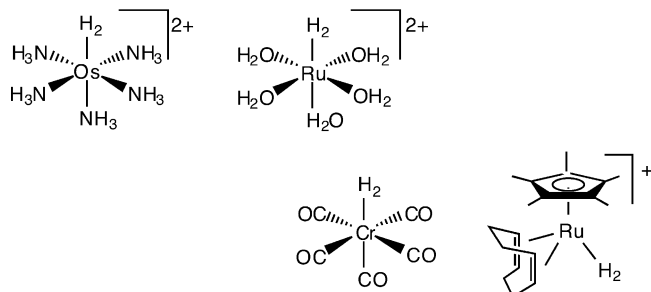
2. Types and Synthesis of H₂ Complexes

2.1. Stable H₂ Complexes

Hundreds of stable H₂ complexes have now been synthesized and characterized spectroscopically or structurally, and many others either are thermally unstable, are transient species, or are proposed to contain H₂ ligands. Almost every transition metal from V to Pt is represented (V, Ni, and Pd form only low-temperature stable species), and one lanthanide complex⁵⁶ is known. Only the very early transition metals and actinides have thus far not been observed to form stable H₂ complexes. As will be detailed below, the coupling constant J_{HD} in isotopomeric HD complexes is the best diagnostic for molecular hydrogen binding, i.e., the presence of a stretched H–H bond, and can be as high as 35 Hz versus <2 Hz for classical hydride complexes. The great majority of complexes contain octahedrally coordinated d⁶ metals that are relatively low-valent (divalent or lower), primarily because of the favorable electronic situation for side-on coordination of σ bonds to such metal centers. Virtually all H₂ complexes are coordinatively saturated, and the few that are not normally contain π -donating halide or pseudohalide ligands, e.g., RuHX(H₂)(PR₃)₂ (X = Cl, I, SR).^{57,58} Paramagnetic σ complexes are extremely rare, but apparent high-spin Fe and Mo H₂ complexes have recently been reported.⁵⁹

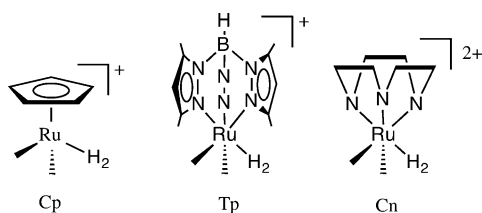
Most H₂ complexes are cationic because the increased electrophilicity of the metal reduces M → H₂ backdonation (BD) that leads to oxidative addition (OA) of H₂. Neutral complexes normally contain a mixture of donor ligands, usually phosphines, with at least one π -acceptor ligand such as CO or strong trans-effect ligands such as hydride to moderate BD, as will be discussed further below. H₂ complexes can be stabilized by classical nitrogen-donor ancillary ligands such as ammine, e.g., [Os(NH₃)₅(H₂)]²⁺, and its ethylenediamine analogues, which have very long H–H distances ($d_{HH} = 1.34$ Å) more characteristic of dihydrides.⁶⁰ These complexes indeed were initially believed to be dihydrides. As shown below, complexes containing

only aqua,⁶¹ CO,^{21,22} or carbon⁶² coligands are known but in some cases are only marginally stable. The highly acidic



pentacarbonyl Cr–H₂ complexes (and monophosphine and W derivatives) were recently observed by low-temperature NMR.^{21,22} The first example of an H₂ complex with carbene coligands, $[\text{Cp}^*\text{Ir}(\text{bis-carbene})(\text{H}_2)]^{2+}$, exhibits a much shorter H–H distance (1.04 Å) than its bis-phosphine analogues that contain highly elongated H₂ (1.45 Å).⁶³

The group 8 triad contains the overwhelming majority of dihydrogen complexes, with Ru and Os displaying the greatest variety of fragment types, especially “half-sandwich” complexes with cyclopentadienyl-type ligands (Cp, Tp, and Cn).⁴² As will be discussed in section 8.2.3, the H₂ ligands



in these and related cationic complexes can be quite acidic, especially in highly electrophilic dicationic species. The most common fragment in the group 8 triad is $[\text{MH}(\text{H}_2)\text{P}_4]^+$, where there are >45 different variants, almost half of which are for Ru (P = phosphorus donor, primarily in a planar array). Such series are ideal for correlating structural, electronic, and physical properties, e.g., H–H distance with J_{HD} , as will be discussed below.⁶⁴ This is particularly the case for the series $[\text{Os}(\text{H}_2)(\text{L})\text{N}_4]^{+/2+}$ ($\text{N}_4 = 4\text{NH}_3$ or 2 ethylenediamine), which contains over two dozen members.^{60,64a,b}

Several isoelectronic series exist across the periodic table, e.g., $\text{Mo}(\text{CO})(\text{H}_2)(\text{PP})$, $[\text{Mn}(\text{CO})(\text{H}_2)(\text{PP})]^+$, and $[\text{Fe}(\text{CO})(\text{H}_2)(\text{PP})]^{+2}$ (PP = diphosphine) and $\text{W}(\text{H}_2)(\text{CO})_3(\text{PR}_3)_2$, $[\text{Re}(\text{H}_2)(\text{CO})_3(\text{PR}_3)_2]^+$, and $[\text{Os}(\text{H}_2)(\text{MeCN})_3(\text{PR}_3)_2]^{+2}$.^{5,6} The dicationic complexes of iron, the metal most relevant to biological enzymes such as hydrogenases, often can bind H₂ more tightly than the cationic or neutral analogues because increased electron donation from H₂ offsets decreased backdonation (BD) from the metal. Note that the Os complex does not contain π -acceptor CO ligands that generally stabilize H₂ coordination against oxidative addition to hydride ligands. Instead, the dipositive charge on the metal reduces backdonation that otherwise might promote oxidative addition. Highly electrophilic cationic metals are thus excellent targets for design of σ complexes because increased σ donation to M stabilizes the interaction but can never cause the σ bond to rupture.

Isolable bis-H₂ complexes are rare, e.g., $\text{RuH}_2(\text{H}_2)_2(\text{PR}_3)_2$ (R = cyclohexyl (Cy) and cyclopentyl (Cyp)),^{35,65} $[\text{RhH}_2(\text{H}_2)_2(\text{PCy}_3)_2]^+$,⁶⁶ and $\text{Tp}^*\text{RuH}(\text{H}_2)_2$.⁶⁷ The first neutron diffraction structure of a bis-H₂ complex was determined on

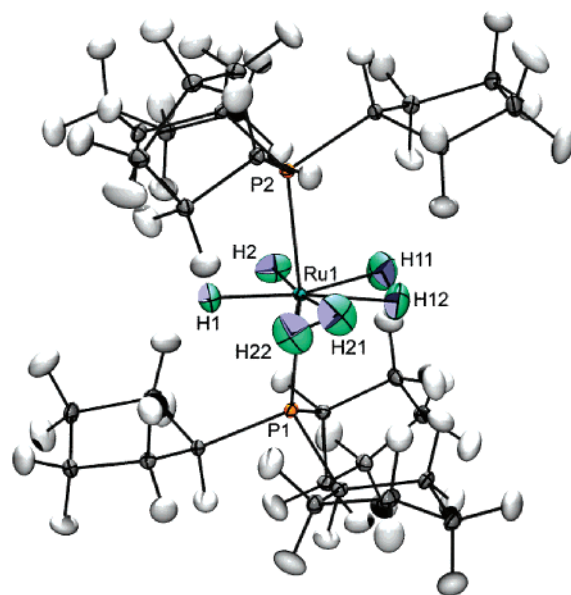
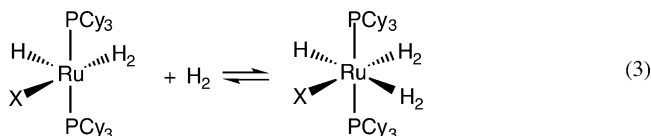


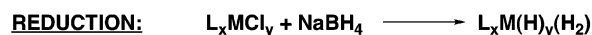
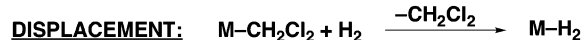
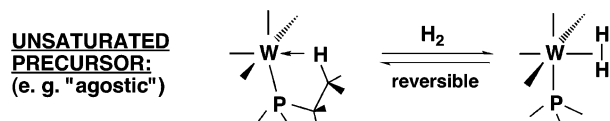
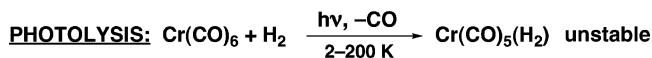
Figure 2. Structure of $\text{RuH}_2(\text{H}_2)_2(\text{PCy}_3)_2$ from a neutron diffraction study.

$\text{RuH}_2(\text{H}_2)_2(\text{PCy}_3)_2$ and showed cis-H₂ ligands with very short $d_{\text{HH}} = 0.825(8)$ Å (Figure 2).⁶⁵ The novel, X-ray characterized 16e species $\text{RuHX}(\text{H}_2)(\text{PCy}_3)_2$ (X = Cl, I) add a second H₂ ligand in equilibrium fashion (eq 3, observable only in solution).^{57,58}



Only about a dozen polynuclear dihydrogen complexes are known, and these are primarily dinuclear hydride- and/or halide-bridged Ru, Os, and Ir complexes containing H₂ bound to only one of the metals.⁶ Bridging H₂ ligands have not been definitively proven by diffraction methods, and indeed, it can be extremely difficult to determine conclusively whether or not even mononuclear complexes contain classical hydride ligands versus a nonclassical H₂ ligand (or how many of each). This is especially a problem in polyhydride complexes that contain both classical hydrides and η^2 -H₂ that undergo dynamic exchange even at the lowest temperature accessible by solution NMR. The classic example is $\text{RuH}_2(\text{H}_2)(\text{PPh}_3)_3$, which, as mentioned above, had long been speculated to contain molecular H₂ binding but had defied attempts to definitively prove it by diffraction methods.³⁹ Not surprisingly, as shown by Heinekey,⁶⁸ there have been cases where misassignments have been made, even for complexes containing only two hydrogens on a metal. About a dozen complexes exist that possibly may contain coordinated H₂ and/or have d_{HH} in the “gray zone” (1.4–1.6 Å) between formulation as H₂ or dihydride complexes. Such complexes have been referred to as “compressed hydrides” with NMR features differing from elongated H₂ complexes; for example, J_{HD} increases with temperature for the former and decreases for the latter.^{34,69} These are relative terms, since the H–H bond is always stretched on binding, and indeed, as will be shown below, a near continuum of d_{HH} exists.^{6,35,69,70}

Dihydrogen complexes may also exist in solutions of organometallic complexes as equilibrium or transient species



PROTONATION OF HYDRIDE:

by acids as weak as ROH

Deprotonation of bound H₂ by bases as weak as Et₃O

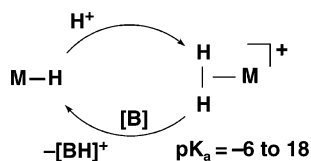
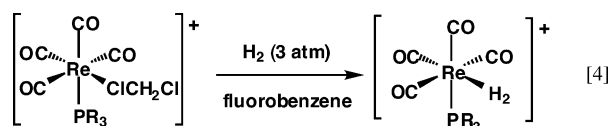


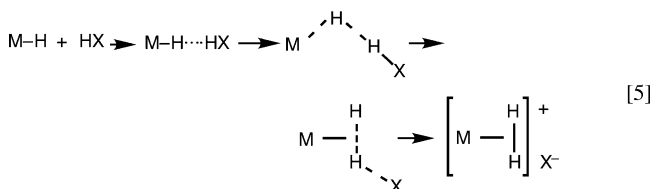
Figure 3. Synthetic methods for H₂ complexes.

that cannot be observed spectroscopically. Weak interactions of H₂ with surface species, bare metal ions, and main group Lewis acids/bases are known and will be discussed in sections 2.2.2 and 11.3. Short d_{HH} as low as 1.5 Å ("hydrogen pairing") are proposed to be present in certain intermetallic rare-earth hydrides, as evidenced by solid state ¹H NMR^{71,72} and theoretical calculations.⁷³ The observation, for example, of a characteristic splitting pattern (Pake doublet) at 140 K gives a d_{HH} of 1.48 ± 0.02 Å in CeNiInH_{1.0}, suggesting that the hydrogens may occupy nearest-neighbor tetrahedral sites separated by about 1.5 Å (2.1 Å had generally been believed to be the closest possible spacing in metal hydrides).⁷¹

Several synthetic routes to H₂ complexes are available (Figure 3) and will be discussed in detail below. The simplest method is reaction of H₂ gas with a coordinatively unsaturated complex or one that is effectively unsaturated, such as W(CO)₃(PR₃)₂, which contains an agostic interaction of a C–H bond weakly occupying the sixth site (eq 1). Displacement of a weakly bound "solvento" ligand such as dichloromethane or a coordinated anion can be utilized, although a less coordinating solvent such as fluorobenzene may need to be employed.⁷⁴ By far the most common method of



preparation is protonation of metal hydride complexes (eq 5).^{33,44,55,75} Reaction proceeds via observable hydrogen bond-

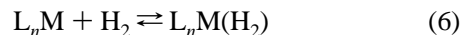


ing of the acid (which can be as weak as alcohols) to the basic hydride.^{55,75} This method has been widely applicable because it does not require an unsaturated precursor that often either does not exist or is difficult to synthesize. Neutral polyhydride complexes L_nMH_x are often easy targets for protonation to cationic hydrido-H₂ complexes, [L_nM(H₂)H_{x-1}]⁺,

which frequently are more robust than complexes prepared from H₂ gas.

2.1.1. Complexes Synthesized by Addition of H₂ Gas to an Unsaturated Precursor

A common method of preparation is the reaction of H₂ gas at about 1 atm pressure with a coordinatively unsaturated precursor complex, ML_n (eq 6):



The precursor complex can be a formally 16e species possessing an agostic C–H interaction that is in effect displaced by the incoming H₂ ligand, as was shown above in eq 1. The agostic interaction can readily displace the η²-H₂ if excess H₂ is not present, facilitating the reversibility of the binding. This is the case for the original series of H₂ complexes, M(H₂)(CO)₃(PR₃)₂ (M = Cr, Mo, W; R = Cy, *i*-Pr) and certain others formed directly by reversible addition of H₂ gas to an isolated, formally unsaturated, precursor complex (Table 1). Virtually all of the precursors are "operationally unsaturated", i.e. formally 16e species stabilized by agostic interactions, π-donation from halide ligands, or hydride ligands. In a few cases, the precursor has an anion such as triflate or solvent (e.g. CH₂Cl₂) occupying the coordination site that can reversibly be displaced by H₂, as in eq 4 above and further discussed below. The percentage of H₂ complexes synthesized by H₂ addition to precursors is actually surprisingly small (~10–15%). The reactions are generally carried out in noncoordinating or weakly coordinating organic solvents such as toluene or CH₂Cl₂, although solid–gas reactions can also be used.^{76–79} Low-coordinating anions such as B[3,5-C₆H₃(CF₃)₂]₄[−], abbreviated as BAR_f, are often needed to stabilize cationic M and prevent anion binding to M, especially for M = Mn, Re in Table 1. For example, the complex [Re(H₂)(CO)₃(PCy₃)₂]⁺ with BF₄ anion loses H₂ at low temperature, but the complex with less coordinating BAR_f can be isolated as a solid at room temperature.⁸⁰

2.1.2. Complexes with the Most Weak, Reversible H₂ Binding and the Shortest H–H Distances

The Cr(H₂)(CO)₃(PR₃)₂ complexes are among the most unstable H₂ complexes isolable as solids at 25 °C.⁸² The deep-blue precursor, Cr(CO)₃(PCy₃)₂, was prepared initially by Hoff.⁸³ In solution, the latter binds H₂ (or N₂) only at high pressures (>10 atm). The H₂ complex is stable under H₂ but, immediately on dissolving in toluene, loses all bound H₂ as H₂ gas, which vigorously effervesces from solution to give a deep-blue solution of Cr(CO)₃(PCy₃)₂. Such a large difference in stability between solution and solid states is rare in chemistry. It appears that coordinated H₂ can effectively be "trapped" in the less flexible solid state, possibly as a result of product solubility differences. This is reasonable in that the H₂ is not merely leaving the coordination site in these complexes; the whole molecule must rearrange to give back the agostic interaction with more acute P–Cr–P, Cr–P–C, and P–C–C bond angles. Also, in toluene, transient solvent binding might induce rapid H₂ loss kinetically by mass action effects, although hydrocarbon binding could never actually be observed by NMR for any of these group 6 systems, even at low temperature. Evidence for H₂ substitution by hydrocarbon solvents (toluene or even hexane) is seen for the series of iridium(III) complexes,

Table 1. Complexes Prepared by Reversible Addition of H₂ to a Known Precursor Complex

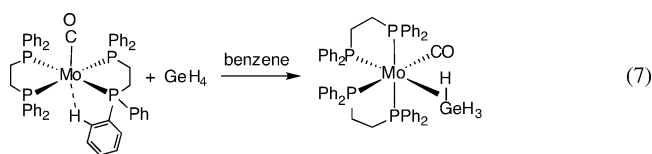
complex ^a	precursor structure	H ₂ lability	ref
M(H ₂)(CO) ₃ (PR ₃) ₂ (M = Cr, Mo, W)	agostic	v high to med	2, 82 ^b
<i>trans</i> -Mo(H ₂)(CO)(PP) ₂	agostic	med	88c, 89
[Mn(H ₂)(CO) ₃ (P) ₂] ⁺	agostic, ^c solvento ^d	high	99
<i>trans</i> -[Mn(H ₂)(CO)(PP) ₂] ⁺	agostic	high	97, 98, 100
<i>trans</i> -[Mn(H ₂)(CO)-{P(OR) ₃ } ₄] ⁺	agostic?	high	<i>e</i>
Tc(H ₂)Cl(dppe) ₂	trig bipy	med	<i>f</i>
[Re(H ₂)(CO) ₃ (PR ₃) ₂] ⁺	agostic	med	80, 204c, 360 ^g
[Re(H ₂)(CO) ₄ (PR ₃) ₃] ⁺	solvento	high	74, 277
[Re(H ₂)(CO) ₂ (triphos) ₂] ⁺	agostic	med	<i>h</i>
[CpRu(tmeda)(H ₂) ⁺	2-leg piano stool	low	76, 77
[Ru(H ₂)H(PP) ₂] ⁺		med to high	202 ^{i-k}
[M(H ₂)(CN)(PP) ₂] ⁺ (M = Fe, Ru)	anion-coord	med	274, 275
[M(H ₂)(L)(PP) ₂] ²⁺ (M = group 8; L = CO, CNH)	anion-coord	med	266, 274, 275 ^l
Ru(H ₂)H ₂ (CO)(P ⁱ Bu ₂ Me) ₂	sq pyr	high	<i>m, n</i>
[Ru(H ₂)Cl(PP) ₂] ⁺	trig bipy	v high to med	105–109 ^l
Ru(H ₂)Cl ₂ (P-N)(PR ₃)	sq pyr	high	<i>o</i>
M(H ₂)Cl(H)(CO)(PPR ₃) ₂ (M = Ru, Os)		med	<i>p, q</i>
(H ₂)(dppb)Ru(<i>m</i> -Cl) ₃ -RuCl(dppb)	dimer	high	<i>r, s</i>
[Os(H ₂)Cl(PP) ₂] ⁺	trig bipy	low	225 ^{c,l}
OsH ₃ Cl(H ₂)(P ⁱ Pr ₃) ₂	distorted oct	low	<i>t, u</i>
OsH ₂ (X)(Y)(H ₂)(P ⁱ Pr ₃) ₂ (X, Y = Cl, Br, I)	distorted oct	med	<i>t, u</i>
Ir(H ₂)H ₂ Cl(PR ₃) ₂	trig bipy	v high	78, 79
<i>trans</i> -Ir(H ₂)HX ₂ (PR ₃) ₂ (X = Cl, Br)	sq pyr?	v high	166 ^{v,w}
Ir(H ₂)(H)(diphpyH)(PR ₃) ₂	agostic	med	<i>x</i>
[PtH(H ₂)(PR ₃) ₂] ⁺	anion/ solvento	v high	101 ^{y,z}

^a Abbreviations: P-N = *o*-diphenylphosphino-*N,N*-dimethylaniline; diphpyH = 2,6-diarylpyridine. ^b Khalsa, G. R. K.; Kubas, G. J.; Unkefer, C. J.; Van Der Sluis, L. S.; Kubat-Martin, K. A. *J. Am. Chem. Soc.* **1990**, *112*, 3855. ^c P = PCy₃. ^d P = P{(OCH₂)₃CMe}₂. ^e Albertin, G.; Antonietti, S.; Bettiol, M.; Bordignon, E.; Busatto, F. *Organometallics* **1997**, *16*, 4959. ^f Burrell, A. K.; Bryan, J. C.; Kubas, G. J. *J. Am. Chem. Soc.* **1994**, *116*, 1575. ^g Albertin, G.; Antonietti, S.; Garcia-Fontan, S.; Carballo, R.; Padoan, F. *J. Chem. Soc., Dalton Trans.* **1998**, 2071. ^h Bianchini, C.; Marchi, A.; Marvelli, L.; Peruzzini, M.; Romerosa, A.; Rossi, R.; Vacca, A. *Organometallics* **1995**, *14*, 3203. ⁱ Saburi, M.; Aoyagi, K.; Takahashi, T.; Uchida, Y. *Chem. Lett.* **1990**, 601. ^j Jimenez-Tenorio, M.; Puerta, M. C.; Valerga, P. *J. Am. Chem. Soc.* **1993**, *115*, 9794. ^k Schlaf, M.; Lough, A. J.; Morris, R. H. *Organometallics* **1997**, *16*, 1253. ^l Rocchini, E.; Mezzetti, A.; Ruegger, H.; Burckhardt, U.; Gramlich, V.; Del Zotto, A.; Martinuzzi, P.; Rigo, P. *Inorg. Chem.* **1997**, *36*, 711. ^m Poulton, J. T.; Sigala, M. P.; Eisenstein, O.; Caulton, K. G. *Inorg. Chem.* **1993**, *32*, 5490. ⁿ Heyn, R. H.; Macgregor, S. A.; Nadasdi, T. T.; Ogasawara, M.; Eisenstein, O.; Caulton, K. G. *Inorg. Chim. Acta* **1997**, *259*, 5. ^o Mudalige, D. C.; Rettig, S. J.; James, B. R.; Cullen, W. R. *J. Chem. Soc., Chem. Commun.* **1993**, 830. ^p Gusev, D. G.; Vymenits, A. B.; Bakhmutov, V. I. *Inorg. Chem.* **1992**, *31*, 1. ^q Esteruelas, M. A.; Sola, E.; Oro, L. A.; Meyer, U.; Werner, H. *Angew. Chem., Int. Ed. Engl.* **1988**, *27*, 1563. ^r Joshi, A. M.; James, B. R. *J. Chem. Soc., Chem. Commun.* **1989**, 1785. ^s Chau, D. E. K.-Y.; James, B. R. *Inorg. Chim. Acta* **1995**, *240*, 419. ^t Gusev, D. G.; Kuznetsov, V. F.; Eremenko, I. L.; Berke, H. *J. Am. Chem. Soc.* **1993**, *115*, 5831. ^u Kuhlman, R. L.; Gusev, D. G.; Eremenko, I. L.; Berke, H.; Huffman, J. C.; Caulton, K. G. *J. Organomet. Chem.* **1997**, *536–537*, 139. ^v Gusev, D. G.; Bakhmutov, V. I.; Grushin, V. V.; Volpin, M. E. *Inorg. Chim. Acta* **1990**, *177*, 115. ^w Bakhmutov, V. I.; Vymenits, A. B.; Grushin, V. V. *Inorg. Chem.* **1994**, *33*, 4413. ^x Albeniz, A. C.; Schulte, G.; Crabtree, R. H. *Organometallics* **1992**, *11*, 242. ^y Gusev, D. G.; Notheis, J. U.; Rambo, J. R.; Hauger, B. E.; Eisenstein, O.; Caulton, K. G. *J. Am. Chem. Soc.* **1994**, *116*, 7409. ^z Stahl, S. S.; Labinger, J. A.; Bercaw, J. E. *Inorg. Chem.* **1998**, *37*, 2422.

IrXH₂(H₂)(PⁱPr₃)₂ (X = Cl, Br, I), which, like the Cr(0) complex, readily liberates H₂ on dissolution in hydrocarbons.⁸⁴ The Cr and Ir complexes contain the most weakly, reversibly bound H₂ ligands in an isolable species. They have very short *d*_{HH} (0.85 Å, solid-state NMR for Cr⁸² and neutron

diffraction for Ir⁸⁵), and the Cr complex has one of the highest *J*_{HD} measured, 35 Hz, for all H₂ complexes. Cr(CO)₅(H₂), which is stable only at low temperature, has *J*_{HD} = 35.8 Hz, which would correspond to *d*_{HH} = 0.84 Å from a known correlation (eqs 18 and 19 below).^{21,22} The highest value for an isolated complex, 37 Hz, has been reported for [RuH(H₂)(BINAP)(dpen)]⁺, although no structural details are available.⁸⁶

In addition to mass action effects, *entropy* effects are also often critical in determining the relative stabilities of these weak complexes because enthalpies of ligand binding can be as low as 15 kcal/mol for M–H₂ or even lower for alkane complexes. This is particularly true when σ ligands are competing for binding sites against external ligands such as H₂O and N₂ and at the same time against intramolecular agostic interactions. The latter are favored because addition of an external ligand (“two particles to form one”) has an *entropic* cost, *TΔS*, of ≈10 kcal/mol at room temperature.⁸⁷ Other complexes prepared according to eq 6 are listed in Table 1 along with the structure of the precursor complex if known. Several 16e precursors have true 5-coordinate geometries without agostic interactions, and H₂ binds highly reversibly to them. The 16e complex, Mo(CO)(Ph₂PC₂H₄-PPh₂)₂, was the first to show coordination of H–H, Si–H (silane coordination), and agostic C–H bonds to the same metal fragment and also coordinates germanes, HGeR₃, via Mo(η²-Ge–H) bonding, including GeH₄.^{88–91}

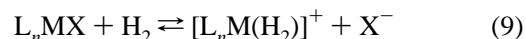


2.1.3. Complexes Prepared from H₂ Gas by Ligand Displacement or Reduction

A related method of synthesis from H₂ gas involves displacement of a labile ligand (eq 8)

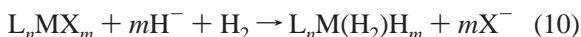


Neutral ligands L' which have been displaced include H₂O,^{61,92,93} N₂,^{94,95} NH₃,⁹⁶ CH₂Cl₂,^{74,97–101} and PMe₂Ph.¹⁰² One of the simplest conceivable H₂ complexes, [Ru(H₂O)₅(H₂)]²⁺, is formed by displacement of an aqua ligand from the hexaqua complex by pressurized H₂ in aqueous solution.⁶¹ Although it cannot be isolated, NMR indicates it has *d*_{HH} of 0.90 Å on the basis of the observed *J*_{HD} of 31.2 Hz. Displacement of a charged ligand, X[−], by H₂ has occasionally been employed for synthesis (eq 9).



Complexes prepared as in eq 9 are [M(H₂)H(depe)₂]⁺, M = Fe, Ru, Os,¹⁰⁴ [M(H₂)Cl(depe)₂]⁺, M = Ru, Os,^{105–108} [Ru(H₂)H(dcyep)₂]⁺,¹⁰⁹ and [Os(H₂)H(CO)(P-*i*-Pr₃)₂]⁺, where X = BH₄[−].¹¹⁰ Often, a group 1 metal cation such as Na⁺ or alternatively Tl⁺ is present to precipitate with the anion. Remarkably, H₂ directly displaces a normally strongly bound chloride ligand in Re(CN-*t*-Bu)₃(PCy₃)₂Cl in CH₂Cl₂, without such help to give [Re(CN-*t*-Bu)₃(PCy₃)₂(H₂)]Cl where the Cl becomes the counteranion.¹¹¹

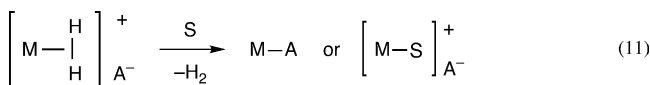
The syntheses of polyhydride complexes containing η²-H₂, such as RuH₂(H₂)(PPh₃)₃, can be accomplished by hydride reduction according to eq 10.¹¹²



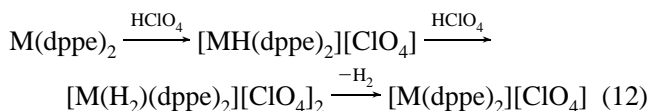
Common sources of hydride in eq 10 are NaH, NaBH₄, and LiAlH₄, and the anion, X⁻, is usually chloride or bromide. Complexes include ReH₇(PR₃)₂,¹¹³ [FeH(H₂)(pp₃)]⁺,¹¹⁴ M(H₂)-H₂(PR₃)₃ (M = Fe, Ru),^{36,38,115} Ru(H₂)H₂(cytpp),^{116,117} and Rh(H₂)H₂(HB(3,5-Me₂pZ)₃).¹¹⁸

2.1.4. Protonation of a Hydride Complex

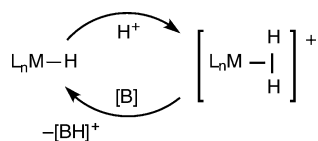
A very common and convenient method of preparation of H₂ complexes is the addition of H⁺ to a hydride or polyhydride complex, as shown in eq 5 above. In most cases, the resulting complex is cationic, and the proton source can range from strong acids such as HBF₄·Et₂O or triflic acid to very weak acids, even alcohols. The reactions are usually carried out below room temperature (ca. -60 °C), especially with strong acids, which often need to have low-interacting anions such as BF₄ or BAr_f. This method was first employed by Crabtree in 1985 by reaction of IrH₂(PPh₃)₂(bq) (bq = benzoquinolate) with PhCH(SO₂CF₃)₂,^{92,93} and a variety of H₂ complexes too numerous to list in detail have been prepared by protonation. The large class of half-sandwich complexes, [Cp'M(H₂)(L)(L')] ⁺ (M = Fe, Ru, Os; Cp' = cyclopentadienyl derivative), have all been prepared by protonation, for example. Normally, the low-temperature protonation initially gives a [M-H₂]⁺ complex, but on warming, rearrangement to a dihydride or equilibrium mixture sometimes results. Occasionally the product is unstable toward loss of H₂ and coordination of anion or solvent (S) if the electronics and thermodynamics of the system do not favor H₂ binding. The stability of H₂



complexes prepared by protonation thus varies greatly: some are stable only below room temperature and cannot be isolated as solids, and others are among the most robust H₂ complexes known. Generally, the lability of an H₂/hydride system increases upon protonation or multiple protonation. Thus, M(dppe)₂ (M = Ni, Pd, Pt) had been reported in 1966 to give a dicationic complex on double protonation (eq 12), which in light of current knowledge can be speculated to occur via a monohydride and an unstable H₂ complex, which readily loses H₂.¹¹⁹



Needless to say, complexes formed by protonation, especially where HA is a strong acid, are readily *deprotonated*, even by bases [B] as weak as diethyl ether, and are highly sensitive to solvent media and trace water. Much of these properties

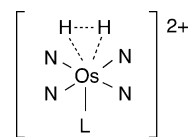


relate to the high acidity of certain H₂ complexes, which

can have pK_a as low as -6, e.g., when generated from triflic acid, as will be discussed below in section 8.2.3.

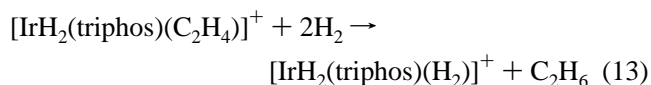
2.1.5. Other Methods of Preparation

Some less common preparations have been reported. The reduction of complexes of Re^V or Os^{III} in the presence of a source of protons and electrons (H⁺ and Mg or Na) gives the complexes ReCl(H₂)(PMePh₂)₄¹²⁰ and [Os(H₂)(NH₃)₅]²⁺, respectively. The latter and its ethylenediamine (en) congeners are unique in containing pure σ-donor ligands, and a large series of such complexes have been prepared with a variety of ligands (L) trans to the H₂.^{60,121-126} The dipositive



charge is rare among H₂ complexes and undoubtedly is responsible for arresting oxidative addition. However, the *d*_{HH} is very long, ca. 1.35 Å, in these species, indicating they are closer to being dihydrides. The reaction of Ru(cod)(cot) with PCy₃ and H₂ gives RuH₂(H₂)₂(PCy₃)₂,³⁵ and protonation¹²⁷ of [RuH₅(PⁱPr₃)₂]⁻ gives RuH₂(H₂)₂(PⁱPr₃)₂. These are among only a handful of well-characterized complexes that contain more than one η²-H₂ and have received extensive study by Chaudret and co-workers.³⁵

Decomposition of OsH(η²-H₂BH₂)(CO)(PⁱPr₃)₂ in alcohols produced OsH₂(H₂)(CO)(PⁱPr₃)₂,¹²⁸ which, despite its facile loss of H₂ and wide use as a hydrogen transfer catalyst, was initially believed to be a tetrahydride and was not shown¹²⁹ to have an η²-H₂ ligand until 10 years after its original synthesis. This is yet another dramatic example of how difficult it can be to prove the presence of H₂ ligands. Another unusual synthesis involves hydrogenation of an ethylene complex either in solution or even in the solid state at 60 °C (eq 13).^{130,131}



2.2. H₂ Complexes Unstable at Room Temperature

Many H₂ complexes are unstable at room temperature, in some cases those formed by protonation (eq 5). However, they often can still be studied by low-temperature NMR methodologies and determined to have η²-H₂ by measurement of *J*_{HD} and *T*₁. Virtually any metal system that eliminates H₂ gas via any route (protonation, photolysis, heating, etc.) must do so by a transient H₂ complex as demanded by the principle of microscopic reversibility. Obviously, the transient will have widely varying degrees of stability, roughly corresponding to the various points along the reaction coordinate toward OA along which H₂ complexes can be arrested. The sections below describe identification of H₂ complexes by non-NMR methods.

2.2.1. Organometallic Complexes Observed at Low Temperature in Rare Gas or Other Media

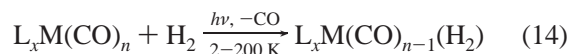
The first spectroscopic evidence for H₂ coordination was obtained in matrix-isolated Cr(CO)₅(H₂) by Sweany virtually at the same time as that for W(CO)₃(PR₃)₂(H₂). The investigations of low-*T* stable H₂ complexes (Table 2) in solid

Table 2. Low-Temperature-Stable H₂ Complexes and Surface-Bound H₂

complex	conditions	$\nu(\text{H-H}), \text{cm}^{-1}$	ref
ScH _x (H ₂) _n ($x = 2, 3$)	Ar matrix		<i>a</i>
YH ₂ (H ₂) _n	Ar matrix		<i>a</i>
V(CO) ₅ (H ₂)	heptane, Xe soln		<i>b</i>
CpV(CO) ₃ (H ₂)	heptane, Xe soln	2642	<i>c-e</i>
CpNb(CO) ₃ (H ₂)	heptane, Xe soln	2600 (equil)	<i>c-f</i>
(indenyl)Nb(CO) ₃ (H ₂)	heptane, Xe soln, PE		<i>e, f</i>
CrH(H ₂) ₂ ($X = 1, 2; n = 1, 2$)	Ar, Ne matrix		<i>g, h</i>
CrO ₂ (H ₂)	laser ablation, IR, theory	[3950] _{calc}	<i>i</i>
CrO ₂ (H ₂) ₂	laser ablation, IR, theory	2728, 2640	<i>i</i>
Cr(CO) ₅ (H ₂)	matrix, Xe soln, PE	3030	18–20, 22 ^j
	heptane, photoacoustic		134 ^k
	matrix, Xe soln		18, 19b
Cr(CO) ₄ (H ₂) ₂	Xe soln		<i>l-n</i>
Cr(CO) _n (L)(H ₂) ($n = 3, 4; L = \text{olefin or diolefin}$)	Xe soln		<i>o</i>
(arene)Cr(CO) ₂ (H ₂)	Xe soln, PE	3080	19b, 134
Mo(CO) ₅ (H ₂)	heptane, photoacoustic		<i>j</i>
	matrix		<i>p, q</i>
CpMoH(CO) ₂ (H ₂)	Ar matrix, Xe soln, PE		<i>m, n, r</i>
Mo(CO) _n (L)(H ₂) ($n = 3, 4; L = \text{olefin or diolefin}$)	matrix	3200	<i>s</i>
(arene)Mo(CO) ₃ (H ₂)	Kr, Xe matrix, laser ablated		<i>t</i>
Mo(H ₂) _n ($n = ?$)	matrix, soln, and gas phases, PE	2711	19b, 22, ^{o,u} 134
W(CO) ₅ (H ₂)	Xe soln		<i>l-n</i>
W(CO) _n (L)(H ₂) ($n = 3, 4; L = \text{olefin or diolefin}$)	matrix		<i>p, q</i>
CpWH(CO) ₂ (H ₂)	gas phase, scCO ₂		<i>x</i>
CpMn(CO) ₃ (H ₂) ($x = 1, 2$)	Xe soln		<i>o</i>
Cp*Mn(CO) ₂ (H ₂)	heptane		<i>y</i>
(C ₅ E ₅)Mn(CO) ₂ (H ₂)	matrix		<i>z</i>
MnX(CO) ₄ (H ₂) ($x = \text{Cl, Br}$)	Xe soln	2973	<i>aa</i>
Fe(CO)(NO) ₂ (H ₂)	Xe soln		<i>o</i>
Fe(C ₄ H ₄)(CO) ₂ (H ₂)	PE		<i>r</i>
Fe(CO) ₃ (H ₂)(DF)	Xe soln	{2976, 3100}	<i>aa</i>
Co(CO) ₂ (NO)(H ₂)	matrix		<i>bb</i>
CoH(H ₂)(CO) ₃	matrix		<i>bb</i>
Co(CH ₃)(H ₂)(CO) ₃	matrix		<i>cc</i>
CpIr(CO)(H ₂)	matrix		<i>dd</i>
Ru(H ₂) _x (CO) _n ($x = 1, 2; n = 1, 2$)	Ar matrix, laser ablated		
RuO ₂ (110)(H ₂)	surface, HREELS	2960	471, 472
RhH _x (H ₂) ($x = 0-3$)	Ar, Ne matrix, theory		<i>g</i>
[RhH ₂ (H ₂)] ⁻	Ar, Ne matrix, theory		<i>g</i>
Ni(CO) ₃ (H ₂)	Ar matrix		96
NiCp ₂ '(H ₂)	matrix	3250	<i>s</i>
Ni(510)(H ₂)	surface, EELS	3205	195
Ni(111)(H ₂)	surface, HREELS		<i>ee</i>
Pd(H ₂) _x ($x = 1-3$)	Kr, Xe matrix, laser ablated	2971 ($x = 1$)	152
Pd ₂ (H ₂)	laser ablated		152b
Pd(210)(H ₂)	surface, HREELS		<i>ff</i>
Cu ₂ H ₂ (H ₂) _x ($x = 1, 2$)	Ar matrix		457
Cu ₃ (H ₂)	Ar matrix		457
CuCl(H ₂)	Ar matrix		<i>gg</i>
[Cu ⁺ -zeolite-H ₂]	theory, IR		<i>hh</i>
[Cu ₂ -(H ₂) _n] ⁺	mass spec, surface ionization		<i>ii</i>
CuH(H ₂)	theory, matrix		<i>jj, kk</i>
AgH(H ₂)	theory, matrix		<i>jj, kk</i>
AuH _x (H ₂) ($x = 1, 3$)	Ar, Ne matrix, theory		<i>jj-oo</i>
MH ₂ (H ₂) ($M = \text{La, Ce, Pr}$)	Ar matrix, theory		<i>a, pp</i>

^a Wang, X.; Chertihin, G. V.; Andrews, L. *J. Phys. Chem. A* **2002**, *106*, 9213. ^b George, M. W.; Haward, M. T.; Hamley, P. A.; Hughes, C.; Johnson, F. P. A.; Popov, V. K.; Poliakov, M. *J. Am. Chem. Soc.* **1993**, *115*, 2286. ^c Haward, M. T.; George, M. W.; Howdle, S. M.; Poliakov, M. *J. Chem. Soc., Chem. Commun.* **1990**, 913. ^d Haward, M. T.; George, M. W.; Hamley, P.; Poliakov, M. *J. Chem. Soc., Chem. Commun.* **1991**, 1101. ^e Childs, G. I.; Gallagher, S.; Bitterwolf, T. E.; George, M. W. *J. Chem. Soc., Dalton Trans.* **2001**, 1711. ^f Wang, X.; Andrews, L. *J. Phys. Chem. A* **2002**, *106*, 3706. ^g Wang, X.; Andrews, L. *J. Phys. Chem. A* **2003**, *107*, 570. ^h Zhou, M.; Zhang, L.; Shao, L.; Wang, W.; Fan, K.; Qin, Q. *J. Phys. Chem. A* **2001**, *105*, 10747. ⁱ Walsh, E. F.; Popov, V. K.; George, M. W.; Poliakov, M. *J. Phys. Chem.* **1995**, *99*, 12016. ^j Poliakov, M.; Howdle, S. M.; George, M. W. *Process Technol. Proc.* **1996**, *12*, 67. ^k Jackson, S. A.; Upmacis, R. K.; Poliakov, M.; Turner, J. J.; Burdett, J. K.; Grevels, F.-W. *J. Chem. Soc., Chem. Commun.* **1987**, 678. ^l Jackson, S. A.; Hodges, P. M.; Poliakov, M.; Turner, J. J.; Grevels, F.-W. *J. Am. Chem. Soc.* **1990**, *112*, 1221. ^m Jia, G.; Lin, Z.; Lau, C. P. *Eur. J. Inorg. Chem.* **2003**, 2551. ⁿ Howdle, S. M.; Healy, M. A.; Poliakov, M. *J. Am. Chem. Soc.* **1990**, *112*, 4804. ^o Sweany, R. L. *J. Am. Chem. Soc.* **1986**, *108*, 6986. ^p Sweany, R. L. *Organometallics* **1986**, *5*, 387. ^q Childs, G. I.; Cooper, A. I.; Nolan, T. F.; Carrott, M. J.; George, M. W.; Poliakov, M. *J. Am. Chem. Soc.* **2001**, *123*, 6857. ^r Grinval'd, I. I.; Lokshin, B. V.; Rudnevskii, N. K.; Mar'in, V. P. *Dokl. Acad. Nauk SSSR* **1988**, *298*, 1142. ^s Wang, X.; Andrews, L. *J. Phys. Chem. A* **2005**, *109*, 9021. ^t Andrea, R. R.; Vuorman, M. A.; Stufkens, D. J.; Oskam, A. *Recl. Trav. Chim. Pays Bas* **1986**, *105*, 372. ^u Ishikawa, Y.; Weersink, R. A.; Hackett, P. A.; Rayner, D. M. *Chem. Phys. Lett.* **1987**, *142*, 271. ^v Ishikawa, Y.; Hackett, P. A.; Rayner, D. M. *J. Phys. Chem.* **1989**, *93*, 652. ^w Zheng, Y.; Wang, W.; Lin, J.; She, Y.; Fu, K.-J. *J. Phys. Chem.* **1992**, *96*, 9821. ^x Johnson, F. P. A.; George, M. W.; Bagratashvili, V. N.; Vereshchagina, L. N.; Poliakov, M. *Mendeleev Commun.* **1991**, 26. ^y Sweany, R. L.; Watzke, D. *Organometallics* **1997**, *16*, 1037. ^z Gadd, G. E.; Upmacis, R. K.; Poliakov, M.; Turner, J. J. *J. Am. Chem. Soc.* **1986**, *108*, 2547. ^{aa} Sweany, R. L.; Russell, F. N. *Organometallics* **1988**, *7*, 719. ^{ab} Bloyce, P. E.; Rest, A. J.; Whitwell, I.; Graham, W. A. G.; Holmes-Smith, R. *J. Chem. Soc., Chem. Commun.* **1988**, 846. ^{ac} Wang, X.; Andrews, L. *J. Phys. Chem. A* **2000**, *104*, 9892. ^{ad} Kresse, G. *Phys. Rev. B* **2000**, *62*, 8295. ^{ae} Schmidt, P. K.; Christman, K.; Kresse, G.; Hafner, J.; Lischka, M.; Gross, A. *Phys. Rev. Lett.* **2001**, *87*, 096103. ^{af} Plitt, H. S.; Bar, M. R.; Ahlrichs, R.; Schnöckel, H. *Angew. Chem., Int. Ed. Engl.* **1991**, *30*, 832. ^{ag} Solans-Monfort, X.; Branchadell, V.; Sodupe, M.; Zicovich-Wilson, C. M.; Gribov, E.; Spoto, G.; Busco, C.; Ugliengo, P. *J. Phys. Chem. B* **2004**, *108*, 8278. ^{ah} Manard, M. J.; Bushnell, J.; Bernstein, S. L.; Bowers, M. T. *J. Phys. Chem. A* **2002**, *106*, 10027. ^{ai} Balabanov, N. B.; Boggs, J. E. *J. Phys. Chem. A* **2001**, *105*, 5906. ^{aj} Andrews, L.; Wang, X. *J. Am. Chem. Soc.* **2003**, *125*, 11751. ^{ak} Bayse, C. A.; Hall, M. B. *J. Am. Chem. Soc.* **1999**, *121*, 1348. ^{al} Bayse, C. A. *J. Phys. Chem. A* **2001**, *105*, 5902. ^{am} Wang, X.; Andrews, L. *J. Am. Chem. Soc.* **2001**, *123*, 12899. ^{an} Wang, X.; Andrews, L. *J. Phys. Chem. A* **2002**, *106*, 3744. ^{ao} Wilson, S. P.; Andrews, L. *J. Phys. Chem. A* **2000**, *104*, 1640.

or liquid rare gas media have continued to be a subdiscipline that has gone hand-in-hand with studies of stable complexes, as shown in reviews by Sweany¹³² and Poliakoff.¹³³ In most cases, the preparations involve photochemical displacement of CO either in a rare gas matrix or in liquid Xe.



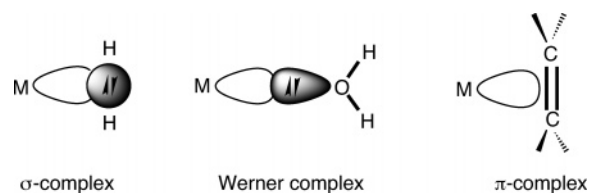
The most intensely studied species are the group 6 pentacarbonyls, $M(CO)_5(H_2)$, which have been observed in rare gas matrices, in liquid Xe solutions at $-70\text{ }^\circ\text{C}$ (a very useful medium), in alkane solvents, and even in the gas phase. As shown in Table 2, these and other complexes have relatively high H–H stretching frequencies in the $3000\text{--}3500\text{ cm}^{-1}$ range, indicative of weakly bound H_2 . As will be discussed in section 5, most stable H_2 complexes have $\nu(HH)$ lower than this. Perhaps the most novel preparation is photolysis of the hexacarbonyls impregnated in polyethylene (PE) disks under H_2 or N_2 pressures to give $M(CO)_{6-n}(L)_n$, where $n = 1\text{--}2$ for $L = H_2$ and $1\text{--}4$ for $L = N_2$.¹³⁴ Reactivity follows the order $Mo > Cr > W$, and H_2 can displace coordinated N_2 in the PE systems. In all media, vibrational spectroscopy provides evidence for H_2 rather than dihydride binding, and the H–H, H–D, and D–D stretching modes are often observed because of the clear spectroscopic window in rare gas media.

In nearly all cases, these complexes decompose rapidly and irreversibly at or near room temperature because of the weak H_2 binding on such CO-rich metals, where less backdonation is present. Their instability is exacerbated because the 16e product of H_2 dissociation is extremely reactive, since it is not stabilized by internal agostic C–H interactions or solvent binding (hydrocarbon solvents are even more weakly bound than H_2). The rate of dissociation of H_2 from $Cr(CO)_5(H_2)$ in hexane at $25\text{ }^\circ\text{C}$ is actually slower than that for many stable species. Thus, this complex and others like it might otherwise be stable under H_2 . One such complex initially presumed to be unstable, $CpMn(H_2)(CO)_2$, has in fact been isolated as a relatively stable solid from supercritical CO_2 ($scCO_2$) at $25\text{ }^\circ\text{C}$ in a flow reactor by photolysis of $CpMn(CO)_3$ in the presence of H_2 and rapid expansion of the $scCO_2$.¹³⁵ $CpMn(H_2)(CO)_2$ is one of the simplest stable H_2 complexes and has by far the lowest molecular weight (178) and highest percentage of H_2 by weight (1.1%) of an isolable transition metal H_2 complex, an important factor in materials for hydrogen storage. Analogues with Cp^* and N_2 , C_2H_4 , and $\eta^2\text{-SiHEt}_3$ ligands have also been prepared, and interchange of these labile ligands can be promoted.¹³⁵

2.2.2. Binding of H_2 to Bare Metal Atoms, Ions, and Surfaces

H_2 has also been found to molecularly bind to metal surfaces such as Ni(510), metal atoms or cations, and small metal atom clusters (e.g. $Cu_2(H_2)_2$, $Cu_2(H_2)_3$, $Cu_3(H_2)$, and $Fe_x(H_2)$ ($x = 3$ or 4) at low temperature (Table 2). Monometallic species such as $Pd(H_2)$ were first studied by Ozin (see section 3.1) and then later by Andrews⁵³ for many metals, including gold. The evidence again is entirely spectroscopic, primarily vibrational and mass spectroscopy. H_2 is believed to be bound in η^2 -fashion on the stepped edges of the Ni(510) surface, which are coordinatively unsaturated. Electron energy loss spectroscopy (EELS) at 100 K shows several bands comparable to those for organometallic H_2

Scheme 1



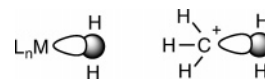
complexes. No such chemisorption is observed on the flat Ni(100) surface, which lacks the residual unfilled d states at the step sites that bind the H_2 . Undoubtedly, side-on molecular H_2 coordination is the first step in the dissociation of H_2 on metal surfaces to form hydrides and is followed by rapid splitting of H–H analogous to OA in homogeneous solution activation.

Diatomic and triatomic Cu and Pd clusters formed by vaporization react with up to three H_2 to form complexes in argon matrices at $7\text{--}15\text{ K}$. Analogous reaction of H_2 with iron clusters forms only Fe_3 or Fe_4 hydrides (Fe_2 is unreactive). Main group species such as alkali halides, boron hydrides, and Lewis bases interact very weakly with H_2 at low temperature (ν_{HH} is perturbed only slightly, see section 5 below).

3. Structure and Bonding of H_2 Complexes

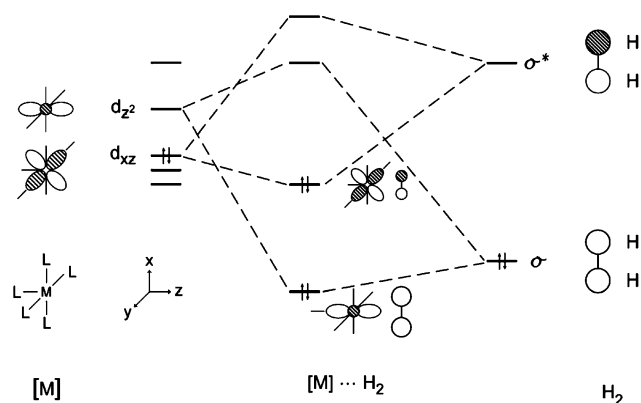
3.1. Theoretical Analysis of Nonclassical Bonding of H_2

Knowledge of the structure and bonding aspects of dihydrogen complexes is critical in understanding their properties, reactions, and dynamics. Several review articles and book chapters focus at least in part on the theoretical aspects of H–H bond coordination and activation,^{5,6,24,48a,136–138} including five in a special volume of *Chemical Reviews* devoted to computational transition metal chemistry.^{25,139–141} The nonclassical 3-center interaction of H_2 with the metal perfectly complements classical Werner-type compounds where a ligand donates electron density through its *non-bonding* electron pair(s) and π -complexes such as olefin complexes in which electrons are donated from bonding π -electrons (Scheme 1). It is remarkable that the *bonding* electron pair in H_2 can further interact with a metal center almost as strongly as a nonbonding pair. The resulting side-on (η^2) bonding in $M\text{--}\eta^2\text{-H}_2$ and other σ -complexes (and bridging hydrides/alkyls⁷) is *nonclassical*, by analogy to the 3c-2e bonding in carbocations and boranes. The M center may be considered to be a “superelectrophile” isolobal with H^+ and CH_3^+ , mimicking carbocation chemistry; that is, a σ complex such as $M^+\text{--}CH_4$ is equivalent to CH_5^+ , which in turn is now viewed as a highly dynamic H_2 complex of CH_3^+ .¹⁴³ H_2 is thus a weak Lewis base that can bind to strong



electrophiles, but transition metals are unique in stabilizing H_2 and other σ -bond complexes by *backdonation* (BD) of electrons from a filled metal d orbital to the antibonding orbital of H_2 (σ^*), a critical interaction unavailable to main group atoms (Schemes 2 and 3).⁵ Although it may seem paradoxical that an antibonding orbital such as H_2 (σ^*) can form a chemical bond, this orbital is only antibonding with respect to the H atoms and can still be bonding with respect

Scheme 2

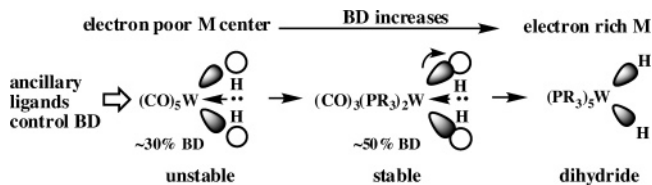


to M and H. Backdonation is a synergistic effect and can relieve the metal center of some of its excess electron density, which in turn can stabilize binding of π acceptor ligands such as CO, olefins, and even H₂. The backbonding interaction was found to present computationally by Hay¹⁴⁴ in our original tungsten–H₂ complex and is analogous⁵ to that in the Dewar–Chatt–Duncanson model^{145,146} for π -complexes, e.g., M-ethylene. Seminal theoretical and experimental studies of Pd(H₂) laid the groundwork for understanding the side-on bonding of H₂, including the presence of BD.^{147–152}

The electronic features of bonding in a metal complex truly are complex. Pauling's electroneutrality principle is important here and states that molecules arrange themselves so that their net charges fall within fairly narrow limits, about +1 to –1 overall, usually less.¹⁵³ Nonmetals such as C, N, or O prefer a charge closer to –1 while metals tend to be closer to +1. An isolated Co³⁺ ion is not an electroneutral species, since it has excessively high positive charge. It will tend to seek to form compounds with good donor ligands such as O^{2–} to form an oxide Cr₂O₃ or, in the case of coordination complexes discussed here, with NH₃ to form ammine complexes. On the other hand, an isolated M(0) atom is relatively too negatively charged (“electron-rich”), so it will prefer to attract and bind to net electron-withdrawing ligands such as CO. Complexes containing only CO ligands such as W(CO)₆ are known and now actually become electron-poor, relatively speaking. Electron balance is important in coordination complexes, and in formation of a ligand field around a metal, electrons tend to redistribute as evenly as possible over all the M–L bonds. Electron-rich complexes are better backbonders, and as we go from left to right in the transition series or down a group to third row metals, backdonation ability increases.

3.2. M → H₂ Backdonation and Influence of CO Ligands on Activation of H₂

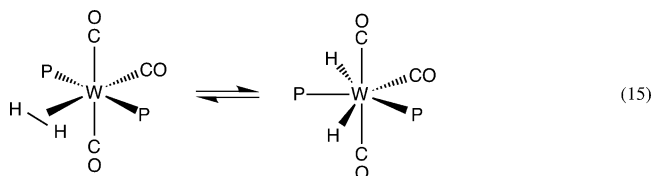
Backdonation of electrons from M to H₂ is crucial not only in stabilizing σ bonding but also in activating H–H toward homolytic cleavage to a dihydride. If BD becomes too strong, e.g., by increasing the electron-donor strength of coligands on M, the σ bond cleaves to form a dihydride because of overpopulation of H₂ σ^* . Replacing electron-withdrawing CO ligands by strongly donating phosphines ruptures the H–H bond in the tungsten system (Scheme 3). More quantitative measures of BD are provided by charge decomposition analysis (CDA) and extended transition state (ETS) analysis.^{154–160} Frenking's CDA calculations break down the bonding into donation and backdonation terms to compare binding of H₂ to that of conventional ligands.^{155–157}

Scheme 3. Backdonation (BD) Is Critical to the Stability of H₂ Complexes and H–H Cleavage

For example, CO is found to be both a good σ donor and a strong π acceptor, consistent with its ability to bind to most metal fragments. Cyanide is a powerful donor but a weak acceptor while N₂ is the opposite: a very poor donor and moderate acceptor. By comparison, H₂ is a slightly better acceptor than N₂ but, unlike N₂, H₂ is a good donor. This is beautifully corroborated experimentally by small molecule interactions with the strongly electrophilic complex [Mn(CO)₃(PCy₃)₂]⁺, which binds H₂ reversibly but not N₂, even at low temperature.⁹⁹ This binding difference may be important in hydrogenases where atmospheric dinitrogen could potentially bind and inhibit H₂ activation at the enzyme's dimetallic core. For W(CO)₅(H₂), donation from H₂ (0.349 e) is greater than BD (0.129 e), as expected for this related electron-poor system.

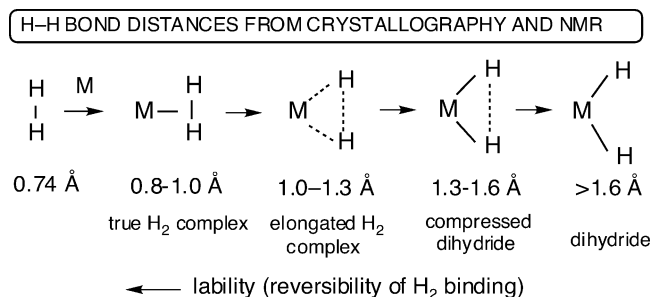
For very electrophilic centers, *loss in BD is almost completely offset by increased electron donation from H₂ to the electron-poor center*. The M–H₂ energy for electron-poor Mo(CO)₅(H₂) is surprisingly similar to that for the more electron-rich, isolable, phosphine complexes. *H₂ is the perfect ligand because it is effectively amphoteric like CO and is perhaps the most adaptable “weak” ligand*, reacting with virtually every unsaturated M fragment. As pointed out by Hoffmann,¹⁶¹ the reason CO is an excellent, ubiquitous ligand is the balance between its good donor/acceptor capabilities and its innate stability. The H₂ ligand offers the same advantages, albeit on a lesser energy scale. These and other electronic factors are important in understanding both activation of H₂ in metalloenzymes and reversible binding of H₂ for purposes of hydrogen storage that will be discussed below.

Because of the above electronic considerations, particularly BD, there is a fine line between H₂ and dihydride coordination, and in some cases, *equilibria* exist between the two forms in solution for W(CO)₃(PR₃)₂(H₂) (R = *i*-Pr; K = 0.25) (eq 15).^{2,3,6} Our seminal studies thus clearly demonstrated

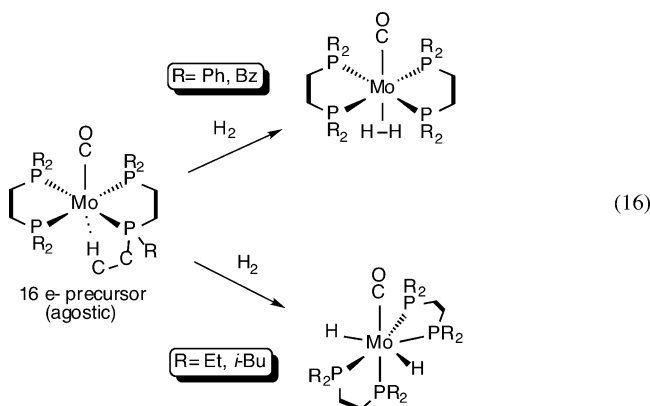


that side-on coordination of H₂ is the first step in H–H cleavage to dihydride. Equally important is that even though a complex may ostensibly be observed to contain only hydride ligands, a low-energy pathway to a coordinated H₂ ligand may exist (e.g., via the reverse of eq 15) that can result in dissociative loss of H₂ as in eq 1. Both processes can be completely reversible, providing the complex is stabilized in the absence of H₂ by either steric protection and/or agostic interaction (eq 1). Although the electronic factors for oxidative addition of H₂ in eq 15 were well-established computationally, the role of steric factors was not. The phosphines are bulky (R = cyclohexyl or isopropyl) and at first were believed to inhibit H₂ splitting to form a 7-coordinate complex.¹⁶² This later was shown to be true to

Scheme 4

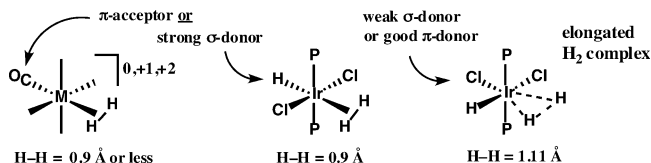


some extent: for less bulky R = Me, the equilibrium lies completely to the right, i.e., the complex is a *dihydride*,¹⁶³ and increasing phosphine size in [Cp*OsH₂(H₂)(PR₃)]⁺ led to elongation of d_{HH} in the H₂ ligand.⁷⁰ Another dramatic demonstration of the fine effects of changing electronics and sterics is H₂ addition to the agostic complex Mo(CO)(R₂-PC₂H₄PR₂)₂, whereby merely changing R controlled whether a H₂ or dihydride complex was stable (eq 16).⁸⁸ The more



electron-donating alkyl diphosphines such as depe (R = Et) lead to increased BD, ultimately favoring H-H rupture to form a dihydride. It would appear that electronic rather than steric factors are more crucial in stabilizing H₂ versus dihydride coordination, since the phosphines with R = *i*-Bu and phenyl (dppe) are similar in size. Changing M in Mo(CO)(dppe)₂ to W also leads to dihydride formation¹⁶⁴ because W is a better backbonder than Mo (third-row metals have more diffuse d orbitals).

Another indication that electronic effects predominate in stabilizing molecular H₂ versus dihydride binding is that H₂ binding was eventually found in complexes containing only very small coligands such NH₃ (section 2.1.5),^{60,121-126} that is, bulky phosphine ligands are not needed to sterically favor 6-coordinate H₂ complexes over 7-coordinate dihydrides. Second, the H-H distances were found to vary greatly completely independent of ligand size and in some of these complexes were well over 1 Å. Both of these observations represented further paradigm shifts. This led to extensive efforts by many researchers to vary the metal, ancillary ligands, and other factors to study the stretching of the H-H bond. Within the large regime of hundreds of L_nM-H₂ complexes, it was possible to map out the entire reaction coordinate for the activation of H₂ on a metal as a function of the degree of backdonation. Complexes with d_{HH} varying enormously from 0.82 to 1.5 Å were found (Scheme 4). This arresting of bond rupture along its entire reaction coordinate is unprecedented in chemistry. Although the d_{HH} ranges shown are arbitrary, each category of complexes has distinct

Scheme 5. σ Complex Favored by Strong trans Ligand and Positive Charge

properties. The d_{HH} is relatively short (0.8–1.0 Å) and reversibly bound in “true” H₂ complexes best exemplified by W(CO)₃(PR₃)₂(H₂), much as in physisorbed H₂, where d_{HH} is <0.8 Å. Elongated H₂ complexes,^{30,34} where d_{HH} = 1–1.5 Å, were first clearly identified in 1991 in ReH₅(H₂)-(PR₃)₂, where neutron diffraction showed a d_{HH} of 1.357(7) Å between two hydrides.¹⁶⁵ Complexes with such very long d_{HH} over 1.3 Å are now viewed as “compressed hydrides”, with NMR features differing from those of elongated H₂ complexes; for example, J_{HD} increases with temperature for the former and decreases for the latter.^{30,69} These are relative terms, since the H-H bond is always stretched on binding, and indeed, a near *continuum* of d_{HH} exists. The activation of H₂ is very sensitive to the nature of M, L, and charge. Strongly donating L, third-row M, and neutral charge favor elongation and splitting of H-H to hydride, while first-row M, electron-withdrawing L, and positive charge shorten d_{HH} and favor molecular H₂ binding.

The ligand trans to H₂ has a powerful influence: strong π -acceptors such as CO (and also strong σ -donors such as H) greatly reduce BD and normally keep d_{HH} < 0.9 Å, as in the Mo complexes. Thus, a σ complex can be designed by placing the potential σ ligand trans to CO or another strong π acceptor (charge is not critical), or also a very strong trans donor ligand such as a hydride. Conversely, mild σ -donors such as H₂O or π -donors such as Cl trans to H₂ elongate d_{HH} (0.96–1.34 Å), as dramatically demonstrated by the isomers of IrCl₂H(H₂)(PR₃)₂ (Scheme 5).¹⁶⁶ The cis-Cl complex in solution but in the solid state is an elongated H₂ complex (d_{HH} = 1.11 Å) due to Ir-Cl...H-Ir hydrogen bonding, illustrating the hypersensitivity of d_{HH} to both intra- and intermolecular effects.¹⁶⁷ Intermolecular interactions (e.g., crystal packing forces) can substantially affect bond lengths, so solution and solid-state d_{HH} may differ. The isomer with hydride trans to H₂ shows d_{HH} to be 0.9 Å, i.e., a true H₂ complex. The reason here is that if the trans ligand is a strong σ -donor such as hydride, there is a powerful trans influence that reduces σ electron donation from H₂ to keep the orbital electron population in balance because the orbitals are shared.¹⁶⁸ This in turn weakens the M-H₂ bonding and contracts d_{HH} even though the complex as a whole is relatively electron-rich and neutral. On the other hand, a *weak* σ -donor ligand trans to H₂ elongates the H₂ as shown in the dicationic complex, [Ru(H₂)(PP)₂]²⁺ (PP = Bz₂PC₂H₄PBz₂), where an agostic aryl C-H interaction is trans to the H₂ ligand.¹⁶⁹ This has the longest d_{HH} (1.05 Å) observed for a *dicationic* Ru-H₂ complex, which would be expected to have a short d_{HH} because of the double positive charge.

The influence of cis ligands is less consequential because the orbitals are independent of each other. Exceptions to the above effects exist to make life interesting: the isomers of Cr(CO)₄(PMe₃)(H₂) have similar J_{HD} (~34 Hz, hence d_{HH} ~ 0.86 Å) whether H₂ is trans to a CO or the good donor PMe₃.²² The strongly electron-withdrawing CO ligands may affect the electronics differently here than in an electron-

rich complex such as the Ir complex above. An even more glaring exception to the principles discussed above is FeH₂(CO)₄, which was prepared in 1931 and was the first organometallic hydride complex.¹⁵³ However, because of its electron-poor nature as in the above Cr complex and in W(CO)₅(H₂) in Scheme 3, it would be expected to be an H₂ complex.⁶ Nonetheless, relatively recent experimental and computational studies confirm that the complex is a dihydride.¹⁷⁰ The nature of the electronic state of the complex plays a large role, as will be discussed below for H₂ addition to iron atoms (section 11.2). As previously emphasized, the dichotomy between H₂ and dihydride coordination is much more complex than could have been imagined.

There is little H–H bonding interaction remaining for $d_{\text{HH}} > 1.1 \text{ \AA}$,³⁴ so at what point is the bond “broken”? Theoretical analyses suggest 1.48 \AA , i.e. twice the normal length.^{64b} In certain “elongated” H₂ complexes, e.g., [OsCl(H₂)(dppe)₂]⁺, the energy barrier for stretching the H–H bond from 0.85 \AA all the way to 1.6 \AA is calculated^{34,69} to be astonishingly low (on the order of 1 kcal/mol !). The H₂ molecule is extremely delocalized: the H atoms undergo large amplitude vibrational motion along the reaction coordinate for H–H breaking (section 6). Remarkably, d_{HH} is both temperature and isotope dependent in [CpM(diphosphine)(H₂)]ⁿ⁺ (M = Ru, Ir; $n = 1, 2$).¹⁷² These phenomena illustrate the prodigious dynamic properties of coordinated H₂ (section 6), which can even exhibit quantum mechanical behavior such as rotational tunneling in inelastic neutron scattering spectroscopy (section 11.4).¹⁷³

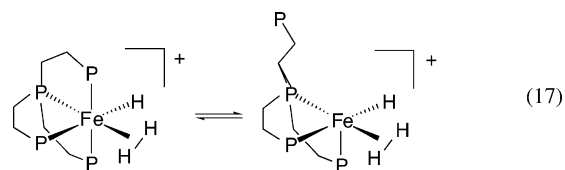
4. Properties and Spectroscopic Diagnostics for H₂ Complexes

4.1. Properties of H₂ Complexes

The properties of H₂ complexes vary tremendously, depending on the degree of activation of the H₂ ligand toward the dihydride form, i.e., the value of d_{HH} , which in turn depends on a multitude of factors as shown in section 3.2.⁶ In some instances, polyhydrides are known that adopt more than one structure in solution or that adopt different structures in solution versus the solid state, e.g., dihydrogen–dihydride and classical tetrahydride forms.¹⁷⁴ True H₂ complexes with short $d_{\text{HH}} < 0.9 \text{ \AA}$ typically have labile H₂ ligands that readily exchange with D₂ and in some cases give isotopic scrambling to HD. Atmospheric N₂ can even displace the H₂ ligand in these complexes (section 8.2.7). Most H₂ complexes are air-sensitive, reacting with oxygen to give decomposition, or very rarely, O₂ binding. The exceptions tend to be cationic species of later metals such as [IrH(H₂)(PPh₃)₂(bq)]⁺, [RuCl(H₂)(PP)₂]⁺, and [PtH(H₂)(PⁱPr₃)₂]⁺. The latter is air-stable even in solution (although it is thermally unstable above $-30 \text{ }^\circ\text{C}$).¹⁰¹ Thus, H₂ complexes are best prepared, handled, and stored under atmospheres of rare gases such as argon or helium containing some hydrogen. Occasionally, the solid complexes can be handled under N₂ or even briefly in air, though it is often necessary to use an argon-flushed glove bag ultimately filled with an argon–H₂ (or D₂) mixture, e.g., when preparing Nujol-mull IR samples of H₂ or D₂ complexes. Air-stability increases toward the later transition elements, down the group, and for complexes that are more hydridic in character (longer d_{HH}). A trace amount of water in the atmosphere or solvent is usually not a problem if excess H₂ is present, since, as will be shown in section 8.2.7, binding of H₂ competes favorably with H₂O binding (an important feature in biological systems).

Another key feature is lability of the H₂ ligand, which has two important connotations, namely *reversibility* and *ease of displacement* by other ligands. Reversibility in the strictest sense means that the H₂ can be removed *in vacuo*, by passage of an inert gas over the complex, or by heating, either in solution or solid states, to regenerate a stable precursor that re-adds H₂ for at least several cycles. Degradation or loss/gain of other ligands must not occur in the process. This property was found for the original W complex and is obviously more common for the complexes prepared from H₂ gas, which are shown in Table 1 (though not all such complexes show facile reversibility). Often the solid will have a measurable H₂ dissociation pressure ($\sim 10 \text{ Torr}$ for W(H₂)(CO)₅(PⁱPr₃)₂), necessitating a H₂-enriched atmosphere over the complex at all times. Reversible color changes, e.g., yellow to deep purple for the Kubas complexes, can occur on H₂ loss in vacuo and re-addition of H₂ and are usually rapid, even in the solid. This is often an easy (and visually impressive) test of reversibility. It is important to note that such *reversibility does not prove the existence of H₂ ligands*, although it may suggest it. Many examples of multimetallic hydrides or even complexes with –SH ligands (section 8.2.5) are known to dissociate and re-add H₂ reversibly.^{45c} Morris has tabulated the stability of a wide variety of H₂ complexes to H₂ loss in both solution and solid states.³⁰ Dissociation of H₂ to generate a vacant coordination site for substrate binding is a critical step in many catalytic hydrogenation and related processes; that is, dihydrogen complexes can function as excellent catalyst precursors.^{40,43,175}

Facile displacement of $\eta^2\text{-H}_2$ by more strongly bound ligands can occur both for the above cases and also for systems that do not bind H₂ reversibly.³⁰ For group 6 and certain other complexes, this includes coordinating solvents such as THF and acetonitrile, although some complexes are stable to H₂ loss even on heating in such solvents. In a tetraphosphine Fe complex, the H₂ is so strongly bound that when it is used as a hydrogenation catalyst for alkynes to alkenes, a free coordination site for the incoming alkyne is provided by detachment of a phosphine arm instead of H₂ loss.¹⁷⁶ However, catalysis by the Ru analogue occurs via



usual H₂ loss,¹⁷⁷ illustrating the difficulty in predicting stability, particularly for the iron group metals.^{30,178}

The photochemical stability of H₂ complexes has not been well-studied, but H₂ dissociation on exposure to visible light has been commonly observed in matrix-isolated species (section 2.2.1). The electrochemistry of H₂ complexes has also not been widely studied and is limited to cyclic voltammetric determinations. Oxidation of H₂ complexes is much more common than reduction because the majority are low valent complexes. Reversible redox systems are quite rare and include ReCl(H₂)(PMePh₂)₄¹⁷⁹ and [Os(H₂)(NH₃)₅]¹⁺,¹²¹ which show respective $E_{1/2}$ values of -0.07 and 0.58 V in organic solvents. In the latter case, oxidation is irreversible in acetone because the resulting Os(III)–H₂ complex reduces acetone to isopropanol, an unusual case where oxidation transforms a complex into a better reducing agent. Irrevers-

ible systems that primarily show anodic peaks are summarized by Jessop and Morris.³⁰ One of the few complexes to be reduced electrochemically is $[\text{FeH}(\text{H}_2)(\text{pp}_3)]^+$, which irreversibly goes to $\text{FeH}_2(\text{pp}_3)$.¹⁸⁰ $\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)$ can be electrochemically oxidized to $[\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)]^+$, whereupon the H_2 ligand becomes highly acidic (protonates weakly basic THF solvent).¹⁸¹ As will be shown, overall positive charge and electron-withdrawing coligands such as CO positioned trans to the H_2 ligand greatly increase its acidity, another critical feature in dihydrogen coordination chemistry relevant to biological activation.

4.2. Spectroscopic and Other Diagnostics for H_2 Complexes

Characterization of and evidence for dihydrogen ligands encompass several spectroscopic and crystallographic techniques, and in some cases more than one may be needed to prove the existence of H_2 binding. X-ray and neutron diffraction and NMR spectroscopy are the major techniques for determination of the structure of H_2 complexes, particularly H–H separation, by far the parameter of most interest yet the most difficult to pinpoint accurately. All stable complexes studied to date feature symmetrically side-on (η^2 -) bound H_2 as in olefin binding in order to maximize backdonation (BD) from M. However, the H–H distances (d_{HH}) span a huge range (Scheme 4), and certain polyhydride complexes studied by neutron diffraction show weak bonding interactions between two hydride ligands with $d_{\text{HH}} = 1.6$ Å in $\text{OsH}_6(\text{P}^i\text{Pr}_3)_2$,¹⁸² $1.49(4)$ Å in $[\text{OsH}_5(\text{PPhMe}_2)_3]^+$,¹⁸³ and $1.36(1)$ Å in $\text{ReH}_7(\text{P}(p\text{-tolyl})_3)_2$.¹⁶⁵ A useful empirical correlation devised by Morris enables one to predict whether or not a certain ML_n fragment will bind H_2 or form a dihydride by determining ν_{NN} for its corresponding dinitrogen complex, $\text{M}(\text{N}_2)\text{L}_n$.¹⁸⁴

The determination of d_{HH} and d_{MH} both accurately and precisely is nearly always a challenge. In certain cases, especially polyhydride complexes, there is ambiguity as to whether H_2 ligands are really present, even in neutron diffraction structures. For example, $[\text{OsH}_5(\text{PPhMe}_2)_3]^+$ was originally formulated as an H_2 complex^{113,185} and then calculationally as a pentahydride, and finally, a neutron diffraction study at 11 K showed that it is indeed closer to a pentahydride with widely varying d_{HH} (1.49, 1.75, and 1.98 Å).¹⁸³ It took eight experimental and theoretical papers from six different research groups over a 25-year period to resolve the structure and bonding in a single complex. Thus, it is not surprising that σ H_2 coordination was not found until the 1980s. Locating hydrogen bound to heavy atoms by X-ray methods is a well-known problem, and even determination of d_{HH} by neutron diffraction is complicated by rapid rotation of $\eta^2\text{-H}_2$ that shortens the observed d_{HH} .^{88c} Solid-state proton NMR can be used to accurately determine d_{HH} with good precision (± 0.01 Å).^{186,187} The first complex studied, $\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)$, showed a d_{HH} of 0.890 ± 0.006 Å.¹⁸⁷ These values are nearly always significantly longer (roughly 0.07 Å on average) than neutron values that are uncorrected for the effects of H_2 rotation. Solid-state NMR directly measures the H–H internuclear separation (rotational and other dynamics are *not* factors) and can be a better gauge than neutron diffraction.

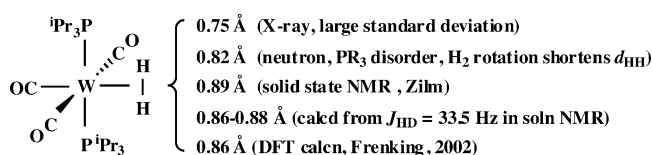
Solution ^1H NMR spectra of $\eta^2\text{-H}_2$ ligands normally give broad uncoupled signals throughout a large range of chemical shifts (2.5 to -31 ppm) that can overlap with those for classical hydrides. NMR can be used to determine d_{HH} in

solution by two different techniques involving measurement of either J_{HD} or relaxation time, T_1 . J_{HD} for the HD isotopomer of an H_2 complex is the premier diagnostic for H_2 versus hydride coordination. The signal for an HD complex becomes a 1:1:1 triplet (D has $I = 1$) with a much narrower line width and is direct proof of the existence of an H_2 ligand, since classical hydrides do not show significant J_{HD} because no residual H–D bond is present. J_{HD} for HD gas is 43 Hz, the maximum value ($d_{\text{HD}} = 0.74$ Å), and lower values represent proportionately shorter d_{HD} . J_{HD} determined in solution correlates well with d_{HH} in the solid state,⁶⁴ and both Morris^{64c} and Heinekey^{64d} developed empirical relationships, shown in eqs 18 and 19:

$$d_{\text{HH}} = 1.42 - 0.0167J_{\text{HD}} \text{ \AA} \quad [\text{Morris}] \quad (18)$$

$$d_{\text{HH}} = 1.44 - 0.0168J_{\text{HD}} \text{ \AA} \quad [\text{Heinekey}] \quad (19)$$

Input data include d_{HH} from X-ray and neutron diffraction methods plus solid-state NMR^{186,187} measurements. For $\text{W}(\text{CO})_3(\text{P}^i\text{Pr}_3)_2(\text{H}_2)$, J_{HD} is 34 Hz, giving $d_{\text{HH}} = 0.86\text{--}0.88$ Å versus 0.89 Å from solid-state NMR and $0.82(1)$ Å from neutron diffraction (uncorrected for the effects of H_2 libration). The value calculated by DFT methods is quite close

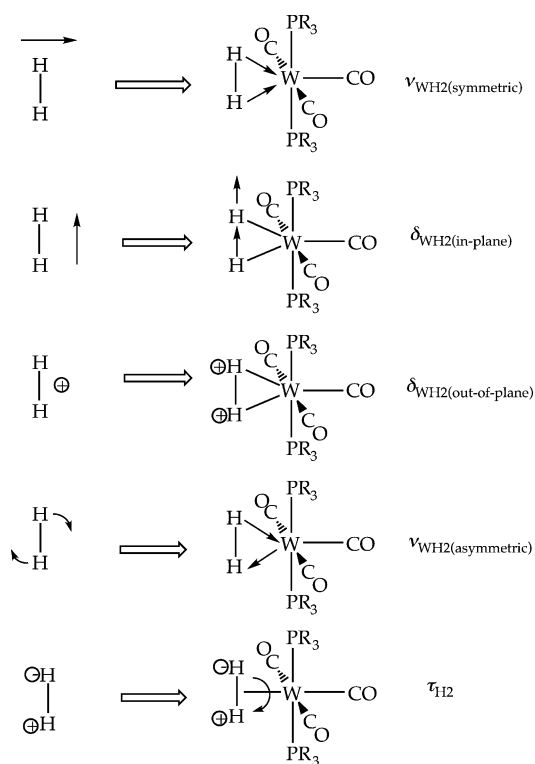


to this, 0.86 Å,¹⁸⁸ and in general, there is remarkably good agreement between experiment and theory in metal– H_2 complexes. Short T_1 values for the H_2 ligand were originally found by Crabtree to be also diagnostic of H_2 coordination (e.g., 4 ms for the W complex here versus >100 ms in hydrides).^{9,36} T_1 values are temperature dependent and go through a minimum, and the value of T_1^{min} is the important diagnostic parameter here. Because T_1 depends on d_{HH} , it is extremely sensitive to the presence of H's that are close together as in an H_2 complex. However, care must be exercised in interpretation because several factors influence T_1 values.^{120,129,189,190} Observed J_{HD} values can also exhibit temperature and even solvent dependence in certain situations, e.g., equilibria between two different structures such as a solvated dihydride of Ir(III) and an H_2 complex of Ir(I).¹⁹¹

5. Vibrational Spectroscopy of H_2 Complexes

Another valuable though underutilized characterization tool is infrared spectroscopy. The vibrational modes for $\text{M}(\eta^2\text{-H}_2)$ are distinct from those for hydrides, which have only two fundamental modes: $\nu(\text{MH})$ at $1700\text{--}2300$ cm^{-1} and a M–H bending mode at $700\text{--}900$ cm^{-1} . However, the initial routine IR spectrum of solid $\text{W}(\text{CO})_3(\text{PR}_3)_2(\text{H}_2)$ showed two bands that were outside these ranges and additionally displayed an unusual low-energy band near 460 cm^{-1} that was the first substantial clue to the novel dihydrogen structure here.^{1–4,6} When diatomic H_2 combines with a M–L fragment to form a $\eta^2\text{-H}_2$ complex, five “new” vibrational modes in addition to ν_{HH} are created which are related to the “lost” translational and rotational degrees of freedom for H_2 (Scheme 6). ν_{HH} is still present, but it is shifted to much lower frequency and becomes highly coupled with a MH_2

Scheme 6

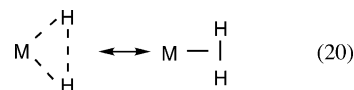


mode, $\nu_{\text{as}}(\text{MH}_2)$.⁴ Thus, six fundamental vibrational modes are expected to be formally isotope sensitive: three stretches, $\nu(\text{HH})$, $\nu_{\text{as}}(\text{MH}_2)$, $\nu_{\text{s}}(\text{MH}_2)$; two deformations, $\delta(\text{MH}_2)_{\text{in-plane}}$ and $\delta(\text{MH}_2)_{\text{out-of-plane}}$; and a torsion (H_2 rotation), $\tau(\text{H}_2)$. The bands shift hundreds of wavenumbers on isotopic substitution with D_2 or HD , which greatly facilitates their assignment. Importantly, the frequencies of the bands for the η^2 - HD complexes lie between those for the η^2 - HH and η^2 - DD isotopomers and are not a superimposition of MH_2 and MD_2 bands as seen for classical hydrides. This is another valuable diagnostic for distinguishing H_2 versus dihydride coordination, although these vibrational modes are often difficult to observe. All six bands have in fact been observed and assigned only in the first H_2 complex, $\text{W}(\text{CO})_3(\text{PR}_3)_2(\text{H}_2)$ ($\text{R} = \text{Cy}, \text{Pr}$), but this may only be due to lack of a concerted effort for other complexes. All but $\nu_{\text{s}}(\text{MH}_2)$, observed in both the IR and Raman spectra, are weak, and many of the bands tend to be obscured by other ligand modes, except for certain complexes such as $\text{Cr}(\text{CO})_5(\text{H}_2)$ that are normally stable only at low temperature.^{18–20,132–135} Table 3 lists the modes observed for selected complexes.

In the Nujol-mull IR spectrum of $\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)$, four bands, $\nu(\text{HH})$ at 2690 cm^{-1} , $\nu_{\text{as}}(\text{MH}_2)$ at 1575 cm^{-1} , $\nu_{\text{s}}(\text{MH}_2)$ at 953 cm^{-1} , and $\delta(\text{MH}_2)_{\text{in-plane}}$ at 462 cm^{-1} , can be observed to shift to lower frequency for the D_2 analogue. The band at 442 cm^{-1} in the D_2 complex is assigned to $\delta(\text{WD}_2)_{\text{out-of-plane}}$. The modes for H_2 rotation about the $\text{M}-\text{H}_2$ axis, $\tau(\text{H}_2)$, and also $\delta(\text{MH}_2)_{\text{out-of-plane}}$ near 640 cm^{-1} are observable only by inelastic neutron scattering (INS) methods, a powerful technique to locate such large amplitude vibrations involving hydrogen.^{4,6,173} These lower frequency deformations and torsions have been the least observed modes in H_2 complexes.

The frequency of most interest, ν_{HH} , varies tremendously and is often near the ν_{CH} region, where it can be obscured because most ancillary ligands such as phosphines have strong ν_{CH} bands. Use of perdeuterated phosphine ligands to eliminate such interference enabled location of ν_{HH} in

$\text{W}(\text{CO})_3[\text{P}(\text{C}_6\text{D}_{11})_3]_2(\text{H}_2)$ as a broad, weak band at 2690 cm^{-1} .^{4,6} About 30 other compounds, including surface and cluster species, exhibit ν_{HH} in a range, $2080\text{--}3200\text{ cm}^{-1}$, that is considerably lower than that for free H_2 gas (4300 cm^{-1}).⁴ As expected, there is a large dependence of ν_{HH} and MH_2 modes on both metal and ligand sets. One might anticipate a correlation of ν_{HH} with d_{HH} and the electron-backdonating ability (electron-richness) of the metal, as found for ν_{NN} and ν_{CO} in similar π -acceptor N_2 and CO ligands. However, as can be seen from Table 3, this is not the case because of the complexity of the bonding and extensive mixing of $\nu(\text{HH})$ and $\nu(\text{MH}_2)$ modes as shown by the normal coordinate analysis of $\text{W}(\text{H}_2)(\text{CO})_3(\text{PCy}_3)_2$.⁴ The latter, in fact, treats the $\text{W}-\text{H}_2$ interaction as a triangulo system, i.e., where direct BD electronic interactions exist between W and H atoms (below, left), rather than as the strictly 3-center bonding representation (below right).



Modes other than ν_{HH} have been less often observed in room-temperature stable complexes, partly because of interference from coligands or difficulty in assignment, especially if hydride ligands are also present. Low-energy modes have been identified mainly by INS methods, e.g., the torsional mode at 200 cm^{-1} for $\text{TpRhH}_2(\text{H}_2)$.¹⁹² Four modes were seen in the Raman spectrum of $[\text{CpRu}(\text{dppm})(\text{H}_2)]\text{BF}_4$, which has an elongated $\text{H}-\text{H}$ bond (1.10 \AA) and one of the lowest reported values for ν_{HH} , 2082 cm^{-1} .¹⁹³ The H_2 in elongated H_2 complexes can also be highly delocalized, and new vibrational modes must be defined (see Scheme 8 and section 6 below).^{69,172b,194} Modes for surface-bound H_2 such as on the stepped edges of a $\text{Ni}(510)$ surface can be observed, and electron energy loss spectroscopy (EELS) at 100 K shows several bands comparable to those for H_2 complexes such as $\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)$.¹⁹⁵

6. Dynamics of H_2 and Hydride Complexes

Long before the “nonclassical” dihydrogen complexes were discovered, classical polyhydride complexes had been known to be stereochemically nonrigid (fluxional) in solution, which was viewed as isolated H -atoms moving over the surface of the metal center.^{196–198} However, their association as H_2 ligands as intermediate steps is now much more attractive. For example, for hydride site exchange in polyhydrides such as ML_4H_4 ($\text{M} = \text{Mo}, \text{W}$; $\text{L} = \text{P-atom donor}$), transient intermediates with a geometry very much like $\text{MH}_2(\text{H}_2)\text{L}_4$ or *trans*- $\text{M}(\text{H}_2)_2\text{L}_4$ with elongated d_{HH} were considered possible even in 1973, long before H_2 complexes were actually discovered (Scheme 7). Since the dihydrogen ligand nearly freely rotates, that is, has a relatively low barrier to rotation ($1\text{--}10\text{ kcal/mol}$), hydride ligand rearrangement could easily take place by rotating the intermediate $\text{H1}-\text{H2}$ ligand as shown. Many new examples of hydride fluxionality and facile intramolecular and intermolecular hydrogen transfer reactions were later discovered, and the principle mechanistic aspects have been reviewed to include systems containing $\eta^2\text{-H}_2$ ligands.^{30,50,140b,199} For example, fast exchange between terminal and bridging hydrides in dinuclear rhenium complexes has been shown calculationally to be facilitated by formation of dihydrogen-containing intermediates,²⁰⁰ which may be an important feature in H_2 ases. As will be shown

Table 3. IR Frequencies (cm⁻¹) for ν_{HH} and MH_2 Modes in H_2 Complexes Compared to d_{HH} (Å)

complex	$\nu(\text{HH})$	$\nu_{\text{as}}(\text{MH}_2)$	$\nu_{\text{s}}(\text{MH}_2)$	$\delta(\text{MH}_2)$	d_{HH}	ref
CpV(CO) ₃ (H ₂)	2642					<i>a</i>
CpNb(CO) ₃ (H ₂)	2600					<i>a</i>
Cr(CO) ₅ (H ₂)	3030	1380	869, 878			19b
Cr(CO) ₃ (PCy ₃) ₂ (H ₂)		1540	950	563 ^b	0.85	82
Mo(CO) ₅ (H ₂)	3080					19b
Mo(CO) ₃ (PCy ₃) ₂ (H ₂)	~2950 ^c	~1420 ^c	885	471	0.87	2
Mo(CO)(dppe) ₂ (H ₂)	2650		875		0.88	88c
W(CO) ₅ (H ₂)	2711		919			19b
W(CO) ₃ (P ^{<i>i</i>} Pr ₃) ₂ (H ₂)	2695	1567	953	465	0.89	2, 4
W(CO) ₃ (PCy ₃) ₂ (H ₂)	2690	1575	953	462	0.89	2, 4
W(CO) ₃ (PCyp ₃) ₂ (H ₂) ^d		1565	938			<i>k</i>
Fe(CO)(NO) ₂ (H ₂)	2973	1374	~870			<i>l</i>
Co(CO) ₂ (NO)(H ₂)	{3100, 2976} ^e	1345	868			<i>l</i>
FeH ₂ (H ₂)(PEtPh ₂) ₃	2380		850	500, 405 ^f	0.82	115
RuH ₂ (H ₂) ₂ (P ^{<i>i</i>} Pr ₃) ₂	2568	1673	822 ^b		0.92	127
Tp [*] RuH(H ₂) ₂	2361				0.90	224
Tp [*] RuH(H ₂)(THT)	2250				0.89	224
[Os(NH ₃) ₅ (H ₂) ²⁺]	2231 ^b				[1.34] ^g	121
[CpRu(dpmp)(H ₂) ⁺]	2082 ^b	1358 ^b	679 ^b	486, 397 ^b	[1.10] ^h	193
Tp [*] RhH ₂ (H ₂)	2238				0.94 ⁱ	67
Pd(H ₂) (matrix)	2971	1507	950		0.85 ⁱ	152 ^{a,b}
Ni(510)-(H ₂) ^j	3205	1185	670			195

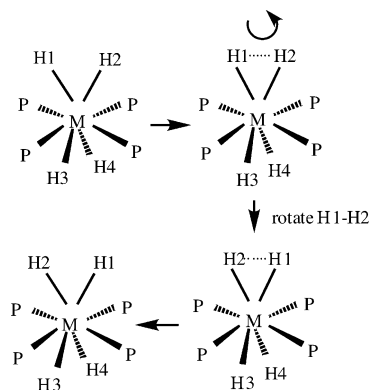
^a George, M. W.; Haward, M. T.; Hamley, P. A.; Hughes, C.; Johnson, F. P. A.; Popov, V. K.; Poliakoff, M. *J. Am. Chem. Soc.* **1993**, *115*, 2286.

^b Assignments unclear; in the case of the elongated Ru and Os complexes, these are highly mixed modes that could involve M–H modes (if present).

^c Estimated from observed D₂ isotopomer bands. ^d Cyp = cyclopentyl. ^e Split possibly by Fermi resonance. ^f Assignment unclear (data from INS).

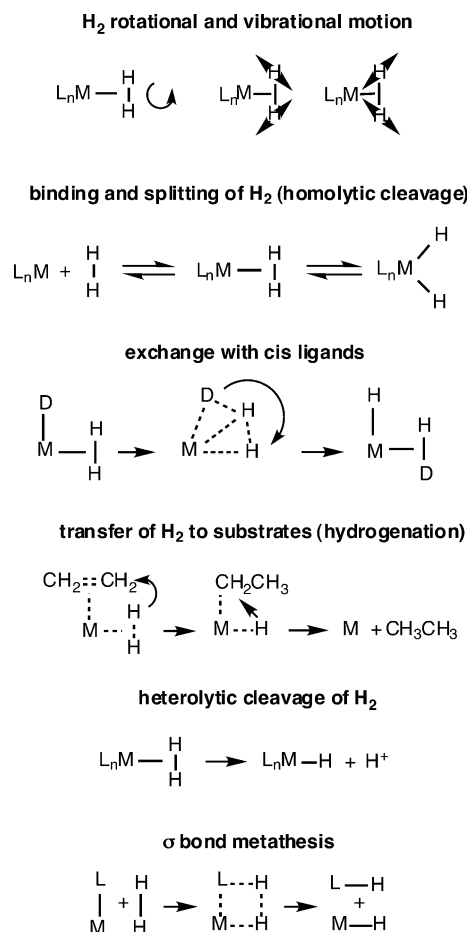
^g For [Os(ethylenediamine)₂(H₂)(acetate)]⁺ (ref 60). ^h For the Cp* analogue (ref 225a). ⁱ Calculated from inelastic neutron scattering data or DFT.

^j Data from EELS spectroscopy. ^k Khalsa, G. R. K.; Kubas, G. J.; Unkefer, C. J.; Van Der Sluys, L. S.; Kubat-Martin, K. A. *J. Am. Chem. Soc.* **1990**, *112*, 3855. ^l Gadd, G. E.; Upmacis, R. K.; Poliakoff, M.; Turner, J. J. *J. Am. Chem. Soc.* **1986**, *108*, 2547.

Scheme 7

below, remarkably facile hydrogen site exchange between cis hydride and H₂ ligands can occur even in the *solid state* at temperatures below 77 K with activation barriers as low as 1.5 kcal/mol.

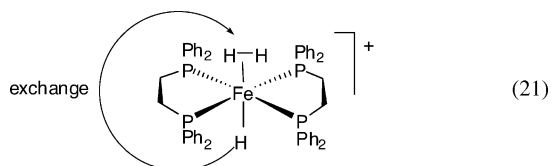
For the H₂ ligand, the structure and dynamics are much more extensive and richer than those for hydride ligands. These can include rotational/vibrational motion of η^2 -H₂, binding and splitting of H₂ (including equilibria between η^2 -H₂/dihydride tautomers), transfer of hydrogen to substrates, heterolytic cleavage of H₂, and σ bond metathesis processes (Scheme 8). Several of these processes can occur simultaneously on a metal center, and all will be discussed in more detail below. Often, these dynamics cannot be frozen out on the NMR time scale even at the lowest attainable temperatures for the system. The H₂ ligand by itself is remarkably dynamic. As discussed above, the first set of equilibria essentially represents the reaction coordinate for H–H bond cleavage/formation, which in several systems takes place in solution at room temperature. In addition to or instead of this process, virtually all complexes with H₂ ligands cis to hydride undergo extremely facile ligand exchange with very low barriers of ~5 kcal/mol or less, as

Scheme 8

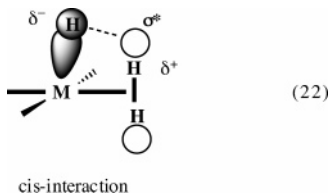
will be discussed below. Finally, in most cases, η^2 -H₂ rapidly rotates (librational motion is more accurate) even in the solid state, further delocalizing the H atom positions over virtually

the entire coordination sphere of a metal complex. One of the key diagnostics for coordination of *molecular* H₂ is in fact the observation by inelastic neutron scattering (section 11.4) of rotational transitions for η^2 -H₂, which cannot exist for classical *atomic* hydrides. Hydrogen reorientation among either chemically equivalent or inequivalent sites is extremely complex and can even involve *quantum mechanical* phenomena such as tunneling and exchange coupling between hydride ligands.²⁰¹

Facile intramolecular site exchange of H atoms between H₂ and hydride ligands is common.^{6,92,202–206} The ¹H NMR signals of the cis H₂ and hydride ligands in [Ir(H₂)H(bq)-(PPh₃)₂]⁺ coalesce at 240 K because of exchange,⁹² and even the hydride trans to H₂ in [Fe(H₂)H(dppe)₂]⁺ exchanges positions with the H atoms of η^2 -H₂.²⁰² Ab initio calculations

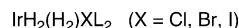
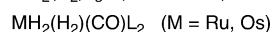
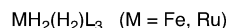
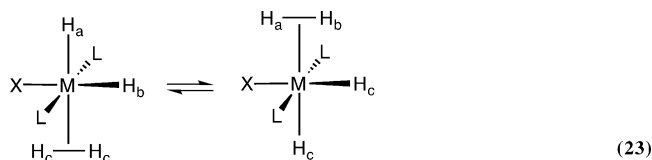


show that a variety of mechanisms are possible for the site exchange.^{203,207} Both experimentally and computationally, complexes that contain a hydride cis to a H₂ ligand often show structural and dynamic features indicative of mutual interaction.^{57,115,166,178,203,208–212} For example, the barrier to H₂ rotation (section 11.4) can be perturbed by the presence of a hydride cis to H₂. Calculations by Eisenstein show that this results from a “cis-interaction”, a hydrogen-bonding like interaction between the hydride ligand and σ^* H₂.^{115,209} This

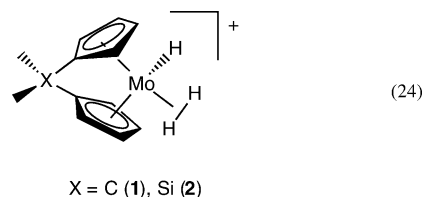


interaction is significant because of its apparent role as the nascent interaction in facile intramolecular hydrogen exchange processes, many of which can be viewed as a type of σ -bond metathesis process (Scheme 8), a term for a more general form of the above hydrogen exchange analogous to olefin metathesis.^{140b,213–216} The H₂ ligand can also interact with other atoms bound to the metal center such as B, Si, and C and undergo interconversions via σ -complex-assisted metathesis (σ -CAM), which is distinct from σ -bond metathesis and oxidative–reductive elimination mechanisms.²¹⁵ Such processes can be considered to be related to the heterolytic cleavage processes discussed below that are relevant to H₂ activation in hydrogenases.

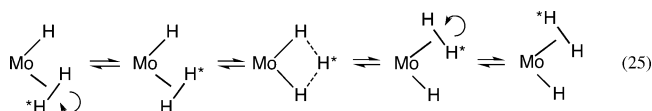
A well-studied extremely fluxional complex is IrClH₂(H₂)(Pⁱ-Pr₃)₂, where INS studies showed the lowest barrier to H₂ rotation (0.51(2) kcal/mol) ever measured for a metal complex.^{85,217} Solid-state ¹H NMR studies on a single crystal provided key initial information on the fluxional behavior.²¹⁸ A transition state with C_{2v} symmetry is attained in this and related systems by stretching the H–H bond followed by concerted migration of metal-bound hydrogens. This transient structure inverts with H_a and H_b forming a new H₂ ligand, all of which occurs in the equatorial plane of the molecule (eq 23). This is a remarkably low barrier for a solid-state process at 77 K involving considerable ligand rearrangement.



Recent studies have been carried out on bis(cyclopentadienyl)Mo type complexes, the first complexes with d² electronic configurations to have cis hydride–dihydrogen ligands. In contrast to [Cp₂MoH₃]⁺, which is a thermally stable *trihydride* complex, the *ansa*-bridged analogues [Me₂X(C₅R₄)₂MoH(H₂)]⁺ (X = C, R = H; X = Si, R = Me) have been independently determined by both Heinekey²²⁰ and Parkin²²¹ to be thermally labile *dihydrogen/hydride* complexes. Rapid dynamic processes interchange the

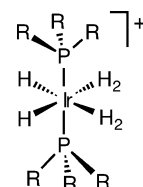


hydride and dihydrogen moieties in these complexes. The bound H₂ ligand in **1** exhibits hindered rotation with $\Delta G_{150}^\ddagger = 7.4$ kcal/mol, comparable to previously reported observations in d² Ta and Nb dihydrogen complexes.²²² However, H-atom exchange is still rapid at temperatures down to 130 K, and eq 25 depicts the dynamic process envisaged, with the central Mo–trihydrogen structure representing a transition state for atom transfer from one side of the molecule to the other. Complex **2** has an X = Si linker and methyl



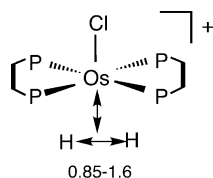
substituents on the ring carbons.²²¹ “Side-to-side” motion of the central hydrogen or deuterium atom as in eq 25 remains rapid on the NMR time scale at all temperatures studied. The barrier to rotation of the H₂ ligand is 9.0 kcal mol^{−1} at 25 °C.

There are only a handful of *bis-H₂ complexes*, which typically additionally have classical hydride ligands and present another example of the very low barriers for exchange of H₂ and hydride ligands situated cis to each other around the equatorial plane of a complex. The complex [IrH₂(H₂)₂-(PCy₃)₂]⁺ is a good example, and separate ¹H NMR resonances for the hydride and H₂ ligands could be observed on cooling of the complex to 188 K.²²³ These peaks coalesce



at 200 K, and Morris³⁰ calculates the ΔG^\ddagger at this temperature to be 8.4 kcal mol^{−1}. Chaudret’s bis-H₂ complexes, RuH₂–

Scheme 9



(H₂)₂(PR₃)₂, are also highly fluxional,³⁵ as is his Tp**Ru*H-(H₂)₂ complex, with the hydride and two η²-H₂ residing on the same side of the complex.²²⁴ Although crystallographic evidence is unavailable, NMR data is compatible with averaging of the H positions in solution, and cis-interactions between the hydrogen/hydride ligands appear likely here.

Last, the hydrogens in elongated H₂ complexes undergo rapid motion in a flat potential energy surface. Certain complexes such as [Cp**Ru*(H⋯D)(dppm)]⁺ and *trans*-[OsX-(H⋯D)(dppe)₂]⁺ (X = H, Cl) showed unusual behavior in the temperature dependence of *J*_{HD}, indicative of highly delocalized bonding.^{64c,g,172,225} In the OsCl complex (Scheme 9, *d*_{HH} = 1.22 Å, neutron diffraction), for example, *J*_{HD} unexpectedly varied from 13.6 to 14.5 Hz depending on both temperature (253–308 K) and solvent.^{64c} Several different explanations evolved, including rapid temperature-dependent interconversion of H₂–dihydride tautomers, but these were discarded in favor of rapid motion of two hydrogen atoms in a flat potential energy surface with a shallow minimum at the neutron-diffraction determined position of 1.2 Å.³⁴

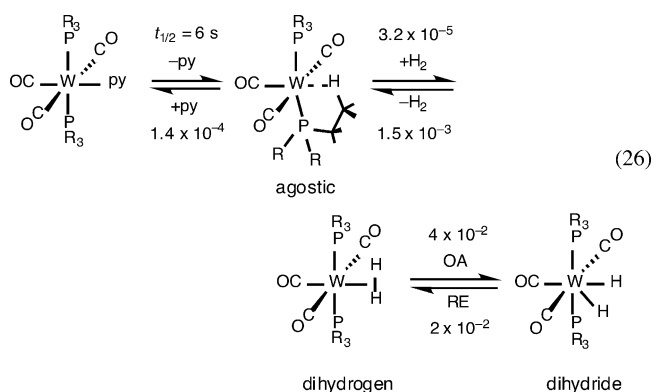
This study led to theoretical investigations that revealed the extraordinarily delocalized nature of the bonding here: *d*_{HH} can vary from 0.85 to 1.6 Å (with concomitant variation in *d*_{MH}) at a cost of only 1 kcal/mol! Subsequent NMR studies by Heinekey¹⁷² of the HD, HT, and DT isotopomers of [Cp**Ru*(H₂)(dppm)]⁺ show remarkably high isotope and temperature dependence of the bond distance (ranging from 1.037 Å for *d*_{DT} at 220 K to 1.092 Å for *d*_{HD} at 286 K) as determined by the various NMR *J* couplings. This is attributed to the extremely flat PES which defines the H–H and M–H interactions in this complex, which allows the zero-point energy differences among the various isotopomers to be directly reflected in *d*_{HH}. The striking change of *d*_{HH} with small changes in temperature is due to thermal population of vibrational excited states that are only slightly higher in energy than the ground state, an unprecedented situation in a readily isolable molecule. In certain cases, new vibrational modes needed to be defined involving a low-energy mode along the reaction coordinate for H₂ splitting and a high-energy mode orthogonal to this (Scheme 8, uppermost line).^{69,172b,194} The very strong temperature dependence of *J*_{HD} for [Ir(dmpm)Cp*H₂]₂⁺ (dmpm = bis-(dimethylphosphino)methane) was modeled simply by the Boltzmann average of the zero-point vibrationally averaged *J*_{HD} of two isomers.^{64g} For this complex and four others, the vibrational corrections to *J*_{HD} were shown to be highly significant and led to improved agreement between theory and experiment. The zero-point vibrational correction is important for all complexes. Depending on the shape of the potential energy and *J*-coupling surfaces, for some of the complexes, higher vibrationally excited states can also contribute to the vibrational corrections at temperatures above 0 K and lead to a temperature dependence.

7. Thermodynamics, Kinetics, and Isotope Effects for H₂ Binding

Solution calorimetric measurements on reactions of H₂ complexes and their precursor complexes were first carried out by Hoff and co-workers on W(CO)₃(PCy₃)₂ and W(CO)₃-(PCy₃)₂(H₂).²²⁶ Pyridine was reacted with both of these complexes to form W(CO)₃(PCy₃)₂(py). The enthalpy term for reaction with W(CO)₃(PCy₃)₂, Δ*H*^o, was −18.9 ± 0.4 kcal/mol in toluene, and that for reaction with the H₂ complex was −9.5 ± 0.5 kcal/mol under an H₂ atmosphere. The difference in enthalpies corresponds to the enthalpy of H₂ addition to W(CO)₃(PCy₃)₂, which is exothermic by 9.4 ± 0.9 kcal/mol. Note that these enthalpies are not the true binding energies because an agostic interaction is being displaced in W(CO)₃(PCy₃)₂ (see eq 1). Thus, the energy of the agostic interaction should be added to the measured enthalpies to obtain the true binding energies but could only be estimated to be about 10 kcal/mol.

Calculations indicate that 5 kcal mol^{−1} of the interaction is assigned to the net agostic interaction associated with moving from a nonagostic local minimum configuration of the PCy₃ ligands to the agostically bonded global minimum.²²⁷ Therefore, the binding energy of H₂ in W(CO)₃-(PCy₃)₂(H₂) can best be approximated to be 20 ± 7 kcal/mol. This agrees well with the values from theoretical calculations, 17–20 kcal/mol. H₂ is often a stronger ligand than one might have imagined, much like N₂, with which it is electronically similar in terms of π-acceptor strength. However, as will be shown below, H₂ is a much better σ donor than N₂ and is a *more versatile ligand than any other weak ligand (and many strong ligands) in terms of the variety of L_nM fragments to which it binds*. H₂ can coordinate or oxidatively add to both highly electrophilic and electron-rich L_nM. Thus, H₂ can be competitive with weak to moderately strong pure σ donors such as THF, water, and dichloromethane, and mass action effects are critical, as will be discussed below. Bonding strength is highly dependent on degree of H₂ activation, and much like hydrides, elongated η²-H₂ ligands cannot easily be displaced even by moderate donors such as acetonitrile.

The thermodynamic and kinetic reaction profile for H₂ addition to W(CO)₃(PR₃)₂ and equilibrium H–H cleavage has been determined for R = Cy, ⁱPr.²²⁶ The results of stop-flow kinetic studies of displacement of H₂ by pyridine (py) are given in eq 26, which shows reaction rates in terms of *t*_{1/2} (in seconds; pseudo-first-order conditions; [py] = [H₂] = 0.01 M; [W] = 5 × 10^{−4} M). In the first step of the



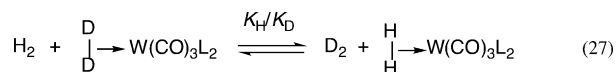
reaction sequence shown in reverse, pyridine dissociates to generate a vacant site at M on the slow time scale of seconds.

The agostic species W(CO)₃(PCy₃)₂ can then react with either pyridine or H₂ with *t*_{1/2} of 140 and 32 ms, respectively, where the rate constant *k* = 2.2 × 10⁶ M⁻¹ s⁻¹ for H₂ reaction. If the H₂ complex is formed, it may dissociate H₂ and regenerate W(CO)₃(PCy₃)₂ within 1.5 ms (*k* = 469 s⁻¹) or undergo reversible oxidative addition (OA), where *K* = ~0.25 (298 K), to form the dihydride tautomer with *t*_{1/2} = 40 ms. Under these conditions, the ratio of the rate of binding of H₂ to the rate of H₂ dissociation to the rate of OA is roughly 1200:25:1. The most surprising feature here is *the rate of dissociation of H₂ is faster than the rate of OA* by at least 1 order of magnitude. Thus, H₂ binds and dissociates many times prior to OA, which has vital importance in understanding σ bond activation processes and attendant homogeneous catalytic reactions in general. The barrier to breaking the σ bond in σ complexes is the dominant (and variable) factor in reaction rates rather than the binding of the σ ligand. The complete reaction profile for H₂ addition to W(CO)₃(PR₃)₂ has been determined. The enthalpy of activation, Δ*H*[‡], for loss of coordinated H₂ is 16.9 ± 2.2 kcal/mol, which implies a barrier of 6.9 ± 3.2 kcal/mol for the forward reaction between W(CO)₃(PCy₃)₂ and H₂, based on Δ*H*^o measured for the latter reaction, 10.1 kcal/mol.

Direct measurements of the rate constants and activation volumes for the binding of H₂, D₂, N₂, C₂H₄, and CH₃CN to the agostic complex W(CO)₃(PCy₃)₂ have recently been carried out, including both theoretical and experimental studies with time-resolved step-scan FTIR and UV–vis spectroscopy.²²⁸ The second-order rate constant for H₂ addition (*k* = 2.0 × 10⁶ M⁻¹ s⁻¹) was similar to that found by Hoff above. This rate is faster than that for N₂ addition but slower than acetonitrile binding.

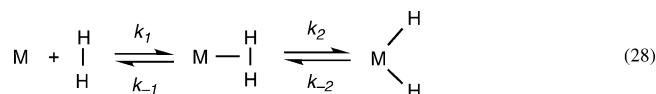
Isotope effects can be very informative in understanding chemical reactions. Both kinetic and equilibrium (or thermodynamic) effects can provide crucial information about reaction mechanisms that is unavailable from other methods. However, isotope effects often are poorly understood or may even seem paradoxical. Unlike the situation in organic chemistry, the ability of metal sites (enzymes included) to reversibly coordinate substrates prior to rate determining steps complicates the original isotope effect “rules” formulated by organic chemists. For example, the nature of equilibrium isotope effects for H₂ versus D₂ addition to metal complexes has been understood only recently. The situation can become even more complex for σ ligands that can undergo homolytic or heterolytic cleavage, either of which can also be reversible. A “normal” isotope effect occurs when the rate of reaction of an unlabeled compound is faster than that for the corresponding labeled species, i.e., *k*_H/*k*_D > 1. It is “inverse” for *k*_H/*k*_D < 1, and this terminology also applies to equilibrium isotope effects (EIEs), *K*_H/*K*_D.

The vibrational complexity of M–H₂ coordination (six modes) as shown in section 5 gives rise to an *inverse equilibrium isotope effect*; that is, D₂ binds slightly more strongly than H₂.⁴ For example, *K*_H/*K*_D = 0.70 for W(CO)₃-



(PCy₃)₂(H₂). This may be of consequence in isotopic studies of H₂ reactions, e.g., deuterium exchange reactions. Related to this is the tendency for D to concentrate in the hydride site in certain (but not all) hydride(H₂) complexes versus in η²-H₂.²⁰⁶

There is very limited data on kinetic isotope effects (KIEs) for H₂ coordination/dissociation or cleavage equilibria as shown in eq 28. For H₂ loss from the W(CO)₃(PCy₃)₂

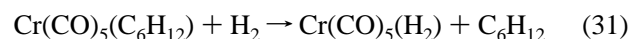


fragment, *k*₋₁ = 469 s⁻¹ for H₂ and 267 s⁻¹ for D₂, giving *k*^H₋₁/*k*^D₋₁ = 1.7.^{226d} Applying the EIE data above and the following expressions, this gives *k*^H₁/*k*^D₁ = 1.2 for H₂-binding.

$$K_{\text{H}}/K_{\text{D}} = k_{\text{H}1}^{\text{H}}/k_{-1}^{\text{H}} \times k_{-1}^{\text{D}}/k_{\text{D}1}^{\text{D}} \quad (29)$$

$$k_{\text{H}1}^{\text{H}}/k_{\text{D}1}^{\text{D}} = K_{\text{H}}/K_{\text{D}} \times k_{-1}^{\text{H}}/k_{-1}^{\text{D}} = 0.7 \times 1.7 = 1.2 \quad (30)$$

In comparison, the reaction in eq 31 occurs 1.9 times faster for H₂ than D₂ (10⁴ s⁻¹).²⁰



The subsequent rate of loss of H₂ (2.5 s⁻¹) is five times faster than that for D₂, consistent with stronger binding of D₂ over H₂.

The directly measured kinetic isotope effects for the forward and reverse reactions for the formation of W–L (L = H₂ and D₂) from W(CO)₃(PCy₃)₂, obtained by the photoinduced method of Grills et al., are 1.3 ± 0.2 and 1.4 ± 0.3, respectively, in toluene at 25 °C.²²⁸ These are slightly smaller than Hoff’s value of 1.7 but probably within the respective experimental errors.

8. Biological Activation of H₂ in Hydrogenase Enzymes

8.1. Introduction and Structure and Function of Hydrogenases

The biological activation of H₂ in hydrogenase metalloenzymes is a main focus of this article and others in this Thematic Issue. They are redox enzymes that evolved billions of years ago in micro-organisms and catalyze *completely reversible* interconversion of H₂ and protons/electrons to either utilize H₂ as an energy source or dispose of excess electrons as H₂ (eq 32) at very high rates (10⁴ turnovers/s).^{229–245}



This is a rare true equilibrium process much like that in the hydrogen electrode; for example, there is a fine dependence on H₂ pressure whether H₂ is produced or consumed by the micro-organism. From isotope exchange evidence such as the catalytic reaction shown in eq 33 (wherein the HD/H₂ ratio is pH-dependent), it is inferred that the H₂ molecule is split *heterolytically* on the metal center rather than homolytically.



Heterolysis of H₂ on transition metal complexes is a well-known process in inorganic chemistry, and catalysis and will be discussed in detail below along with other aspects of H₂ coordination on metals that form a marvelously close

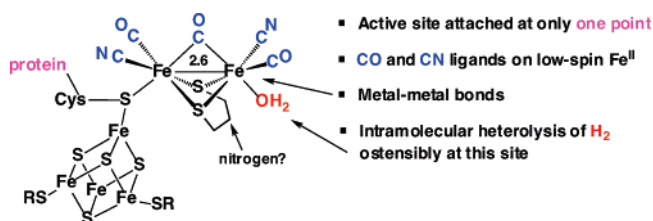
relationship to H₂ activation in Nature. Importantly, the microscopic reverse of heterolysis is formation of H₂ via, for example, protonation of a hydride ligand to form metal-coordinated H₂ that can dissociate to provide fuel for a hydrogen economy. This would be a step in the reverse of eq 32 where the electrons could come from solar photocatalytic water splitting. There is also hope that replacement of platinum in fuel cells for H₂ oxidation could be achieved using base metal catalysts (iron, nickel, etc.) modeled on the active sites in H₂ases. Since the literature on hydrogenases and modeling studies of their active sites is vast and will be addressed by other authors in this volume, only a brief introduction will be given.

Three basic types of H₂ase active sites have been identified. The most prevalent contain Ni in combination with Fe, but a select few contain only Fe and are classified as iron-only [FeFe] H₂ases. A third class was originally thought to be metal free but has recently been identified to contain iron. Although the active site is deeply buried (e.g. ~30 Å from the protein surface), channels generally exist for both proton and H₂ diffusion away from it. Amino acid residues carry protons away, and studies of xenon binding identify hydrophobic channels for H₂ gas ingress to or egress from the active site.²⁴⁶ The [NiFe] systems generally function to consume H₂ and are less active, but more resistant to oxidation, than the anaerobic [Fe] enzymes, which usually produce H₂. X-ray crystallography of the [FeFe] system indicates that there is an accessible site on Fe for H₂ binding and cleavage, but the activation site on the [NiFe] systems is not clearly established. Hydride ligands, both bridging and terminally bound, are likely to be transiently involved at some stage in the activation processes on both types of enzymes. The utilization of a bimetallic site in H₂ases is intriguing because H₂ is easily activated on a large array of mononuclear organometallic complexes without need for a second M. The M–M bonds (Ni–Fe and Fe–Fe) present in the H₂ases would then be expected to serve a useful function in Nature, perhaps as the initial site of metal protonation. Electron transfer to an attached Fe–S cubane redox-active cluster could also be facilitated. All these aspects that relate to organometallic chemistry will be covered below.

Nature has evolved extremely efficient ways to use the more abundant first-row metals such as Fe and Ni in metalloenzymes rather than the precious metals widely used as industrial catalysts. Most notably, the active sites of H₂ases feature the first biological systems with CO and cyanide ligands as intrinsic constituents, which are coordinated to dinuclear Fe–Fe bonded centers, such as shown in Scheme 10 for an iron-only H₂ase.

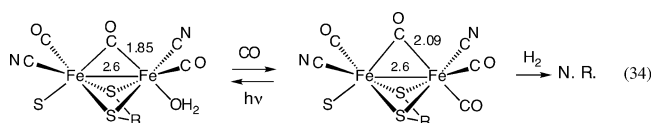
Although infrared spectroscopy provided the first evidence that CO and CN are present in H₂ases (see below), the structure of the active site determined by protein crystallography of *C. Pasteurianum* (1.8 Å resolution)²³³ by Peters in 1998 captured the attention of organometallic chemists in startling fashion. This structure and the related structure of *Desulfovibrio desulfuricans* (1.6 Å)²⁴⁷ pointed to a remarkable similarity between H₂ activation on organometallic centers and biological systems. Five CO and/or CN ligands are identified to be bound to a dinuclear iron center in *C. Pasteurianum*, including one in a bridging position. The bridging diatomic ligand is undoubtedly CO and not CN, which is not known to bridge through carbon only. Bridging CO ligands are common in organometallic chemistry and are often found in polynuclear clusters. An electron-

Scheme 10



transfer Fe₄S₄ “cubane” cluster is directly attached to Fe via a cysteine thiol bridge as shown in Scheme 10, which represents the most probable structure of the active site with one CN and CO on each Fe. An Fe–Fe bond (2.6 Å) is present in both *C. Pasteurianum* and *D. desulfuricans* that is typical of dithio-bridged organometallic Fe–Fe systems. It is important to note here that the dinuclear Fe core contains mostly *exogenous* ligands with the only attachment to the protein being through the cysteinyl sulfur bridging to the Fe₄S₄ cluster, i.e., a nearly independent organometallic complex within a protein pocket. The cyanide ligands probably engage in hydrogen bonding to the protein, which may be an important function for this biologically unprecedented moiety. Also noteworthy is the dithiolate ligand linked by a three-atom bridge, which was later speculated (and supported calculationally) to contain a nitrogen as the middle atom (as an amine group) to aid in the heterolysis of H₂.²⁴⁸ Such a precisely positioned pendant base would serve as a highly efficient proton relay to shuttle protons from the active site to exit channels in the protein, minimizing reorganization energies associated with, e.g., the approach of an external base for proton transfer. DuBois has extensively studied inorganic model systems with such pendant amines that heterolyze H₂, as will be shown below (e.g., see Scheme 13).

Mossbauer spectroscopy indicated that the Fe oxidation state is 2+ in the reduced form but Fe^{II}Fe^{III} in the oxidized form, but the states are not well established and Fe^IFe^{II} is equally probable.²⁴⁹ Although, as will be shown, CO ligands are crucial in the active site, additional CO is a known inhibitor of H₂ activation by the enzyme and irreversibly binds to the site occupied by the water molecule (eq 34), as shown crystallographically.²⁵⁰ This mimics the behavior in



organometallic systems where CO is well established to be a much stronger ligand than either H₂O or H₂. Furthermore, X-ray diffraction studies of a single crystal of the CO adduct after photolysis show dissociation of the CO and replacement by H₂O. The Fe–C distance to the μ -CO is significantly elongated when CO is bound trans to it, reflecting the strong competition for obtaining M→CO backdonation engendered between mutually trans π -accepting CO ligands. The terminal CO trans to the μ -CO is thus more labile than the other CO ligands, which are trans to electron-donating sulfur donors that enhance π -electron acceptance by CO. This leads to stronger Fe–CO bonding, again a characteristic feature in organometallic chemical bonding. The electronic influence of a ligand on the ligand trans to it is normally quite powerful (“trans influence”) and is a major tenet in all of metal coordination chemistry (see section 3.2). These and other

important inorganic chemistry principles will be discussed below in relation to the structure and function of H₂ases.

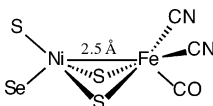
The [NiFe] H₂ases contain at least one NiFe-containing cluster considered as the probable H₂ activation site.²⁵¹ The enzyme's metal center has several states in the activation process and has received extensive theoretical analysis. The crystal structure^{251a} of the "unready" state of *Desulfovibrio gigas* shows a metal–metal bond (2.9 Å) as in the Fe–Fe H₂ase in Scheme 10 and two unlinked bridging thiolates.



The Ni center contains only thiolate ligands, and the cubane cluster is missing. Rather than a bridging CO as in Scheme 10, a bridging X (H₂O, OH[−], or O^{2−}) is present. Upon further hydrogen activation or reductive titration, the catalytically active Ni–C form binds H₂ as either H₂ or hydride ligands. CO is a competitive inhibitor of H₂ binding, forming a bound Ni–CO complex in *D. vulgaris* that was observed crystallographically,^{251e} which supports the role of Ni as the initial site of H₂ activation.

The early crystallographic data for the active site of *D. gigas* in 1995 and 1996 had revealed only the presence of three exogenous diatomic ligands bound to Fe, and the low resolution (2.54 Å) was incapable of identification as CO and CN. The first evidence for these ligands occurring as prosthetic groups in H₂ases (and indeed any biological molecule) was provided by Bagley, Albracht, and Woodruff in IR studies of *Chromatium vinosum* that showed three high-frequency IR bands at 1944, 2081, and 2093 cm^{−1}. The lower frequency band was assigned to CO, and the higher bands were suspected to be due to another multiple-bonded diatomic such as CN.^{252,253} Later, Happe et al. identified the ligands as two CN and one CO after elegant investigation of band shifts and intensities in ¹³C- and ¹⁵N-enriched samples of *C. vinosum*.²⁵⁴ When Woodruff, a colleague of mine at Los Alamos National Laboratory, queried me about the possibility that the bands could be due to CO, it made perfect sense because such strong acceptor ligands were present in W(CO)₅(PR₃)₂(H₂) and would be expected to favor reversible molecular H₂ coordination versus hydride binding. Irreversible formation of a dihydride complex would shut down a catalytic process here.

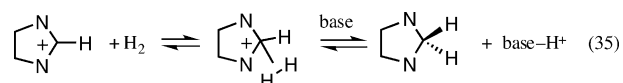
The oxidation/spin states of Ni are controversial, but almost all forms of H₂ases contain low-spin Fe^{II}, which is in the d⁶ electronic state nearly always favored for H₂ binding in organometallic systems. Biologically rare Fe^I is also possible in some of the redox states of these dinuclear M–M bonded systems. The CO and CN ligands favor both low oxidation and low spin states, which will be shown to be crucial in these systems. The crystal structure (2.15 Å resolution) of a *reduced* [NiFeSe] H₂ase from *D. Baculatum* provides insight into the actual catalytically active Ni–C state.²⁵⁵ The overall architecture of the active site is very similar to that in *D. gigas* but with Se replacing one S.



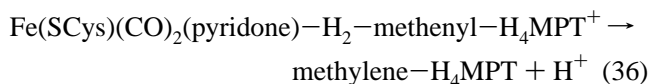
Significantly, however, the putative oxo ligand X present in the unready oxidized form is absent, and the Fe–Ni distance is 0.4 Å shorter than that in the above oxidized *D. gigas*

enzyme. The structure suggests that the closely spaced metals may now be bridged by a hydride, which cannot be seen by X-ray but is supported by theoretical calculations and ENDOR spectroscopy.^{251c} As will be discussed below, metal–metal bonds in organometallic complexes are quite basic and can readily be protonated to form a bridging hydride complex. This could be the first step in the formation of H₂ in H₂ases and may rationalize why two metal atoms are utilized when one would seem to suffice.

Remarkably, a H₂ase (Hmd) found in methanogenic archaea, *Methanobacterium Thermoautotrophicum*, was initially thought to contain no transition metals at all.²⁴³ It catalyzes the reduction of a pterin compound, methenyl–H₄MPT⁺, by H₂ and also produces a proton, as a step in methane formation from CO₂ and H₂. An electrophilic site



where positive charge is delocalized among conjugated N–C–N atoms as modeled by the formamidinium ion in eq 35 appeared to be critical to H₂ activation, as shown by ab initio studies.^{256,257} This mechanism is analogous to the reverse of that for the reversible formation of carbocations and H₂ from alkanes in superacid media, e.g., the isobutane conversion studied by Olah.²⁵⁸ However, recent X-ray absorption spectroscopy and single crystal diffraction studies revealed that a mononuclear iron site is present in the enzyme and octahedrally ligated by two *cis*-CO molecules, a cysteic sulfur atom, a pyridone nitrogen atom originating from the organic skeleton of the Hmd cofactor, an unknown ligand *trans* to a CO, and a hydrogen-bonded water *trans* to the pyridone.²⁵⁹ The mechanism for conversion of the pterin, methenyl–H₄MPT⁺, to methylene–H₄MPT, is now believed to involve a ternary complex catalytic mechanism requiring the presence of all three components (pterin, H₂, and Hmd) for enzymatic activity to occur. Thus, the iron center must be involved in the conversion, which, as for other H₂ases, undoubtedly involves heterolysis of H₂ (eq 36).



It is important to note that Hmd is phylogenetically unrelated to the other H₂ases, and the activity of this enzyme is not reversible and does not function to produce H₂. Although it now appears that a metal center is involved in the above activation of H₂, H₂ was recently reported to split by nucleophilic activation at a single carbon center in a carbene, R₂C, although, in this case, the hydrogens become irreversibly bound to the carbon to form R₂CH₂.²⁶⁰

8.2. Dihydrogen Coordination and Organometallic Chemistry Relevant to H₂ases

8.2.1. Introduction

Formation of stable iron hydrides on more nucleophilic (electron-rich) metal centers than those found in hydrogenases with CO ligands would inhibit or at least slow down function. Nature has thus been opportunistic in designing an electronically finely tuned organometallic site for elec-

trophilic H₂ activation, beating organometallic chemists to the punch 2–4 billion years ago, when microorganisms with these metalloenzymes first appeared. However, the active sites are deceptively complex: synthesis of a complete structural mimic identical to that in Scheme 10 has eluded the intense efforts of inorganic chemists over the past 8 years since the structure was reported. Organometallic models with most of the pieces have been assembled and have been valuable in understanding the structure and functions of H₂-ases. Well-established principles of inorganic, organometallic, and, more specifically, dihydrogen coordination chemistry all apply here, as will be discussed in detail in this section.

Recent developments in metalloenzyme and organometallic chemistry point to a growing link between these seemingly incongruent fields. The chemistry of organometallic compounds (standardly defined as containing one or more metal–carbon bonds¹⁵³) is almost always carried out in nonaqueous media in the absence of oxygen because organometallic compounds often rapidly decompose in the presence of air and/or water. The latter is an alien concept in most life systems, although the active sites in some H₂-ases that are present in anaerobic organisms may indeed be sensitive to oxygen but are protected in some way. Organometallic transition metal complexes typically contain abiological and often highly toxic ligands such as organophosphines and carbon monoxide that would appear to be abhorred by Nature. These notions of incompatibility were thoroughly dispelled by the relatively recent spectacular discovery of not only CO but also cyanide ligands bound to dinuclear Fe–Ni and Fe–Fe sites in H₂ases discussed above. In these often anaerobic life processes it is now abundantly clear that Nature has carried out sophisticated organometallic chemistry at the transition metal cores of hydrogenases. It is indeed humbling to consider that Nature evolved structures and methodologies eons ago that have taken the world's premier inorganic chemists over a century to independently discover and understand in their own field. This may also be said about other life sustaining biological molecules such as DNA and hemoglobin, but the organometallic features found in the dimetallo core of H₂ases had always been relegated to the domain of practiced transition metal chemists and were quite unexpected to see in Nature.

This section will then also discuss the organometallic chemistry performed by the active site of H₂ases, both from a historical perspective as well as highlighting current attempts to understand their structure and function via synthetic models and theory. Questions will be addressed such as why are normally poisonous CO and CN molecules used by H₂ases, the first example of such ligands in naturally occurring biological molecules. Does molecular binding of H₂ to iron occur (at least transiently) as in known transition metal dihydrogen complexes, and can such coordination be observed? The answers will clearly be important in the future design of biomimetic catalysts for hydrogen production. Much is known about the activation of the strong H–H bond toward cleavage on organometallic complexes. Both *homolytic* cleavage of H₂ to metal dihydrides (oxidative addition) and *heterolytic* cleavage of the H–H bond to a metal hydride plus a proton have long been known. Inorganic chemists have established key tenets here, e.g., molecular binding and heterolysis of H₂ are favored by ancillary ligands such as CO. However, it is now clear that Nature has utilized the same strategies in hydrogen activation by H₂ases far longer!

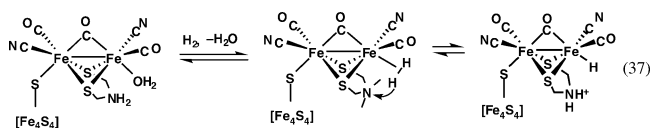
Importantly, the lessons learned from H₂ases and related biological systems may be technologically critical to our

future energy security because these utilize base metals (principally iron) to catalyze hydrogen production at extraordinarily high rates. One of the key challenges in improving chemical processing is the use of nonprecious metal catalysts in aqueous media, i.e., production of fuels, plastics, and consumer products by employing low cost abundant materials in environmentally benign “green chemistry.” Biomimetic production of hydrogen from splitting of water²⁶¹ is of particularly high interest in this regard, especially if it can be fueled by natural resources, e.g., solar energy using direct chemical coupling, as in biological photosystems.^{235,262–264} Nature solved the problem of efficient capture, transport, and storage billions of years ago, through the development of photosynthetic systems. Photosynthesis converts solar energy into high-energy chemical bonds by splitting water to form ATP, NADPH (equivalent to hydrogen), and O₂. Water oxidation is catalyzed by the oxygen-evolving complex of photosystem II. Hydrogenases from various microorganisms catalyze the production of hydrogen from protons and electrons at extraordinarily high rates using nonprecious metals, principally iron. Despite decades of effort, scientists have not yet come close to mimicking these natural systems. Two major scientific barriers persist: developing efficient (molecular) catalysts for water oxidation and H₂ production, and coupling these reactions to a photochemical energy source. Knowledge about hydrogen activation on transition metals, e.g., splitting of the H–H bond both homolytically and heterolytically, will be crucial in these pursuits, since the *microscopic reverse* is H–H bond formation and elimination as hydrogen gas, i.e., production of hydrogen fuel.

Knowledge about the key bonding concepts in organometallic chemistry also aids in understanding the structure and function of H₂ases. The Chatt–Dewar–Duncanson model originally developed for the bonding of the carbon–carbon double bond in olefins to metals is one of the cornerstones of organometallic chemistry.^{145,146,153} The olefin donates π electrons to vacant metal d orbitals and in turn receives “backdonation” (also termed backbonding) from filled metal orbitals into antibonding π^* orbitals of the multiple bond (section 3.1). Backdonation explained the relatively high metal–ligand (M–L) bond strength of ethylene and later on the even higher M–L bonding strengths of multiply bonded molecules such as CO and CN now found in the active sites of H₂ases. Although the latter are end-on bonded through carbon rather than side-on bonded as in ethylene coordination, M $\rightarrow\pi^*$ backbonding to these powerful π acceptors is very strong. Indeed, CO has been characterized to be a “universal ligand” to lower-valent metal centers,¹⁶¹ and metal carbonyl complexes such as Fe(CO)₅ and Ni(CO)₄ were among the earliest discovered organometallic compounds. As discussed in section 3.2, backdonation also greatly enhances the bonding energy of molecular H₂ to metals, where, in this case, the metal donates electrons into the H–H σ^* orbital.

Organometallic linkages were first recognized in biology in the metal–alkyl groups in cobalamins in the early 1960s, giving birth to bioorganometallic chemistry.¹⁵³ However, there have not been many examples of M–C bonds in Nature and certainly none as sophisticated as those in H₂ases. Biological activation and production of small molecules containing very strong “inert” σ -bonds such as H₂ by H₂-ases and CH₄ by methane mono-oxygenases have been known for many decades, but the structure and mechanisms had remained mysteries. Remarkably, the unexpected ability of dihydrogen (H₂) molecules to bind to metals to form stable

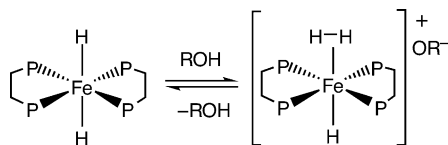
molecular hydrogen complexes (adducts analogous to hemoglobin–O₂) was not recognized until the early 1980s. As described above, the discovery of the first H₂ complex W(CO)₃(PR₃)₂(H₂) led to a new field of chemistry involving nonclassical three-center two-electron interaction of the H–H bond with a metal center with some similarity to olefin π coordination. As originally noted by Crabtree,²⁶⁵ several properties of the H₂ ligand, such as its greatly enhanced acidity compared to elemental H₂ (see below) and its ability to compete with N₂ ligands, clearly must be considered in relation to the structure and function of enzymes such as H₂ases and N₂ases. For example, these enzymes catalyze H/D exchange between H₂O and D₂ (eq 33), which an acidic H₂ ligand can easily promote via heterolytic cleavage of the coordinated H–H bond (eq 37), the key step in biological H₂ activation, as will be discussed below. It is believed that



a proton may initially transfer within the active site to either a thiolate sulfur or a basic group on the thiolate bridge in the Fe–Fe H₂ases. In order for this to occur, H₂ must ligate competitively with water as well as atmospheric N₂, and this is the case in organometallic systems, as will be shown below. The electronics at the metal center M must also be just right: H₂ is a better ligand⁹⁸ than N₂ on electrophilic M, but if M is too electrophilic, water may bind more strongly than H₂. An organometallic biological active site with a mix of strong acceptor and donor ligands such as CO and CN is advantageous here and also for heterolytic splitting of H₂.

8.2.2. Formation of H₂ Ligands by Protonation and Factors That Control H₂ Binding and Activation in H₂ases

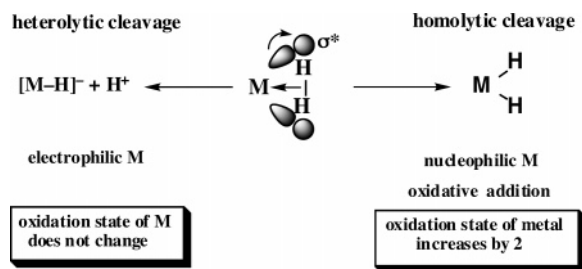
As discussed in section 2.1.4, a common method to form H₂ ligands is protonation of a metal hydride complex. Importantly, double protonation of a neutral complex can lead to formation of H₂ gas via an unstable H₂ complex that releases H₂ as in eq 12. As will be discussed below in section 8.2.11, this is a likely mechanism for formation of H₂ in H₂ases, although here this would occur at a dinuclear metal center. Iron hydride complexes are well-known to be protonated by acids to form dihydrogen complexes.²⁰² In one case, even very weakly acidic alcohols were found to be capable of reversibly protonating a hydride.^{202b} This dem-



onstrates that protons in biological systems should be quite capable of protonating the metallo site of H₂ases to form H₂ ligands that can dissociate H₂ and in a reverse process bind and split H₂.

Transition metals are unique in stabilizing H₂ complexes by M(d π) \rightarrow H₂(σ^*) backdonation (section 3.2), and the degree of backdonation is critical to the activation of H₂ toward homolytic cleavage. Increasing the electronic population of H₂(σ^*) via backdonation causes the H–H bond to elongate and eventually rupture, and examples of complexes with H–H distance (d_{HH}) varying from 0.82 to 1.6 Å have been

isolated and characterized by crystallography, NMR, and other means (Scheme 4). Several factors can stabilize molecular H₂ binding versus oxidative addition to a stable dihydride complex that would be undesirable in the function of H₂ases. These are (1) electron-withdrawing ancillary ligands such as CO, particularly trans to the σ ligand, (2) positively charged metal centers, i.e., cationic rather than neutral complexes, (3) less electron-rich first row metals such as iron (versus, e.g., Ru), and (4) orbital hybridization, i.e., octahedral coordination and a d⁶ electronic configuration. It is thus significant that the active sites of H₂ases have most all of these attributes (factor 2 may or may not be relevant or necessary here). *The nature of the ligand trans to H₂ is most often an important factor in determining whether H₂ binds molecularly and is heterolytically cleaved (versus homolytically cleaved to a dihydride or an elongated H₂ complex that is essentially a dihydride).*^{5,6,362} The *trans influence*, i.e. the electronic influence of the ligand trans to the ligand of interest (section 3.2), is crucial here, as it is in all of coordination chemistry. Complexes such as W(CO)₃(PR₃)₂(H₂) and [FeH(H₂)(dppe)₂]⁺ have either the strong acceptor CO or the high trans-effect hydride ligand positioned trans to H₂. Their H–H distances are <0.9 Å, indicative of true H₂ complexes that characteristically have labile, reversibly bound H₂, properties that are crucial to the rapid binding and loss of H₂ in enzymatic catalysis. The CO ligands, when either trans or cis to H₂, greatly reduce backbonding and stabilize molecular H₂ binding. This clearly must be their function in H₂ases, since there would seem to be no other reason for Nature to employ this toxic molecule. Importantly, d_{HH} is normally <0.9 Å (thus, H₂ is quite labile) in complexes with CO trans to H₂, regardless of ligand set or overall charge. Conversely, complexes with mild σ -donor ligands such as H₂O trans to H₂ or π -donors such as Cl have elongated H–H bonds (0.96–1.34 Å) because of increased backbonding. If the trans ligand is a strong σ -donor such as hydride, there is a powerful trans labilizing effect that reduces donation from H₂, which once again weakens M–H₂ binding and contracts d_{HH} as shown in Scheme 5. The important concept is that *the influence of the trans ligand on H₂ activation is generally greater than that of the cis ligands.* This large dependence on fragment stereochemistry can be critical in understanding how hydrogen is activated in both inorganic and biological systems. In H₂ases, the unusual CN ligand is not a strong acceptor and is an excellent electron donor that serves to preserve a low-spin state for the active site. Thus, it must be concluded that CO is the crucial ligand in controlling the electronics of the system regarding increasing the electrophilicity of the binding site to enhance both reversible molecular binding and heterolytic cleavage of H₂ (see below). Remarkably, highly electrophilic dicationic fragments such as [Fe(CO)(Ph₂PC₂H₄PPh₂)₂]²⁺ can still bind H₂ trans to CO in a stable fashion via the enhanced σ donation from H₂, offsetting the greatly reduced backdonation.²⁶⁶ This must be the case in the [Fe] H₂ases in which both irons are surrounded by CO, including in one case a bridging CO. This would disfavor OA of H₂ to give nonlabile metal hydrides and increase the acidity of iron-bound H₂ toward heterolysis. The IR value of 1945 cm⁻¹ believed to be due to Fe-bound CO in the Ni–Fe H₂ase *C. vinosum* is quite high and characteristic of a fairly electrophilic metal center. An important experimental finding is that IR spectral changes occur when the H₂ atmosphere over the fully activated enzyme is replaced by CO gas. The ν_{CO} for the CO ligand that binds to the Ni, which is the apparent site of H₂ activation, is even higher, 2060 cm⁻¹,^{251e,253} and this

Scheme 11. Dual Pathways for σ Bond Cleavage

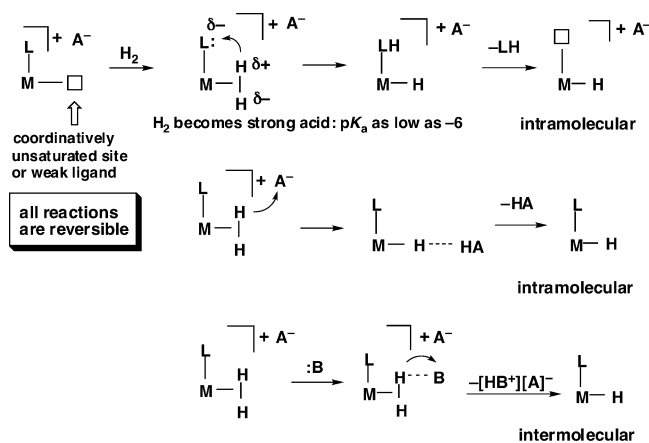
indicates a very electrophilic site. This site is possibly more electron-poor than those in organometallic carbonyl complexes such as $\text{Fe}(\text{CO})_5$ ($\nu_{\text{CO}} = 2013 \text{ cm}^{-1}$), and as will be discussed below, the acidity of H_2 bound to it could far exceed normal physiological pH values.

8.2.3. Heterolytic Cleavage and Acidity of H_2 Coordinated to Metal Complexes

The very unusual (for biology) ligand set around Fe in H_2 ases bears resemblance to many organometallic octahedral fragments that bind and activate hydrogen toward cleavage. The anionic cyanide complex, $\text{Co}(\text{CN})_5^{3-}$, was one of the first organometallic complexes found to homolytically cleave H_2 , forming the monohydride $\text{CoH}(\text{CN})_5^{3-}$, a rare example of the hydrogens transferring to two metals. Such metal centers are very electron-rich because of the strongly donating CN ligands, which favors oxidative addition of H_2 to form hydride complexes, most often dihydrides, as in Scheme 11. The latter are very common in inorganic chemistry, especially as industrial catalysts for homogeneous hydrogenation reactions.

Significantly, the oxidation state of the metal increases by two here (one in the less common case of the Co complex), and the stereochemistry around the metal changes because of the increase in the number of ligands. An H_2 ligand occupies only one coordination site in, e.g., a 6-coordinate complex but cleaves to form two hydrides, giving a 7-coordinate complex with a different arrangement of ligands where hydrides may even be distal to each other, as in eqs 15 and 16. Large oxidation state changes and drastic stereochemical rearrangements might be expected to diminish the extremely rapid rates of H_2 splitting/formation in hydrogenases. Even more importantly, in hydride complexes the hydride ligands are tightly bound and difficult to release as H_2 , clearly not an advantageous property for reversible uptake and release of hydrogen in either organometallic chemistry or biology. A second pathway involving *heterolytic cleavage*, wherein the $\text{H}-\text{H}$ bond is effectively broken into H^+ and H^- fragments, would be expected to enhance facile H_2 catalytic activation (Scheme 11).^{30,31,46,267,268} This is one of the oldest, most significant, and widespread reactions of coordinated H_2 , and importantly, here *neither the metal oxidation state nor the coordination number changes*. The earliest homogeneous (solution-phase) catalytic hydrogenation processes go back to 1938 and indeed involved heterolysis of H_2 as the key step.^{267,269} In such systems, the metal center is generally electron-poor (electrophilic), which can be accomplished by ligating π -acceptor groups such as CO to the metal and/or placing a positive charge on the complex (cationic complex). There are two pathways for heterolytic cleavage on H_2 complexes, which are most often generated either by addition of H_2 gas to unsaturated precursors (section 2.1.1) or by protonation of a $\text{M}-\text{H}$ bond (section 2.1.4). A

Scheme 12



proton can split off from the H_2 ligand and either migrate to an external Lewis base (intermolecular) or directly transfer to a coligand or anion (intramolecular) as in Scheme 12. On electron-poor cationic complexes, the H_2 ligand is highly acidic, i.e., polarized toward $\text{H}^{\delta+}-\text{H}^{\delta-}$, where the highly mobile H^+ readily transfers. Free H_2 is an extremely weak acid with a $\text{p}K_a$ estimated to be 49 in THF, but when H_2 is bound to a highly electrophilic cationic M, *the acidity of H_2 gas can be increased spectacularly, up to 55 orders of magnitude*.^{30,31,42,46,268} The $\text{p}K_a$ of H_2 can become as low as -6 , and the acidity of $\eta^2-\text{H}_2$ is as strong as that of sulfuric or triflic acid. *Intramolecular* heterolysis involves proton transfer to a cis ligand L (e.g., H or Cl) or to the counteranion (A^-) of a cationic complex. This reaction is especially facilitated if the cis ligand is Lewis basic, e.g., an amine or thiolate ligand. The basic group does not have to be attached directly to the metal but can be a component of a ligand positioned near to the metal, as will be shown in section 8.2.5. This is the process most relevant to the heterolytic cleavage of H_2 on H_2 ases. *Intermolecular* heterolysis involves protonation of an external base B to give a metal hydride (H^- fragment) and the conjugate acid of the base, HB^+ , i.e. the reverse of the protonation reaction (eq 5) used to synthesize H_2 complexes. It is critical to note that all reactions in Scheme 12 can be reversible, which is an important feature in designing molecular catalysts for hydrogen production by, for example, mimicking biological H_2 activation. As pointed out by DuBois, the heterolytic cleavage of H_2 should be at or near equilibrium to avoid high-energy intermediates.²⁷⁰ This implies the hydride (H^-) acceptor ability of the metal and the proton (H^+) acceptor ability of the base (either external or internal) must be energetically matched to provide enough energy to drive the heterolysis of H_2 , but this reaction should not be strongly exergonic.

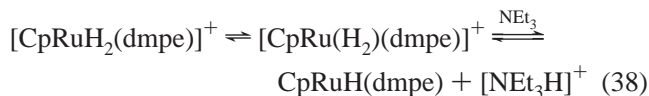
Positive charge and electron-withdrawing coligands such as CO, particularly when trans to H_2 , greatly increase the acidity. Electron deficient cationic and dicationic H_2 complexes with strong short $\text{H}-\text{H}$ bonds ($<0.9 \text{ \AA}$) and weakly bound H_2 , such as $[\text{Cp}^*\text{Re}(\text{H}_2)(\text{CO})(\text{NO})]^+$ and $[\text{Re}(\text{H}_2)(\text{CO})_4(\text{PR}_3)]^+$, are among the most acidic complexes, with $\text{p}K_a$ values determined to be as low as -2 (Table 4). Note that the value for the *neutral* Ru complex is very much higher, 36 (as measured in THF). The highly acidic complexes typically have relatively high values of J_{HD} for their η^2 -HD isotopomers, although $\text{p}K_a$ values do not correlate well with J_{HD} except within specific complex types such as $[\text{FeH}(\text{H}_2)(\text{depe})_2]^+$ versus $[\text{FeH}(\text{H}_2)(\text{dppe})_2]^+$. A good example of the effect of positive charge is $\text{W}(\text{CO})_3(\text{PCy}_3)_2-$

Table 4. Reported pK_a Values (Pseudo-aqueous Scale) and Corresponding J_{HD} of Selected H₂ Complexes, Emphasizing Highly Acidic Species

complex ^a	pK _a	J _{HD} , Hz	ref
[Cp*Re(H ₂)(CO)(NO)] ⁺	-2	27	103
[Re(H ₂)(CO) ₄ (PPh ₃)] ⁺	-2 to 1	33.9	74, 272
[FeH(H ₂)(depe) ₂] ⁺	~16	28	364
[FeH(H ₂)(dppe) ₂] ⁺	12.1	30	364
[FeH(H ₂)(dtfpe) ₂] ⁺	7.8	32	364
RuH ₂ (H ₂)(PPh ₃) ₃	36		268b ^b
[CpRu(H ₂)(dmpe)] ⁺	10.1	22.1	271
[CpRu(H ₂)(dppe)] ⁺	7.5	dihydride tautomer	c
[CpRu(H ₂)(dppe)] ⁺	7.0	24.9	c
[CpRu(H ₂)(dfpe)] ⁺	-5	29.1	276
[OsCl(H ₂)(dppe) ₂] ⁺	7.4	13.9	64c
[Os(CH ₃ CN)(H ₂)(dppe) ₂] ²⁺	-2	21.4	31
[Os(CO)(H ₂)(dppp) ₂] ²⁺	-5.7	32.0	d

^a depe = 1,2-bis(diethylphosphino)ethane; dppe = 1,2-bis(diphenylphosphino)ethane; 1,2-bis(diphenylphosphino)ethane; dfpe = (C₂F₅)₂PC₂H₄P(C₂F₅)₂; dtfpe = 1,2-bis[di-(*p*-trifluoromethylphenyl)-phosphino]ethane; dppp = 1,2-bis(diphenylphosphino)propane; dmpe = 1,2-bis(dimethylphosphino)ethane. ^b Morris, R. H. *Inorg. Chem.* **1992**, *31*, 1471. ^c Jia, G.; Morris, R. H. *J. Am. Chem. Soc.* **1991**, *113*, 875. ^d Rocchini, E.; Mezzetti, A.; Ruegger, H.; Burckhardt, U.; Gramlich, V.; Del Zotto, A.; Martinuzzi, P.; Rigo, P. *Inorg. Chem.* **1997**, *36*, 711.

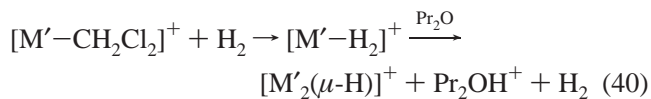
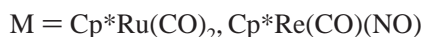
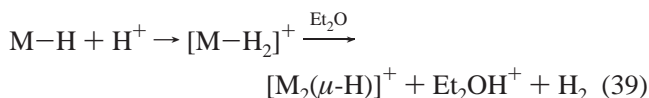
(H₂), which can be deprotonated only by strong bases such as alkoxides and KH but can be electrochemically oxidized to [W(CO)₃(PCy₃)₂(H₂)]⁺ that now is acidic enough to protonate weakly basic THF solvent.¹⁸¹ Crabtree first demonstrated heterolysis of η²-H₂ as in Scheme 12 by isotopic labeling studies to show that H₂ in [IrH(H₂)(benzoquinolate)-(PPh₃)₂]⁺ is deprotonated by LiR in preference to the hydride.⁹² A milder base, NEt₃, was shown by Chinn and Heinekey²⁷¹ to specifically deprotonate the η²-H₂ tautomer in the equilibrium mixture (84:16 ratio of η²-H₂ to dihydride form) in eq 38:



This indicated a pK_a of 17.6 in CH₃CN, and, more importantly, NMR evidence showed that the H₂ tautomer is deprotonated more rapidly than the dihydride form, which showed a *greater kinetic acidity of the H₂ ligand* (the dihydride is actually a slightly stronger acid with a pK_a of 16.8). The main reason H₂ complexes have greater kinetic acidity than classical hydrides of similar structure is that deprotonation of an H₂ complex involves *no change in coordination number*. Also, the η²-H₂ can become polarized toward H^{δ-}-H^{δ+}, and H⁺ is exceedingly mobile, especially for cationic complexes.

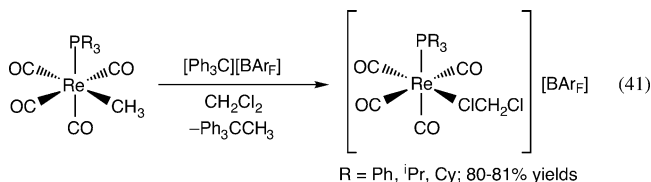
8.2.4. Intermolecular Heterolytic Cleavage of Coordinated H₂

One of the best examples of *intermolecular* heterolytic cleavage of η²-H₂ is the protonation of ethers by extremely electrophilic cationic H₂ complexes containing electron-withdrawing ligands such as CO (eqs 39 and 40).^{74,103,272}



In all cases, a hydride-bridged complex is the product even though the mononuclear hydride M-H is known in eq 39 and is used to generate the thermally unstable H₂ complex by protonation with HBF₄. A mononuclear hydride complex is not observed by NMR in eq 40, indicating a strong thermodynamic preference for the μ-H dimer. Interestingly, hydrogenase enzymes heterolytically activate H₂ and have dinuclear active sites that are capable of forming bridging hydrides by reversible protonation of M-M bonds. The pK_a of bound H₂ in eqs 39 and 40 can be estimated to be near -2 (the pK_a of Et₂OH⁺ is -2.4 in sulfuric acid²⁷³), although the irreversible formation of the μ-H product provides a driving force for deprotonation that could raise the effective pK_a of the H₂ complex a few units. A notable difference between eqs 39 and 40 is that [Re(H₂)(CO)₄(PR₃)]⁺ is *synthesized directly from reaction of H₂ with an isolable precursor*,⁷⁴ while the Cp complexes are formed by protonation of a hydride with a strong acid.¹⁰³ Only a few other examples of highly acidic η²-H₂ directly generated from H₂ gas are known.^{130,274-277}

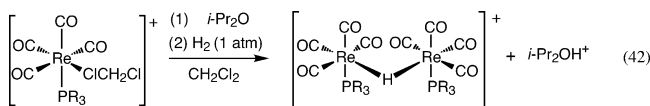
A crucial initial step in heterolysis of σ bonds is generation of a complex with either a coordinatively unsaturated site or more commonly a site occupied by a weak, easily displaceable ligand such as a solvent molecule. Dichloromethane is very convenient here because it is an excellent solvent for cationic complexes and forms isolable complexes despite the high lability of the CH₂Cl₂ ligand. A good synthetic route to CH₂Cl₂ complexes is abstraction of a methyl ligand using a trityl salt with a low coordinating anion such as BAR_f (B[3,5-C₆H₃(CF₃)₂]₄⁻). For example, treatment of [cis-Re(Me)(CO)₄(PR₃)] (R = Ph, Cy) with [Ph₃C][BAR_f] in CH₂Cl₂ solution produced [cis-Re(CO)₄(PR₃)(CH₂Cl₂)]-[BAR_f], where the CH₂Cl₂ is bound via a lone electron pair on Cl.⁷⁴ The fact that CH₂Cl₂ (as well as Et₂O) complexes



are isolable is attributed to the strong electrophilicity of the 16e [Re(CO)₄(PR₃)]⁺ fragment. The importance of a non-interacting counterion for weak ligand binding, such as dichloromethane in this and other highly electrophilic systems, is reflected by the isolation of species such as cis-Re(CO)₄(PPh₃)(F₃B) and cis-Re(CO)₄(PPh₃)(OTeF₅) where the *anion* is coordinated rather than, for example, CH₂Cl₂.²⁷⁸⁻²⁸⁰ Although dichloromethane has been traditionally thought of as a noncoordinating solvent, the isolation of stable CH₂Cl₂ complexes has been a recurring theme in recent literature,^{101,281-286} particularly for extremely electron deficient cationic metal centers with low-interacting anions such as BAR_f. Another strategy for generating unsaturated sites for H₂ addition is abstraction of a chloride ligand by silyl cations.^{64f,201} Reaction of [Cp*Ir(P-P)Cl][B(C₆F₅)₄] (P-P = diphosphine) with [Et₃Si][B(C₆F₅)₄] in methylene chloride

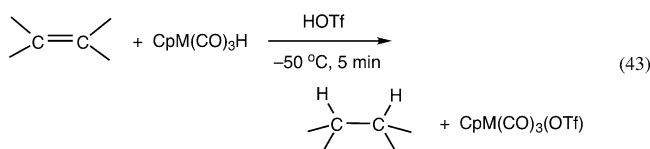
under 1 atm of hydrogen gas afforded the dicationic compressed dihydride complex $[\text{Cp}^*\text{Ir}(\text{P}-\text{P})\text{H}_2][\text{B}(\text{C}_6\text{F}_5)_4]_2$.^{64f}

Regarding displacement of the very labile CH_2Cl_2 ligand by H_2 in $[\text{Re}(\text{CO})_4(\text{PR}_3)(\text{CH}_2\text{Cl}_2)]^+$, no peaks attributable to the expected $\eta^2\text{-H}_2$ complexes were observed in ^1H NMR spectra taken at -80 to 20 °C under H_2 atmosphere in $\text{CD}_2\text{-Cl}_2$ solution.^{74,277} However, when solutions in noncoordinating $\text{C}_6\text{D}_5\text{F}$ were placed under 3 atm of H_2 , broad resonances for $\eta^2\text{-H}_2$ were observed at -4.69 ppm for $[\text{cis-Re}(\text{CO})_4(\text{PPh}_3)(\text{H}_2)][\text{BARf}]$. The addition of H_2 was completely reversible, but the H_2 complexes could not be isolated due to loss of H_2 and decomposition in $\text{C}_6\text{H}_5\text{F}$ solutions. The HD complexes were prepared, and the J_{HD} coupling constants were measured to be 33.9 and 33.8 Hz for the PPh_3 and PCy_3 complexes, respectively. The high J_{HD} observed for these complexes is consistent with those observed in other electrophilic cationic $\text{M}(\text{H}_2)$ systems and suggested a short H–H distance of ~ 0.87 Å and a bonding picture in which the metal– H_2 σ interaction is greatly enhanced relative to the backbonding interaction. Although the ^1H NMR signals for coordinated H_2 were not observed in CD_2Cl_2 solutions of $[\text{Re}(\text{CO})_4(\text{PR}_3)(\text{H}_2)]^+$, heterolytic activation of H_2 was evident in CH_2Cl_2 by protonation of free diisopropyl ether. When $^i\text{Pr}_2\text{O}$ (4–10 equiv) was added to CD_2Cl_2 solutions of the CH_2Cl_2 complexes followed by placement under H_2 atmosphere, complete conversion to the hydride-bridged dimers $\{[\text{cis-Re}(\text{CO})_4(\text{PR}_3)]_2(\mu\text{-H})\}[\text{BARf}]$ was observed.



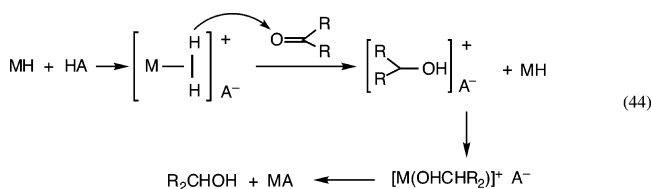
Evidently, CH_2Cl_2 and H_2 complexes existed in equilibrium in CH_2Cl_2 solution, but the exchange was too fast on the NMR time scale to observe the intermediate $[\text{Re}(\text{CO})_4(\text{PR}_3)(\text{H}_2)]^+$ complex that protonated the ether. The pK_a of the H_2 complex was estimated to be approximately 1 to -2 . Heinekey observed similar deprotonation of $[\text{Cp}^*\text{Re}(\text{CO})(\text{NO})(\text{H}_2)][\text{BF}_4]$ with Et_2O to give a hydride-bridged dimer.¹⁰³ Surprisingly, the nature of the anion was found to be important in the deprotonation of $\text{trans-}[\text{FeH}(\text{H}_2)(\text{dppe})_2]^+$ by Et_3N .²⁸⁷ The reaction rate was accelerated by BF_4^- and PF_6^- and decelerated in the presence of bulkier BPh_4^- , which hinders the approach of base via intermediate structures containing $\text{Fe-H}_2\cdots\text{N}$ and $\text{Fe-H}\cdots\text{H}\cdots\text{N}$ dihydrogen bonds (see eq 5, which shows the reverse reaction, the protonation of a hydride).

The heterolytic activation of H_2 in the above system is particularly interesting in that it may be applicable to reactions in which ionic hydrogenation of hindered substrates from a metal catalyst and H_2 is desired. In 1989 Bullock reported the first examples of ionic hydrogenation wherein a mixture of an organometallic hydride such as $\text{CpMoH}(\text{CO})_3$ and a strong acid such as HO_3SCF_3 reduces sterically hindered olefins to alkanes via protonation to carbocations followed by hydride transfer from the metal hydride (eq 43).²⁸⁸ Several other examples have since been reported,



including hydrogenation of alkynes and ketones.^{289–291} It is

likely that an acidic H_2 (or dihydride) complex is involved in the proton-transfer step of some of these reactions (eq 44). This system is significant in that it indicates that H_2

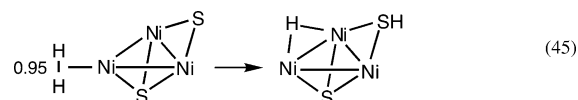


ligands can be *directly* reactive in catalysis via proton transfer and not just as an intermediate to formation of catalytically active dihydride ligands.

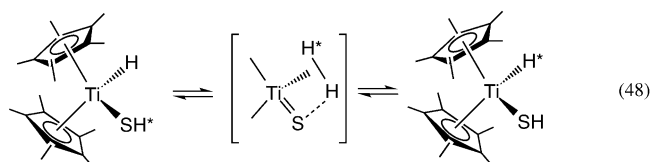
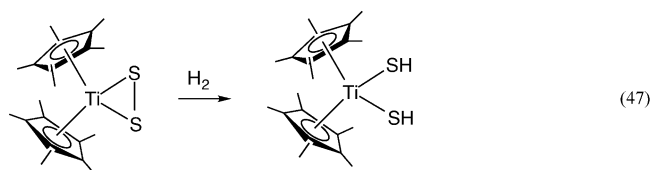
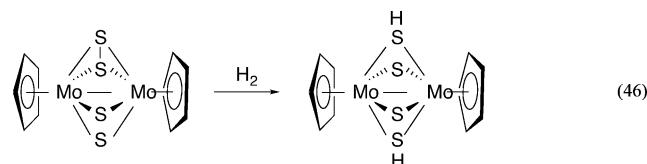
Although the primary focus of this article is on heterolysis of established dihydrogen and σ bond complexes, considerable research has been carried out on heterolytic activation of hydrogen involving classical hydride systems or unidentified transient species. Important data on the thermodynamics of H_2 splitting and the hydride donor abilities of $[\text{MH}(\text{PP})_2]^+$ ($\text{M} = \text{Ni}, \text{Pd}, \text{Pt}$; $\text{PP} =$ diphosphine) have been reported by DuBois and Curtis.^{270c,292} The dicationic complexes $[\text{M}(\text{PP})_2]^{2+}$ heterolytically cleave H_2 in equilibrium fashion in the presence of bases such as amines to give protonated amine and $[\text{MH}(\text{PP})_2]^+$. The involvement of a dihydrogen (and/or dihydride) complex could not be directly identified, illustrating the frequent problem encountered in activation of σ bonds, namely whether the mechanism involves a σ complex, i.e., $\text{M}(\eta^2\text{-H}_2)$ (or generically $\text{M}(\eta^2\text{-X-H})$), or oxidative addition to $\text{M}(\text{X})(\text{H})$.

8.2.5. Intramolecular Heterolytic Cleavage of H_2

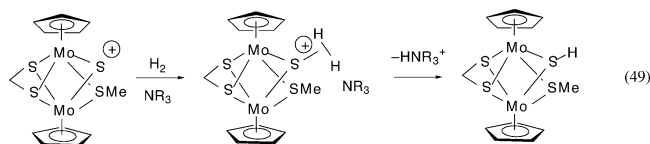
Intramolecular heterolytic cleavage of H_2 is one of the oldest reactions of H_2 and is among the first homogeneous catalytic conversions. $\eta^2\text{-H}_2$ can protonate a counteranion or a basic ancillary ligand, either at the M-L bond or at a ligand lone pair. Intramolecular heterolysis of H-H is most likely an essential step in many diverse systems ranging from industrial processes to the function of metalloenzymes such as hydrogenases. These include heterogeneous catalysis such as in the world's largest man-made chemical reaction, hydrodesulfurization (HDS) of crude oil on metal sulfides, typically MoS_2 and RuS_2 . Heterolysis of H_2 on these and other sulfides to form M-H and M-SH groups is well-known^{293,294} and has been modeled calculationally on NiS and a Ni_3S_2 cluster.^{293b,294} A transient Ni-H_2 species is calculated to be stable by ~ 16 kcal/mol and energetically capable of transferring one H to S (eq 45).^{293b} H_2 also readily



reacts with a select few organometallic sulfides to give SH complexes (eq 46) which can show exchange behavior (eq 48).^{295–298} Although the mechanism of eq 46 is unknown,^{295,298} a four-center S_2H_2 transition state can be envisioned, since there are no vacant coordination sites available on the metal. $[(\text{triphos})\text{Rh}(\mu\text{-S})_2\text{Rh}(\text{triphos})]^{2+}$ reversibly forms $[(\text{triphos})\text{Rh}(\mu\text{-SH})_2\text{Rh}(\text{triphos})]^{2+}$ under H_2 .²⁹⁷ Equation 47 represents the first example of H_2 addition to a nonbridging disulfide complex.²⁹⁶ An undetected H_2 complex may explain NMR evidence for H-atom exchange in eq 48, including the protons in dissolved H_2 gas.²⁹⁶ A

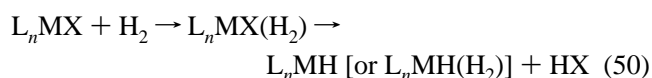


related Mo–S system shows reaction of H₂ with saturated cationic sulfide-bridged complexes in the presence of a base (NR₃), which may be explainable by direct attack of H₂ on sulfur to form a 3c2e S–H₂ interaction, followed by intermolecular heterolytic cleavage of H₂.^{295,298} Although this

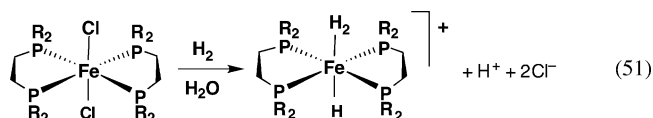


type of reaction is quite rare, it is possible that activation of H₂ could be entirely *sulfide ligand-based* in these reactions as well as in certain biological and industrial catalyst systems. Unlike the active sites in H₂ases, there is no open (or displaceable) site on the metal for H₂ coordination and heterolysis. The richness and versatility of *Mo-based* clusters in undergoing such unique reactions that can involve internal Mo–S redox processes could relate to their presence in nitrogenase enzymes and in HDS catalysts (W analogues do not display the reactivity in eqs 46 and 49).^{299,300} The Mo–SH groups formed in the above reactions can act as reducing agents toward, for example, SO₂, where hydrogenation to elemental sulfur and H₂O was found to occur.^{7,301}

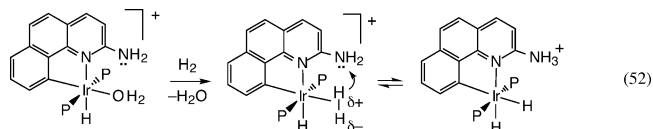
Intramolecular heterolysis of H₂ with elimination of HX (X = Cl) is commonly observed under homogeneous reaction conditions.^{44,106,302–304}



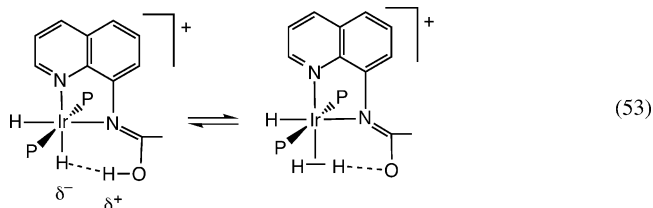
The mechanism in most cases follows that in Scheme 12 where the proton transfers to a cis ligand X. This reaction is useful for preparative and catalytic chemistry; for example, a metal halide (including bridging X) can be converted to a metal hydride in the presence of base or under phase-transfer or high-pressure conditions. In some cases, a dihydrogen-(hydride) complex can be directly prepared via heterolytic cleavage of H₂ and subsequent displacement of chloride by H₂.^{106,303} This can even be done in aqueous solution for water-soluble phosphines (R = methoxypropyl).³⁰³ In the Ru analogue, the H₂ ligand is found to participate in intermolecular hydrogen bonding in solution.^{303b}



Another important type of heterolytic cleavage of H₂ highly relevant to that presumed to take place at the active site of H₂ases is shown in eq 52.^{305,306} The conversion is

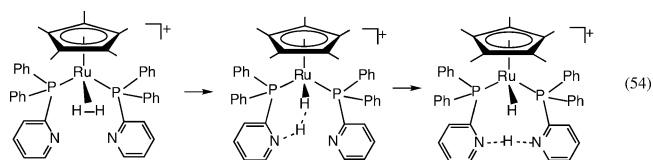


completely reversible by removing the H₂ gas from solution and is remarkably sensitive to phosphine size and ion-pairing effects. A similar proton transfer occurs to a Ru-bound NH₂ (amido) ligand on heterolysis of H₂ on (PCP)Ru(CO)(NH₂) (PCP = 2,6-(CH₂PBu₂)₂C₆H₃).³⁰⁷ An ammonia ligand is formed which then dissociates to give (PCP)RuH(CO). Such “ligand-assisted heterolysis” of the type M(amide) + H₂ → MH(amine) had earlier been found by Fryzuk at about the time M–H₂ complexes were first discovered, and thus, intermediate H₂ coordination was not initially speculated to be a part of the mechanism of such processes.³⁰⁸ These reactions are possibly facilitated by intramolecular hydrogen bonding interactions, e.g., eq 53, where the OH and IrH hydrogens scramble via rotation of the H₂ ligand. The

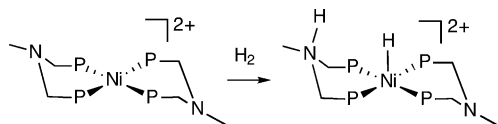


H···H interactions (1.75–1.9 Å) here and related systems are referred to as “proton-hydride bonding” by Morris^{127,309} and “dihydrogen bonding” by Crabtree,^{310–312} who, along with others,^{313–315} have studied or reviewed such *unconventional hydrogen bonds* that include M–H···H–M′, M–H···H–X, and X–H···σ interactions in general (X = C, N, P, O, etc). Remarkably, the H₂ ligand in water-soluble Ru diphosphine dihydrogen complexes has recently been found to hydrogen bond to bulk solvent.^{303b} These complexes can represent intermediates in the heterolytic splitting of H₂ and illustrate both the basicity of the M–H bond and the acidity of η²-H₂. The interactions can be comparable in strength to classical X–H···(lone pair) hydrogen bonds (3–7 kcal/mol). The discovery of the dihydrogen bond and new findings in this area have given significant rebirth of interest in hydrogen bonding in transition metal chemistry^{316,317} that can parallel well-known hydrogen-bonding effects in biological systems.

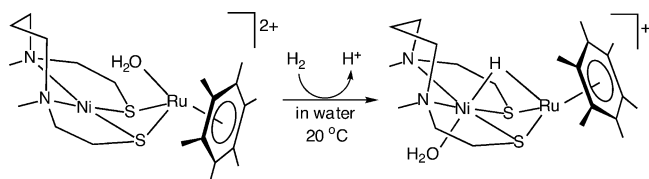
Related H₂ heterolysis also occurs via intramolecular proton transfer between nitrogens on Ru complexes containing phosphinopyridine ligands (eq 54).³¹⁸ Reversible heterolysis of H₂ occurs via dihydrogen bonding involving a protonated pyridine group similar to that in eq 53. An



Scheme 13



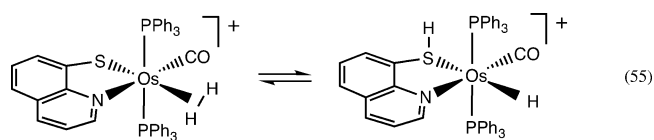
Scheme 14



additional intramolecular proton-transfer process is proposed to occur between the nitrogens of the pyridine rings on adjacent phosphine ligands; that is, DFT calculations show that a proton can be “handed off” from one ring to another via a symmetrical proton-bridged transition state. The complex catalyzes deuterium exchange with methanol- d_4 , where initially 50% of the Ru-bound H_2 is labeled after 7 min. DuBois found that a Ni(II) complex heterolyzes H_2 to form a Ni hydride bond and a protonated pendant amine.^{270c} Although an intermediate H_2 complex was not observed, DFT calculations on a closely related model complex indicated one exists with an energy 2.1 kcal/mol above that of the reactants.^{270a}

A heterolysis of H_2 on a Ni–Ru complex to form a bridging hydride complex directly relevant to the function of NiFe H_2 ases was recently reported by Ogo and co-workers (Scheme 14).³¹⁹ This system is unique in that it undergoes the crucial reaction with H_2 under ambient conditions in water to give the Ni(μ -H)Ru structure analogous to that proposed to occur in the active form of the enzyme, albeit with Ru instead of Fe and different coligands (see Figure 4).

The first direct observation of equilibrium between an acidic H_2 complex and a corresponding hydride complex with a protonated ancillary ligand is shown in eq 55.³²⁰ Here a



proton migrates from H_2 to a thiolate ligand trans to it, possibly via base-assisted heterolysis (initial intermolecular proton transfer to solvent) or initial intramolecular transfer to a phosphine ligand. Several other cases of η^2 - H_2 ligands reacting intramolecularly with thiolate and sulfide ligands are known or believed to be intermediate steps in, for example, SH ligand formation from reaction of sulfides with H_2 .^{297,298,321–333} and are relevant to biological systems such as H_2 ases. Particularly related to modeling the heterolysis of H_2 in H_2 ases is the work of Rauchfuss, who showed how the hydrido(hydrosulfide) complex $[Ir_2H_2(\mu-H)(\mu-SH)(\mu-S)(PPh_3)_4]$ is obtained from a double hydrogenation of the dinuclear iridium(II) complex $[Ir_2(\mu-S)_2(PPh_3)_4]$. In the stepwise process, the first added H_2 molecule undergoes *homolytic cleavage* while the second process is purely *heterolytic*.³²⁴ The related dicationic complex $[(\text{triphos})Rh(\mu-S)_2Rh(\text{triphos})]^{2+}$ [$\text{triphos} = CH_3C(CH_2PPh_3)_3$] is known to reversibly activate two dihydrogen molecules and produce the bis(μ -hydrosulfido) product $[(\text{triphos})(H)Rh(\mu-SH)_2Rh-$

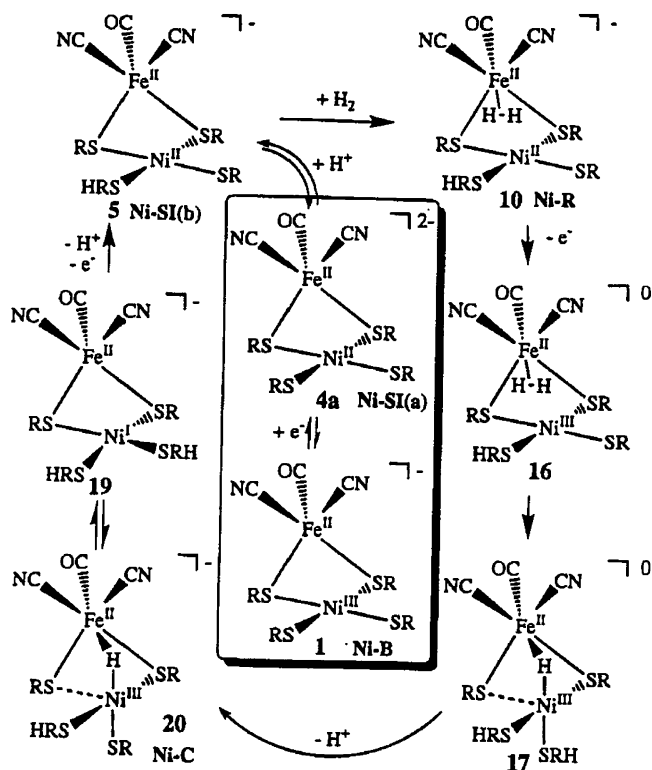
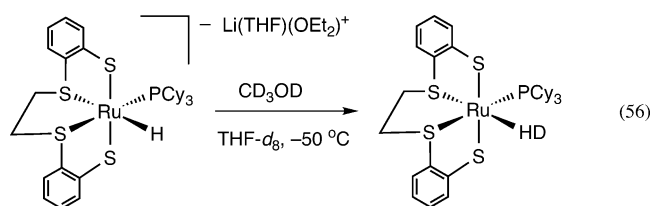


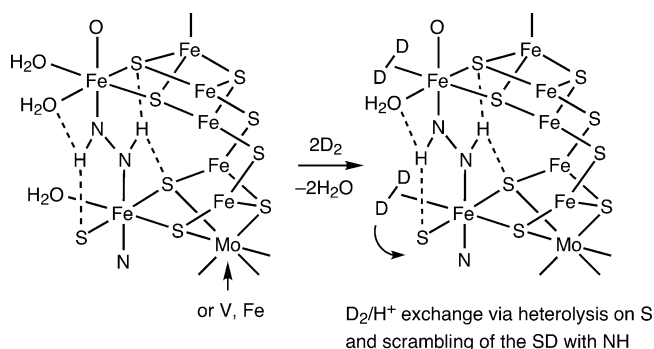
Figure 4. Possible mechanism for hydrogenase function as suggested by the calculations of Niu et al.³⁸³

$(H)(\text{triphos})]^{2+}$.^{297,321} DFT calculations show that each d^6 metal ion in a model complex, with local square pyramidal geometry, is able to anchor one H_2 molecule in the side-on coordination.³²¹ This is followed by heterolysis of the H–H bond over one adjacent and polarized Rh–S linkage and is repeated for addition of the second H_2 molecule. NMR experiments, including para-hydrogen techniques, identified that double heterolysis occurs in stepwise fashion, although there was no experimental evidence for a Rh- (H_2) adduct, probably due to its very short lifetime. The computational results support the energetic feasibility of the whole process, including its reversibility, which is favored by the unique proximity of electrophilic metal centers and nucleophilic sulfur atoms. In this case, the process compares (but is not exactly equal) to σ -bond metathesis, since the newly formed Rh–H and S–H bonds stem from H–H and Rh=S bonds. The mechanism differs from that for the above neutral Ir_2S_2 core, perhaps because the Rh complex is dicationic and more electrophilic, favoring double heterolysis.

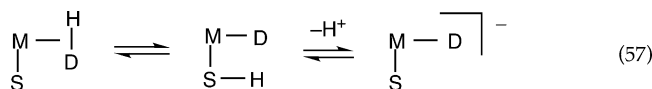
In order for proton transfer from a η^2 - H_2 ligand to a coordinated base to occur, the pK_a of the H_2 ligand and the protonated base must be similar (for a reversible process). Morris has estimated that coordinated alkanethiol ligands have pK_a values between 5 and 10, which matches well with the acidity of many H_2 ligands.³³¹ Protonation of an anionic Ru hydride using CD_3OD gives an unstable HD complex (eq 56).³²⁸ This reaction can be reversed by displacing the



Scheme 15

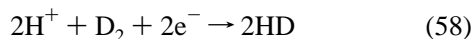


H₂ by DMSO to give Ru(DMSO)(PCy₃)(S₄), which yields Na⁺[RuH(PCy₃)(S₄)]⁻ and MeOH when treated with H₂ in the presence of NaOMe. This demonstrates that H₂ can be heterolytically cleaved at M–S sites, and a mechanism had been elucidated for an analogous neutral Rh–hydride system.^{326,327} In this case, the electrophilic metal and the basic thiolate donors attack the η²-H₂ in concerted fashion to give an identifiable thiol hydride species, [RhH(PCy₃)(^{bu}S₄-H)]⁺. The similarity between the Ru and Rh systems suggests that the HD (or a D₂) ligand in eq 56 can be intramolecularly cleaved (eq 57), which is essential to rationalize the D₂/H⁺ exchange between D₂ and EtOH that these complexes catalyze. For the Ru system, the thiol hydride could not be

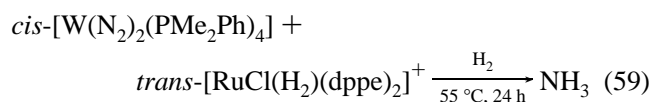


detected, while, for the Rh system and also [IrH₂(HS(CH₂)₃-SH)(PCy₃)₂]⁺ (which similarly catalyzes D₂/H⁺ exchange),³³⁰ the H₂ complex could not be seen but is a transient. A related system, Ni(NHPⁿPr₃)(S₃) clearly shows that heterolysis of D₂ can also occur at nickel sites, which may be relevant to H₂ activation in [FeNi] hydrogenases.³²⁹

Regarding the structure and function of nitrogenases in producing ammonia from N₂, Sellmann has studied several model systems wherein heterolytic activation of H₂ occurs on sulfur ligands.³³⁴ A core geometry based on a hybrid of the FeMoco active site structure with a dinuclear diazene complex, [Fe(“N₄S₄”)]₂(μ-N₂H₂), is a proposed model (Scheme 15). In nitrogenase (section 9), H₂ reduction is proven by the formation of HD from D₂ gas and protons derived from H₂O, which occurs only in the presence of N₂ (eq 58).

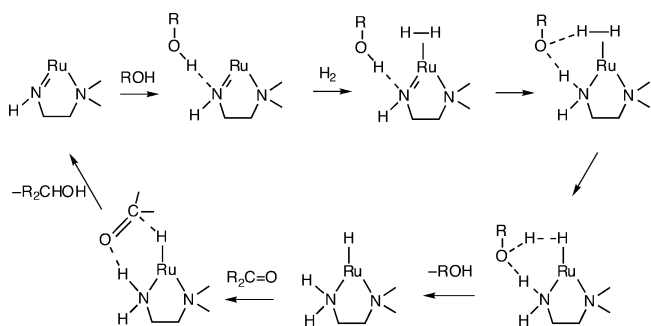


Sellmann's model is claimed to be consistent with the severe constraints imposed on this “N₂-dependent HD formation” from D₂ and protons. Other modeling studies have shown that protons can be transferred from acidic H₂ ligands in cationic Ru–H₂ complexes to N₂ ligands in W(N₂)₂(P)₄ complexes (P = phosphine donor), in some cases even forming ammonia (eq 59).^{335,336}

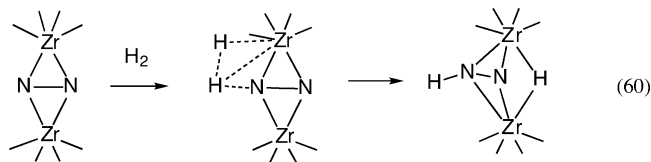


Detailed studies with several Ru(H₂) complexes showed that the yield of NH₃ critically depended upon the pK_a value of the Ru(H₂) complexes.³³⁶ When the W–N₂ complex was

Scheme 16



treated with 10 equiv of [RuCl(H₂)(dppe)₂]⁺ (dppe = 1,2-bis(diphenylphosphino)ethane) with pK_a = 6.0 under 1 atm of H₂, NH₃ was formed in up to 79% total yield (free NH₃ plus NH₃ released on base distillation). If the pK_a of the Ru–(H₂) complex was increased to ~10, the yield of ammonia decreased remarkably. Heterolytic cleavage of H₂ was proposed to occur at the Ru center via nucleophilic attack of the coordinated N₂ on the coordinated H₂, where the coordinated N₂ is protonated and a hydride remains at the Ru atom. Only a very limited number of reactions of bound N₂ with H₂ are known, e.g., eq 60, which slowly occurs in toluene over 1–2 weeks for a dinuclear Zr complex capped by macrocyclic ligands with N and P donor atoms.^{337,338}

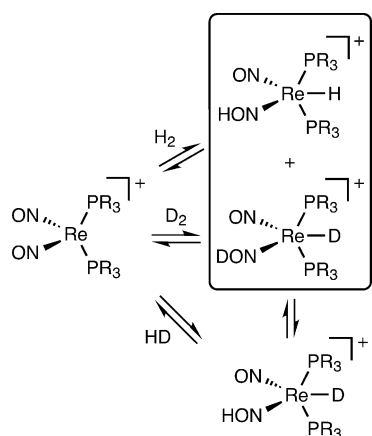


However, here the reaction stopped at the stage of N₂H, and no NH₃ was formed. Chirik recently found NH₃ is produced on reaction of H₂ with a similar μ-N₂ complex containing two methyl-substituted Cp ligands on each Zr.³³⁹ Remarkably, side-on N₂ bonding and NH₃ production occurred only upon a seemingly insignificant change from pentamethylated to tetramethylated Cp ligands. A related hafnocene system hydrogenated the N₂ ligand but did not produce NH₃.³⁴⁰ Heterolysis of H₂ also occurs on a Fe(μ-N)Fe species to form Fe(μ-NH)(μ-H)Fe species, but NH₃ was not seen.³⁴¹ Ammonia and hydrazine have been seen to form in bis(diphosphine)iron systems that are proposed to heterolyze H₂ to form protons. Here, H₂ becomes the actual source of electrons for N₂ reduction.³⁴²

The catalytic system discovered by the recent Nobel laureate, Ryoji Noyori, for asymmetric hydrogenation of simple ketones to alcohols is an elegant example of the importance of heterolytic activation of H₂ in a commercially valuable industrial process. This conversion is catalyzed by *trans*-RuCl₂[(*S*)-binap][(*S,S*)-dpen] (binap = [1,1'-binaphthalene-2,2'-diylbis(diphenylphosphane)]; dpen = diphenylethylenediamine) and is remarkable in several respects.^{343–345} The reaction is quantitative within hours, gives enantiomeric excesses (ee) up to 99%, and shows high chemoselectivity for carbonyl over olefin reduction, and the substrate-to-catalyst ratio is >100,000. The nonclassical metal–ligand bifunctional catalytic cycle is mechanistically novel compared to that of the structurally similar classical ruthenium hydrogenation catalysts (Scheme 16).

The process involves heterolytic splitting of H₂ assisted by coligands (see eqs 47 and 48 and ref 308) and possibly

Scheme 17



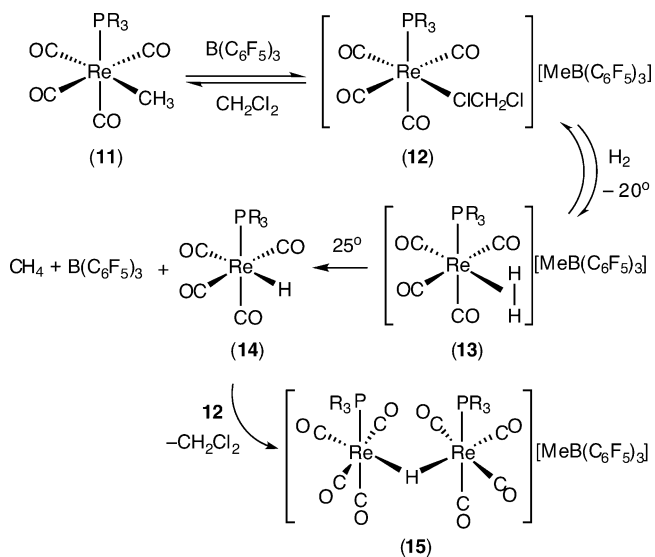
solvent to form a catalytically active Ru(hydride)(diamine) complex as a key step. Computational and experimental modeling studies involving similar heterolysis of H_2 in dihydrogen complexes have been shown by Morris and others to be the critical step in the mechanism of reaction processes related to the Noyori systems.^{86,346–348} Bergens reported the first direct observation of a cationic $[RuH(H_2)(\text{diphosphine})(\text{diamine})]^+$ complex as a putative intermediate, where the H_2 ligand was very labile and had the highest observed J_{HD} (37 Hz) to date.⁸⁶ Evidence suggests that H_2 heterolysis is the key step in Scheme 16 and can be facilitated by alcohols, underscoring the importance of alcohol-containing solvents in promoting heterolysis of H_2 here and in other metal bifunctional catalysis.^{348a,b} Base-assisted heterolysis of coordinated H_2 has been analyzed computationally for a $Rh(H_2)(PH_3)(HCO_2)\cdots NH_3$ model system.³⁴⁹ Both the kinetics and thermodynamics of the metathesis process for transfer of H to the oxygen of HCO_2 were favored by the presence of external amine. In Scheme 16, after the amide nitrogen cleaves H_2 , the resulting NH_2 functionality in the diamine ligand along with the hydride ligand deliver hydrogen to the ketone via a six-membered, pericyclic transition state, giving the alcohol product. Thus, the 18-electron Ru center and the ligands directly cooperate in the bond-breaking and bond-forming processes. The hydride on Ru possesses sufficient nucleophilicity, while the NH moiety exhibits a hydrogen-bonding ability to activate the carbonyl function.

Catalytic H/D scrambling of mixtures of H_2 and D_2 often takes place via intramolecular heterolysis of H_2 , as will be discussed further below. A recent example was proposed to involve cleavage of H_2/D_2 and proton transfer to NO ligands (Scheme 17).³⁵⁰ Although the protonated NO ligands were not actually observed, analogous heterolysis of a Si–H bond in a silane did give a complex with a silylated nitrosyl ligand, Et_3SiON . Reactivity directly analogous to that in Scheme 17, e.g., protonation of similarly π -accepting CO ligands, would not be expected in H_2 ases, since more basic sites are available, but nothing can be ruled out.

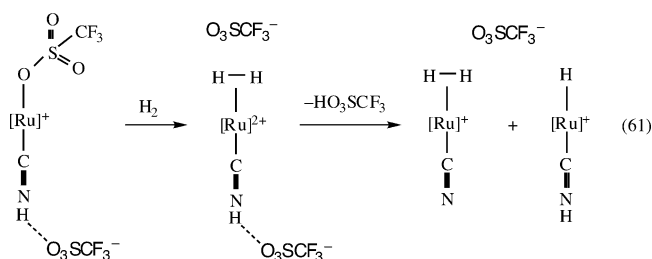
8.2.6. Proton Transfer to Anions

Strong acids such as HCl can be eliminated by proton transfer from η^2-H_2 ligands to the counteranions of highly electrophilic $[L_nM]^+$ complexes. One of the strongest acids known, *triflic acid*, CF_3SO_3H , can even be eliminated from a dicationic H_2 complex formed from reaction of H_2 gas with $[Ru(CNH)(PP)_2][OTf]_2$ (PP = diphosphine), which contained

Scheme 18



a coordinated triflate anion and a protonated cyanide ligand (eq 61).²⁷⁴ Another “superelectrophilic” 16e Ru complex,



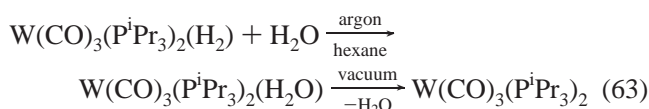
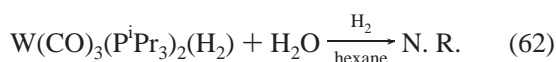
$\{Ru[P(OH)_3](PP)_2\}[OTf]_2$, heterolytically cleaves not only H_2 but other σ H–X bonds in silanes ($HSiR_3$) and boranes ($BH_3\cdot PR_3$) to give $\{RuH[P(OH)_3](PP)_2\}[OTf]$ plus $XOTf$ ($X = H, SiR_3, BH_2\cdot PR_3$).

A further interesting case involves protonation of borane anions where the d⁶ rhenium(I) complex, **11**, is in nearly 1:1 equilibrium with **12**, formed by methyl abstraction by $B(C_6F_5)_3$ to give the $MeB(C_6F_5)_3^-$ counterion (Scheme 18).²⁷⁷ This indicates that the electrophilicity of the $[Re(CO)_4(PR_3)]^+$ fragment is similar to that of $B(C_6F_5)_3$. **12** reacts under H_2 atmosphere below room temperature to form equilibrium amounts ($\sim 5\%$) of the H_2 complex (**13**). On warming the solution, methane, $B(C_6F_5)_3$, and *cis*- $Re(CO)_4(PR_3)H$ (**14**) form, apparently by protonation of the anion $MeB(C_6F_5)_3^-$ by the acidic H_2 in **13**. **14** is not observed by NMR but presumably quickly reacts with unreacted **12** (or **13**) to form the hydride-bridged dimer **15**, which is a “thermodynamic sink” in these systems (see eq 42). Another possible scenario in Scheme 18 is *intermolecular* heterolysis of H_2 , e.g. protonation of the Me group in equilibrium quantities of **11** by the acidic H_2 in **13** to give CH_4 , **12**, and **14**. Regardless of mechanism, this system demonstrates the stability of *hydride-bridged complexes* that have been proposed in the mechanism of H_2 cleavage/formation at the dinuclear active sites in hydrogenases.

8.2.7. Strength of Binding of H_2 Compared to Water and N_2 . Importance of Entropy Effects

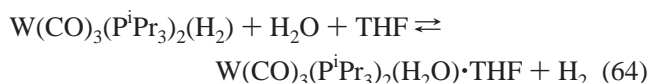
An important question is how can a seemingly weak ligand such as H_2 compete with stronger ligands such as water or even atmospheric dinitrogen that are present in the environ-

ment of life forms. It is illuminating to compare the binding energy of H₂ to that for the aqua ligand, H₂O, the archetypal lone-pair donor in classical coordination chemistry. Addition of excess H₂O to a concentrated tetrahydrofuran (THF) solution of W(CO)₃(PⁱPr₃)₂(H₂) gives instant vigorous effervescence of H₂, even under an H₂ atmosphere.^{226c} X-ray diffraction of the product obtained on crystallization showed it to be W(CO)₃(P-*i*-Pr₃)₂(H₂O)·THF, containing an H₂O ligand replacing the H₂ and lattice solvent (THF). The structure is novel in that the H-atoms on the aqua ligand hydrogen bond to the lattice THF oxygen atom and a CO oxygen on an adjacent molecule. Such hydrogen bonding in organometallic systems is becoming an increasingly recognized phenomenon,³¹⁶ and it is conceivable that hydrogen bonding of protein residues to CO ligands may be present in hydrogenase active sites (although weaker and less consequential than hydrogen bonding to the cyanide ligands). Interestingly, the aqua complex does not precipitate if addition of H₂O to W(CO)₃(PⁱPr₃)₂(H₂) is done in the nonpolar solvent *hexane* under an H₂ atmosphere with a large excess of water present as an immiscible phase.



As soon as the H₂ atmosphere is replaced by argon (eq 63), the less soluble H₂O complex precipitates. Subsequent exposure to vacuum rapidly leads to dissociation of H₂O and precipitation of insoluble W(CO)₃(PⁱPr₃)₂. This demonstrates the extremely delicate reversible nature of the H₂O and H₂ binding and indicates that *H₂ can compete both thermodynamically and kinetically with H₂O as a ligand*. A major factor is mass action, i.e., concentration of unbound ligand in solution. In hexane the low solubility of H₂O limits its maximum concentration to the same order as that of dissolved H₂ (ca. 0.005 M), as opposed to the situation in THF, where the high concentration of miscible H₂O overwhelms that of H₂. Other complexes demonstrating this effect are [Ru{HB(pz)₃}(PPh₃)₂(H₂O)]⁺³⁵² and [Ru(H₂O)₆]²⁺, where an H₂O ligand can be displaced by H₂ under pressurized H₂ even in H₂O solution.⁶¹ One of the first H₂ complexes, [IrH(H₂)(PPH₃)₂(bq)]⁺, was prepared by displacement of H₂O under 1 atm of H₂ in organic solvents.^{92,93}

The fact that H₂ and water can closely compete for the same binding site is clearly relevant to biological activation of H₂ by hydrogenases. The thermodynamic data below show that binding of H₂ should easily occur on large hydrophobic metalloenzyme sites where the effective H₂O concentration is low. The equilibrium constants for displacement of H₂ by H₂O in THF can be determined by IR data at several atm H₂ pressures at 25 to -70 °C.^{226b} The thermodynamic parameters for eq 64 are readily obtained from van't Hoff plots:



$$\Delta H = -4.5 \pm 0.2 \text{ kcal/mol};$$

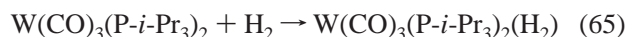
$$\Delta S = -18.8 \pm 2.0 \text{ cal/(mol deg)}$$

Displacement of H₂ by water is exothermic by 3–4

kcal/mol, but hydrogen bonding between coordinated H₂O and solvent appears to play a role in the thermodynamics. Also, bound H₂ has been shown to hydrogen bond to bulk H₂O solvent in a water-soluble Ru-diphosphine complex.³⁵³ In this case, the coordinated H₂ is surprisingly inert to substitution by water. Such species are proposed to be key intermediates in numerous important reactions such as the proton-transfer pathway of H₂ production by hydrogenase enzymes.

The surprisingly high negative entropy change in eq 64 no doubt reflects free THF becoming bound (three particles converting to two). The unfavorable entropy of binding of H₂O is largely the reason why the equilibrium favors H₂ binding at room temperature and H₂O binding at low temperature. ΔG₂₉₈ can be calculated to be 1.1 kcal/mol, i.e., favoring the left side of eq 64. *Entropic factors can thus be critical in competition between weak ligands for binding sites*, as will be seen below for N₂ versus H₂ binding.

The enthalpies of binding of H₂O in eq 64 are relative to H₂, so it is of interest to determine the enthalpy of binding of H₂ to W(CO)₃(PⁱPr₃)₂, which is directly measured to be -11.2 ± 0.5 kcal/mol in toluene at 20 °C (eq 65).



The affinity of H₂ versus other ligands such as N₂ for L_nM varies and can be entropy-dependent. In some cases, N₂ is a better ligand than H₂, and sometimes the opposite is true, or N₂ does not bind at all. Binding a gaseous ligand increases the total entropy of ML_n(H₂) relative to ML_n but does so by a relatively minor amount compared to the entropy lost by the ligand.⁸⁷ On this basis, the total entropy of exchange for eq 66 should depend primarily on the differences in absolute entropies for N₂(g) and H₂(g).



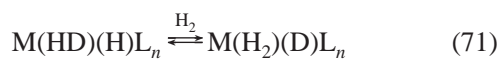
The third-law entropies, S°, of the two gases can be calculated by using standard formulas of statistical thermodynamics.⁸⁷ At room temperature, the entropy is due exclusively to the translational and rotational components. Due to its lower mass and moment of inertia, the absolute entropy of H₂ (31.2 cal/(mol deg)) is 14.6 cal/(mol deg) lower than that for N₂. If eq 66 is re-examined, it is clear that if the total entropies of the complexes in solution exactly canceled, the predicted entropy change would be 14.6 cal/(mol deg). This then favors the right side of eq 66, i.e., H₂ binding, since ΔG = ΔH - TΔS. Thus, because H₂ has the smallest absolute entropy (S°) of any diatomic gas, H₂ will be more competitive in binding relative to N₂ or other small molecules, which may be important in biological activation of H₂. Other factors include the electron-richness of the metal center, which is particularly dependent on overall charge. As the electrophilicity of M increases and M → L backdonation decreases, H₂ becomes an increasingly better ligand than N₂. The disparity here apparently stems from N₂ being a poor σ-donor,^{354–358} weaker than even H₂, although a good π-acceptor like H₂.^{155,355} Summarizing, nonclassically bound H₂ is a more versatile ligand than many classically coordinated ligands such as N₂ in the ability of H₂ to adjust to a larger range of electronic situations. It can also have steric (small size) and entropic advantages over other ligands.

8.2.8. Isotopic Exchange and Other Intramolecular Hydrogen Exchange Reactions

Hydrogen-containing systems readily lend themselves to isotopic substitution or labeling by deuterium and tritium.

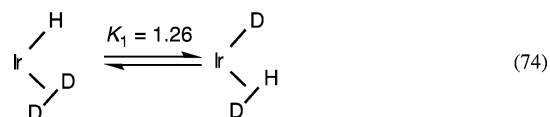
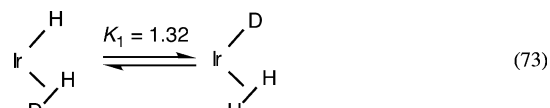
This is most useful in IR and NMR spectroscopic studies, particularly for determining J_{HD} , which is often critical to the proof of molecular H_2 coordination. Importantly, in the context of this article, transition metal-catalyzed H_2/D^+ and D_2/H^+ exchange reactions, where the H^+ and D^+ originate from water or alcohols, are of significant relevance to the study of H_2 ase enzymes.^{327,359,360} For example, the $\text{D}_2/\text{H}_2\text{O}$ exchange catalyzed by H_2 ases has been instrumental in monitoring the activity and studying the mechanism of this important class of enzymes.^{359a-d} Consequently, this exchange process has often been a primary screening tool for functional models of H_2 ases.^{359e-1,360} Such functional models usually invoke heterolytic cleavage of H_2 through the intermediacy of a transition metal dihydrogen complex, as discussed above. Recent interest in performing hydrogenations in aqueous solution has also spurred an interest in synthesis of water-soluble transition metal H_2 complexes and hydrogenation catalysts to catalyze this type of H/D exchange.^{61,303,342,359m,n}

Before isotopic exchange with water is discussed, it should be realized that H_2 , D_2 , and HD ligands can exchange and scramble with each other, with hydride ligands, or with H_2 (or D_2 or HD) gas. Usually, HD or D_2 ligands can be directly coordinated to metal centers by direct addition to unsaturated precursors such as agostic complexes. In some cases, however, a convenient precursor does not exist, and labeling can be done only by facile exchange of the H_2 ligand with HD or D_2 gas, possibly combined with intramolecular isotopic scrambling (eqs 67–71), or by adding a source of D^+ to a hydride complex (eq 72).³⁰



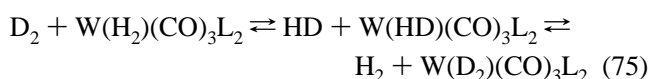
Intramolecular H/D exchange gives essentially a statistical mixture of isotopomers, but not always exactly statistical because deuterium usually prefers to be in the (HD) or (DD) site. Isotopomers can be detected by solution NMR or by IR in low-temperature matrices. Separate resonances for H_2 and hydride site isotopes are observed in the spectra of complexes when no intramolecular exchange occurs, but in cases where eq 68 is fast, only averaged chemical shifts and J_{HD} are observed. In the fast exchange ^1H NMR spectra of isotopomers of nonclassical polyhydrides, a phenomenon called isotopic perturbation of resonance (IPR) occurs.^{94,204a,206} For example, in a partially deuterated $\text{MH}(\text{H}_2)$ complex, each isotopomer (H_3 , DH_2 , and HD_2) shows a separate hydride resonance for the species provided the $\text{M}-\text{H}$ and $\text{M}(\text{H}_2)$ sites have significantly different chemical shifts and sizable deuterium fractionation exists between the sites. There is a nonstatistical site preference for the deuterium isotope that

varies with the degree of deuteration in $[\text{TpIrH}(\text{H}_2)(\text{PR}_3)]^+$ (eqs 73 and 74).²⁰⁶ The equilibrium constants shown are



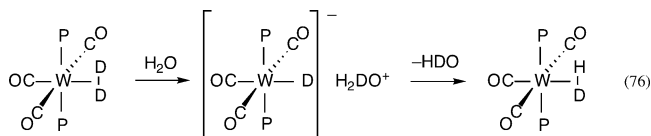
actually Boltzmann factors (statistics not included), but they indicate that the heavier isotope prefers to occupy the hydride site.

H_2 complexes containing hydride ligands, $\text{M}(\text{H}_2)\text{H}_x\text{L}_n$, are usually effective catalysts for $\text{H}_2/\text{HD}/\text{D}_2$ scrambling, but several coordinatively saturated H_2 complexes with no hydrides also catalyze exchange. While the former exchange has several reasonable pathways, scrambling of D_2 with $\text{W}(\text{CO})_3(\text{PR}_3)_2(\text{H}_2)$ and a few other 18e complexes as in eq 75 is more enigmatic.^{2,88b,361-363}



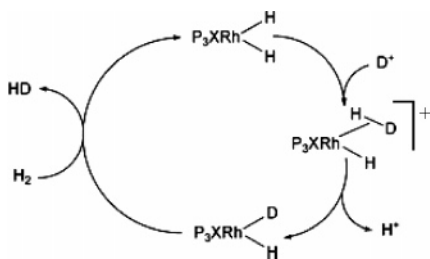
Equimolar amounts of D_2 gas (1 atm) and the H_2 complexes give complete isotope equilibration *even in the solid state* within days for group 6 species or 12 h for $[\text{Re}(\text{CO})_3(\text{PR}_3)_2(\text{H}_2)]^+$ in solution. Prior loss of CO or phosphine to allow D_2 into the coordination sphere followed by isotopic exchange as in eq 69 seems unlikely because ligand loss would be a high-energy process, especially in the solid. Possible mechanisms could involve seven- or eight-coordinate 20e intermediates such as a $(\text{H}_2)(\text{D}_2)$ complex or a dihydride–dideuterium complex, $\text{WH}_2(\text{D}_2)(\text{CO})_3(\text{PR}_3)_2$. However, no evidence exists for either the dihydride form in the solid state or seven- or eight-coordinate complexes of the type discussed here.

Trace quantities of adventitious water may lead to exchange, since isotopic scrambling of the D_2 ligand in $\text{W}(\text{CO})_3(\text{P}-i\text{-Pr}_3)_2(\text{D}_2)$ with H_2O occurs in solution within days^{226c} or less for other metal- D_2 complexes.^{94,326,353,359m,n,360,364-366} A reasonable mechanism for exchange for complexes with one open coordination site is deprotonation of $\eta^2\text{-H}_2$ by the weak base water followed by reprotonation with H_2DO^+ . Such a mechanism may be



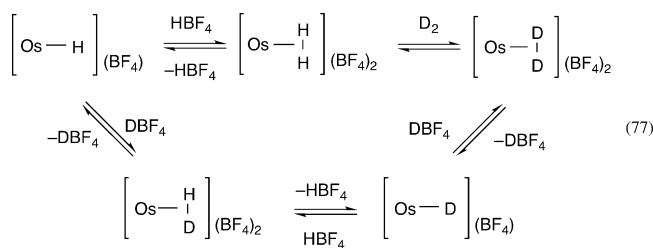
important in isotopic exchange processes in enzymatic systems such as H_2 ases and N_2 ases. As discussed above, $\eta^2\text{-H}_2$ can be quite acidic and is known to hydrogen bond to water.³⁵³ Kovacs proposed a mechanism for $\text{Rh}(\text{TPPMS})_3\text{Cl}$ catalyzed $\text{H}_2/\text{D}_2\text{O}$ exchange (TPPMS = water soluble phosphine) where the catalyst first undergoes oxidative addition of H_2 to make the dihydride (Scheme 19).³⁵⁹ⁿ A hydride ligand can then react with D^+ to form an HD ligand, which can lose H^+ to create isotopic exchange. A similar mechanism was proposed for $\text{TpRuH}(\text{PPh}_3)(\text{CH}_3\text{CN})$ where D_2O initially hydrogen bonds to the hydride ligand, followed by transfer of D^+ to give a cationic HD complex

Scheme 19



with an OD⁻ anion.³⁵⁹ This may also be a possibility in exchanges such as in eq 76, and W(CO)₃(P-*i*-Pr₃)₂(D₂) is known to exist in solution equilibrium with its dideuteride isomer, WD₂(CO)₃(P-*i*-Pr₃)₂.

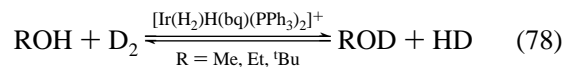
For cationic complexes such as [Os(H₂)(CH₃CN)(dppe)₂][BF₄]₂ formed by protonation of [OsH(CH₃CN)(dppe)₂][BF₄] by [H(OEt₂)BF₄], isotopic exchange with D₂ gas occurs (eq 77).³⁶² Reversible deprotonation of the D₂ ligand by ether present in CD₂Cl₂ solvent is proposed to occur, forming equilibrium amounts of “free” acids, HBF₄/DBF₄ (these are actually present in eq 77 as ether solvates H[OEt₂]BF₄/D[OEt₂]BF₄). This facilitates complete exchange to give the HD complex. The isotopic exchange in CD₂Cl₂ is slow



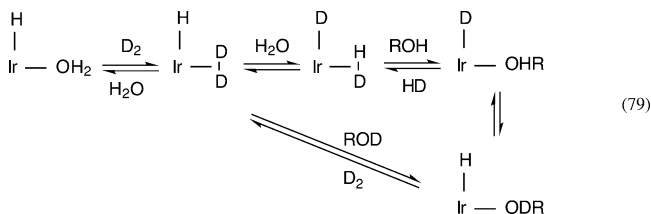
(days), as for the W(CO)₃(PR₃)₂ system, and the deuterio solvent does not become involved (see below). However, in eq 76, much stronger bases than H₂O, such as alkoxides,³⁶⁷ are required to deprotonate the W complex. Also, the rate of H₂/D₂ exchange is much faster than H₂O/D₂ exchange, which is unlikely to occur as above in the solid state and is not seen for solid W(CO)₃(P-*i*-Pr₃)₂(D₂) plus H₂O. This pathway could operate in solution for systems with more acidic η²-H₂, but another explanation is needed for scrambling in group 6 complexes.

In solution, isotopic incorporation of deuterium from deuterated solvents into metal-bound hydrogen is common; for example, reaction of acetone-*d*₆ and [RuCl(dppe)₂(H₂)]⁺ or [OsH(H₂)(PP₃)]⁺ gives the HD isotopomer in 20 min and the fully deuterated complexes in a few hours.^{94,106} Complexes with both hydride and H₂ ligands such as [Ir(H₂)H-(bq)(PPh₃)₂]⁺ and Ir₂H₃(μ-H)(H₂)(μ-Pz)₂(P^{*i*}Pr₃)₂ or unsaturated hydrides such as IrClH₂(P^{*i*}Pr₃)₂ are advantageous for such isotopic exchange. This is because ligand exchange involving H₂, D₂, and substrates with exchangeable protons is facile, and barriers to intramolecular exchange with cis hydride ligands are low. The latter two complexes undergo H/D scrambling with toluene-*d*₈ solvent, which could bind to Ir by adding as a sixth ligand or displacing

H₂.^{78a,368} The cationic Ir complex is an excellent catalyst for deuterium incorporation into alcohols for example (eq 78).³⁶⁰

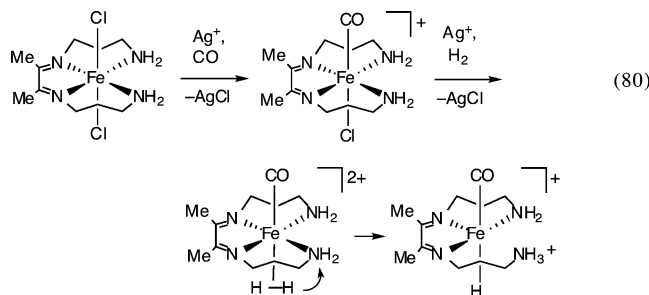


In addition to a possible deprotonation mechanism as in eq 76, a mechanism involving exchange with the cis-hydride is likely here (eq 79).



8.2.9. The Need for a Low-Spin State in H₂ases and the Possible Role of Cyanide Ligands

Another important question is why does Nature utilize toxic cyanide ligands in hydrogenases? CN ligands could be involved in proton transfer or important hydrogen-bonding interactions with protein components. The cyanide complex, [Fe(H₂)(CN)(R₂PC₂H₄PR₂)₂]⁺, is known and can indeed exist as an FeH(CNH) tautomer depending on R.¹⁰⁷ A more likely role for the cyanide ligands relates to the spin state of hydrogenases, which are known to be *low spin* in all redox states. Why then is a low-spin state crucial? The answer comes from fundamental inorganic and organometallic coordination chemistry. In accord with the general principles of transition metal chemistry,^{369,370} the overall ligand field strength strongly influences the spin state of the dimetallic active sites, which generally feature Fe(CO)(CN) moieties linked by thiolate bridges. As will be shown below, this must be taken into account in efforts to model any facet of hydrogenase chemistry. If one assumes that carbonyl (CO) ligands are critical in hydrogenases (section 3.2), their binding to iron must be very strong to both maintain the integrity of the active site and prevent poisoning of the host organism by release of CO. CO is a very powerful ligand and has been characterized to be a “universal ligand” to lower-valent metal centers.¹⁶¹ Strong CO binding to iron in hemoglobin is particularly notorious in regard to the toxicity of CO. Of particular relevance in Fe–heme systems is the spin-state change (spin crossover) from high-spin Fe^{II} (*S* = 2) to low-spin Fe^{II} (*S* = 0) on CO binding,^{369,371–374} which is much less facile in inorganic and organometallic complexes than may generally be appreciated. Anomalously weak CO binding in Cp₂VI(CO) and Cp₂Cr(CO) was noted decades ago independently by Calderazzo³⁷⁵ and Brintzinger,³⁷⁶ both of whom rationalized that spin pairing has to take place upon carbonylation of the high-spin fragments. In his review article on such effects of the spin state, Poli³⁷⁰ notes that “in spite of this early work, the importance of electron pairing in organometallic stability and reactivity has remained essentially unappreciated.” This was encountered in attempts by Kubas to bind CO to iron(II) complexes with nitrogen-donor ligands to model heterolytic cleavage of H₂ as in hydrogenases.³⁷⁷ The intent was to synthesize Fe^{II} complexes with CO trans to H₂ in order to observe *intramolecular* heterolysis of H₂ where a proton transfers to a basic cis N-donor ligand, e.g., via eq 80, similar to that in eq 52.



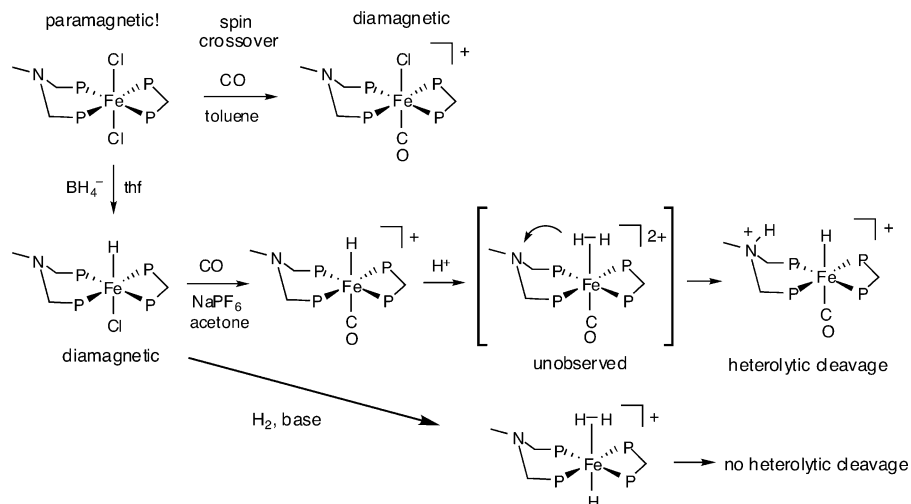
Similarity to the structure of hydrogenases was not of concern, and the multidentate α -diimine ligands had been previously studied on Pd^{II} and Pt^{II} centers.³⁷⁸ The important feature is that the diimines contain basic pendant side arms (the amine groups in eq 80) that could accept a proton from H₂ heterolysis. Intramolecular heterolysis of η^2 -H₂ on Fe^{II} centers as in eq 80 had not been previously directly observed, although while this work was in progress, DuBois³⁷⁹ independently found such heterolysis in a related phosphine system, *trans*-[Fe(X)(Y)(PNP)(dmpm)]⁺ (dmpm = dimethylphosphinomethane), also containing a proximal basic amine group in the chelating PNP ligand (Scheme 20). Although the precursor dichloro complex was high spin, spin crossover to low-spin complexes occurred on CO addition or replacement of Cl by H. Protonation of [FeH(CO)(PNP)(dmpm)]⁺ was observed to give a final product with the proton on the basic N atom of the PNP ligand, implying that an incipient unobserved H₂ ligand, if formed, would heterolytically cleave. However, when a hydride is positioned *trans* instead of CO, H₂ binds but does not heterolyze to protonate the amine. Thus, heterolysis of η^2 -H₂ is much more effective when CO is *trans* to it, in keeping with the principles in sections 3.2 and 8.2.2 outlining how it appears that Nature was opportunistic in employing CO ligands for this purpose.

In eq 80, stepwise removal of chloride ligands from a dichloro precursor using Ag⁺ would have been expected to produce a complex with H₂ *trans* to CO, and the acidic H₂ ligand might then protonate the *cis* pendant amine. However, the very first step unexpectedly proved to be a major barrier: the metal–diimine system *rejected* binding of CO. The apparent rationale here is that the iron is in a high-spin state in the Fe(diimine)Cl₂ precursor and [Fe(diimine)Cl]⁺ fragments formed on Cl abstraction and does not undergo spin crossover to a low-spin state that would appear to be

necessary for stable CO binding. However, DuBois had found that *trans*-FeCl₂(PNP)(dmpm) is also paramagnetic but does not directly react with CO to displace chloride to form diamagnetic [*trans*-Fe(PNP)(dmpm)Cl(CO)]⁺, a rare example of a “spin-blocked” reaction, where a barrier may exist due to the crossing between reactant quintet and product singlet surfaces. Whether spin-state changes inhibit organometallic reactions has been a decades-old debate and has recently been shown computationally by Harvey and Poli to be highly dependent on the system.³⁸⁰ However, this and other current literature indicate that the term “spin-block” (or “spin-forbidden”) should be reserved for *kinetic* effects, and theoretical calculations on CO interaction with model Fe^{II}–diimine centers demonstrated that the lack of CO binding is *thermodynamic* in origin. Addition of CO to a high-spin Fe–diimine model complex was essentially thermoneutral. Thus, in the failed nitrogen donor system (eq 80) versus the successful phosphine donor system (Scheme 20), the *ligand field strength* of the N-donor versus P-donor ligands is of critical importance. The diimine complexes do not bind CO even weakly, but as expected, analogues containing diphosphines with strong ligand fields (strong electron donors) bind CO tightly, even in cationic species.

It thus may seem ironic that binding of CO to hemoglobin is one of the few facile “spin-forbidden” reactions of this toxic molecule with high-spin Fe^{II} centers. On the other hand, Nature has designed hydrogenases to possess low-spin Fe centers that powerfully and purposefully bind CO. Hydrogenases must possess enough electron density at iron to strongly bind CO while maintaining a fine balance of electrophilic character to reversibly bind and heterolytically cleave H₂. The peculiar presence in these enzymes of *cyanide* ligands could then be related to their high ligand-field strength. This would assist in maintaining a low-spin configuration for Fe throughout the large known array of redox state and ligation changes^{381–383,229,232} that occur during the function of the enzyme. Dissociation of either the CO or CN ligands would be destructive to the active site here. Weaker-field ligand sets than CO/CN such as those typically found in enzymes (histidine, cysteine, etc.) would not fulfill this function, since nitrogen-donor ligand sets such as imine/

Scheme 20

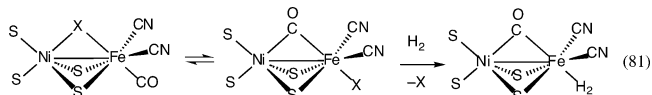


amine in eq 80 give *high-spin* complexes incapable of even weak CO binding. It is significant that CN can be formed biologically along with CO,^{384,385} unlike organophosphines or most other strong field ligands in inorganic and organometallic complexes. In the above context, Rauchfuss had previously also demonstrated the positive influence of cyanide on binding of CO to Fe^{II} and on facilitating carbonylation of Fe^{II} thiolate complexes.³⁸⁶ Darensbourg had speculated that an anionic cyanide would help stabilize a bridging CO ligand.^{248b} Another possible role for a strongly electron-donating cyanide ligand is its influence on the redox potentials, e.g., lowering the electrochemical potential for H₂ production.

8.2.10. Why Do Enzymes Such as H₂ases Have Polymetallic Active Sites with Metal–Metal Bonds?

An obvious question is why are two metals employed by most H₂ases when one would seem to work for H₂ splitting/formation as in organometallic chemistry? The active sites of nitrogenases, oxygenases, and certain other non-heme enzymes also contain two or more transition metals (most often Fe, Mn, Ni, Cu, Mo) in close proximity.^{242,387} Bonding between the metals can involve two electrons or less, as in organometallic dithiolate-bridged Fe dimers where Fe–Fe separations are ~2.6 Å for a normal two-electron bond, ~3.0 Å for a one-electron³⁸⁸ (“half”) bond, and >3.4 Å for no bond.³⁸⁹ These interactions allow complexes to exist in multiple oxidation states interconvertible by reversible one-electron-transfer steps if necessary. Multifunctional Fe₂S₂, Fe₃S₄, and Fe₄S₄ clusters containing Fe–Fe bonds are as common as heme groups in biology and facilitate electron transfer, influence protein structure, and can act as catalysts and sensors.³⁹⁰ Antiferromagnetic coupling via oxo-bridges in methane monooxygenase compounds reduces the Fe–Fe separation to as low as 2.46 Å for O₂ activation.³⁹¹ In nitrogenase, changes in Fe–Fe bonding by electron addition to the MoFe₇S₉ cofactor and/or P-cluster may be crucial to the binding and activation of N₂.^{392–394} Other functions of polymetallic sites include molecular recognition and stabilization of transition states by charge delocalization over multiple atoms.^{242,387}

An obvious question remains though: why are two metals employed by H₂ases? As will be discussed below, the electron-transfer process could be facilitated in some way, e.g., via M–M bonds, but also the active site is much more flexible in terms of stereochemistry and reactivity. The dinuclear site has three types of bridging ligands that can easily shift positions between bridging and terminal sites while the dinuclear configuration is retained: CO, hydride, and even SR (though less likely). These shifts are well-known in organometallic chemistry and could position the critical CO ligand trans to H₂ (e.g., in a bridging location) to favor its heterolysis. This could be especially important in the Ni–Fe H₂ases where the CO is trans to X in the crystal structure but could shift to a bridging position to become trans to the site of H₂ binding and subsequent heterolysis. An H₂ ligand



has yet to be definitively observed to bridge two or more metals in inorganic complexes, so this is unlikely to happen in the enzyme. Bridging *hydride* on the other hand is well-

known and has been proposed in the H₂ase mechanisms. As will be discussed below, M–M bonds can be quite basic and can be protonated,³⁹⁵ perhaps the first step in the H₂ production mode of the enzymes.

8.2.11. Mechanism of Hydrogen Activation in Hydrogenases

Much effort has been carried out in modeling the active site of H₂ases both experimentally and computationally in an effort to understand the mechanism of H₂ activation and is the subject of many publications both in this thematic issue and elsewhere. Therefore, the discussion here will be restricted to application of well-established principles of organometallic chemistry and dihydrogen activation (as detailed above) that could aid understanding the mechanism of biological H₂ activation. Theoretical calculations using data from the X-ray structures provide guidance for the mechanism of H₂ activation and are addressed in the article by Siegbahn in this thematic issue and other publications.³⁹⁶ Some computational aspects will be discussed here in conjunction with the organometallic principles. There are many mechanistic possibilities at the multifaceted dinuclear active sites of H₂ases, and some aspects of H₂ase chemistry are still poorly understood or controversial. However, it is generally agreed that the critical step of the mechanism in H₂ conversion to protons and electrons involves heterolysis of an H₂ ligand initially (and perhaps only transiently) bound to a metal center in the active site. In regard to computational analysis, Siegbahn stated that energies are in general more critical tests of a model than are structures, and it is important that they match the experimental energetics of the H₂ reaction.^{396b,c} The activation of H₂ should have a barrier of ~10 kcal/mol, be slightly exothermic, and most likely include an H₂ complex along the reaction coordinate. His early calculations on modeling the Ni–Fe H₂ases established that the only site to which H₂ binds significantly (binding energy computed to be 3.1 kcal/mol) is the electrophilic Fe (where *d*_{HH} = 0.78 Å). This was later supported by Niu and Hall³⁹⁷ and is consistent with organometallic systems where nickel is not known to form stable H₂ complexes and indeed very few Ni hydrides are known. The estimated barrier height for H–H cleavage is 8.7 kcal/mol, a reasonably low energy in accord with an enzymatic process.

Calculations by other researchers indicate that the Ni site, possibly as high-spin Ni(II), could be involved in the activation, so this is still a controversial area.³⁹⁸ It is likely that a complex with a hydride ligand bridging both metals is an intermediate in the mechanism, as will be discussed below. This was inspired by ENDOR studies that indicate that two types of exchangeable H nuclei are present in the vicinity of the Ni ligands in the Ni–C active form of a [NiFe] enzyme, consistent with *μ*-H.³⁹⁹ More recently, Lubitz directly detected by ENDOR a hydride ligand (presumably formed by heterolysis of H₂) occupying a bridging position at the Ni–Fe center of *Ralstonia Eutropha* in its reduced state.^{251c} Thus, it appears that either the nickel or iron center could be involved in forming an incipient Ni–H₂ complex that undergoes intramolecular heterolysis to form the bridging hydride. Since a bridging *dihydrogen* ligand has yet to be observed in the vast array of inorganic H₂ complexes, it is unlikely that both metals initially cooperate in binding H₂ in a bridging position. DFT calculations by Hall postulate iron as the site of initial H₂ binding/heterolysis and incorporate monoanions as some of the key intermediates (Figure 4).³⁸²

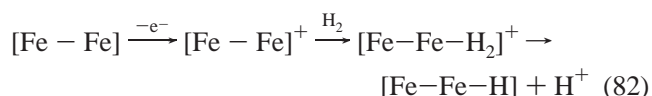
These computations do not take into account the protein backbone or hydrogen bonding of the CN to the protein known⁴⁰⁰ to be present, potentially important considerations. Optimized geometries reveal that H₂ prefers to bind to Fe rather than Ni, and d_{HH} is again 0.77 Å, although the H₂ is trans to CO rather than CN here. The Fe^{II} center is perfectly configured for capture of H₂ as it diffuses to the active site. The H₂ coordination leads to an increase in d_{NiFe} with respect to that in Ni–SI. The proposed mechanism for H₂ activation again features hydride-bridged frameworks for the key intermediates that would be expected to be present on such dinuclear sites, as suggested by Fontecilla-Camps.^{251a} It is notable that heterolysis of H₂ on organometallic complexes can lead to hydride-bridged complexes (Scheme 18 above), although the mechanism is different. The calculated d_{NiFe} values vary greatly in these species as shown, and this flexibility would be expected to facilitate both the electron- and proton-transfer processes (the M–M bond is a possible site for protonation). Although the proposed mechanisms may not be completely correct, the structure/bonding principles mirror those of H₂ activation on organometallic complexes.

The Fe–Fe H₂ases are even more organometallic in character and have been the focus of more modeling studies than the Ni–Fe enzymes. The bridging CO in Fe H₂ases is crucial because it places CO trans to the aqua ligand located crystallographically on Fe (Scheme 10), as in W(CO)₃(PR₃)₂–(H₂O), wherein H₂ is known to displace H₂O, and H₂ binding is favored by 1–2 kcal/mol in terms of ΔG (section 8.2.7). In *C. Pasteurianum*, the probable site for H₂ binding/elimination is thus trans to μ -CO, which would stabilize σ H₂ coordination, favor reversible binding and elimination of H₂, and promote heterolytic cleavage. As discussed above, the CO ligands in H₂ases would appear to be designed by Nature to increase the electrophilicity of the active site, thereby enhancing intramolecular heterolysis of H₂ as in carbonyl-rich [Re(CO)₄(PR₃)(η^2 -H₂)] [MeB(C₆F₅)₃] (Scheme 18). As discussed in section 8.2.7, such electrophilic metal sites as also in [Mn(CO)₃(PCY₃)₂]⁺ greatly favor binding of H₂ over N₂, which is well-known to bind to low-valent organometallic complexes with more electron-rich nucleophilic metal centers. Atmospheric dinitrogen is a potential competing ligand in enzymes with low-valent metallo sites such as H₂ases (Fe(I) and/or Fe(II) oxidation states) that could inhibit their function. This is thus another reason that H₂ases possess some electrophilic character and employ CO ligands for this purpose. The inorganic models for the active sites based on (CO)₂(CN)Fe(μ -SR)₂Fe(CO)₂(CN) type cores also do not bind N₂. It is notable that nitrogenases that *do* bind and activate atmospheric N₂ as their primary function have more nucleophilic metallo centers without electron-withdrawing CO ligands.

The highly electrophilic [Re(CO)₄(PR₃)⁺ center with four CO ligands also coordinates H₂O trans to CO,²⁷⁷ although the aqua ligand is less labile than in the neutral W(CO)₃–(PR₃)₂(H₂O) and appears to be more strongly bound than H₂. Thus, the active site in H₂ases cannot be overly electrophilic or aqua ligands would bind tightly and inhibit H₂ binding. Again, H₂ases have a proper balance of electrophilic and nucleophilic character, with the Fe center in Ni–Fe H₂ases and Fe(2) in Fe–Fe H₂ases being the more electrophilic sites for H₂ Fe–H₂O binding and heterolysis. Binding energies up to 23 kcal/mol have been calculated in a model Fe^{II}–Fe^{II}(H₂O) species for *D. Vulgaris*, but reduction to Fe^I–Fe^{II} can release H₂O to make the site available for

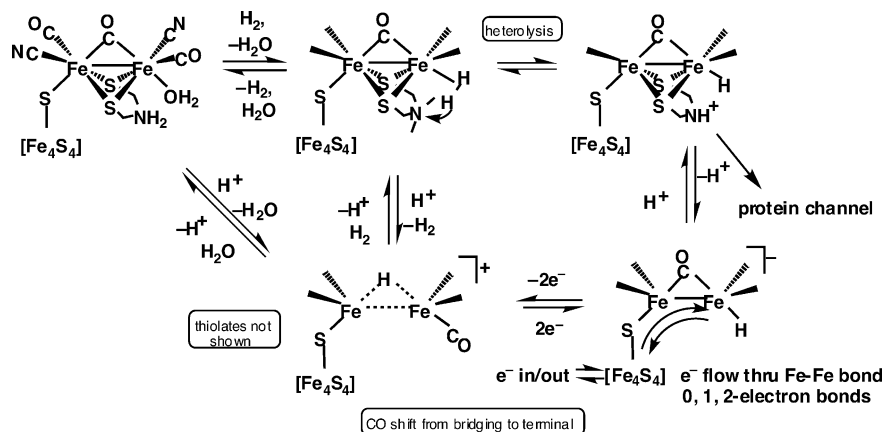
H₂ binding,^{381,401} which may be key to activation of the oxidized inactive form. Addition of H₂ to a Fe^I–Fe^{II} species with an empty coordination site is computed to be exothermic by 6.1 kcal/mol ($d_{\text{HH}} = 0.824$ Å in the resulting η^2 -H₂), and this EPR-active species is postulated to convert to an EPR silent Fe^{II}–Fe^{II}(H₂) form via electron (or proton) transfer.⁴⁰¹

Another of the many variables is the overall charge on the metals in the active site. In organometallic complexes, positively charged (i.e., cationic) metals greatly increase heterolytic cleavage of H₂. Thus, [W(CO)₃(PR₃)₂(H₂)]⁺ is much more easily deprotonated (by ethers) than the neutral complex, which requires a strong base (section 8.2.3).¹⁸¹ In H₂ases, it is likely that the Fe active site is somewhere midway in electrophilicity but could be tuned by oxidation of a neutral active site to a cationic one (or vice versa). Thus, for conversion of H₂ to protons and electrons, heterolysis of H₂ could be “switched on” by initial removal of an electron from the dimetallo core.



The proton would initially be expected to transfer intramolecularly to a basic site and then intermolecularly away from the active site. This process could then be repeated to remove the hydride as a proton. The metal–hydride bond is an interesting paradox in inorganic chemistry in that it can vary from being hydridic (acting as H[–]) to protonic (acting as H⁺) to anywhere in between.^{49,270c,292} Thus, the “hydride” in a metal hydride complex can be fairly acidic (protonic) and removable as a proton, especially if the coligands are CO, e.g., FeH₂(CO)₄. The “hydricity” of hydride complexes has been intensively studied by DuBois and Curtis.^{270c,292}

The active site of *D. desulfuricans* is similar to that of *C. Pasteurianum*, but in lieu of μ -CO, a monatomic oxygen species such as H₂O or OH apparently bridges the irons (it could also be terminal) and Fe(2) is proposed to be coordinatively unsaturated.²⁴⁷ A 1,3-propanedithiolate type ligand bridges the Fe, where R could also be –CH₂NHCH₂– with a basic nitrogen site to accept protons from H₂ heterolysis. Assuming accurate crystallography, one explanation of the structural differences is that the two structures represent different oxidation states and that the open coordination site in *D. desulfuricans* is the potential site for H₂ binding (it may actually be occupied by H₂, since crystallization was done under H₂). Also, shifts of CO between terminal and bridging positions and similar ligand rearrangements are extremely facile in organometallic systems, so in the enzyme mechanism, H₂ and hydride ligands could be positioned trans to a variety of ligands in either bridging or terminal sites. Calculations (below) show that such transformations are nearly barrierless processes on models for the active site. Because of the many easily accessible ligand arrangements and strong trans-ligand influences, the active site is tremendously flexible for either consuming or releasing H₂, adjusting the acidity of η^2 -H₂ for heterolysis, and attaining the relatively low redox potentials typical of these active sites. As stated by Pardo et al. regarding DFT studies on Ni–Fe H₂ases, the channel for H₂ cleavage/formation is very wide, and the enzyme may be a good catalyst because there are many low-energy productive reaction coordinates.³⁹⁸ With this in mind, Scheme 21 presents one (of many) reasonable mechanism for

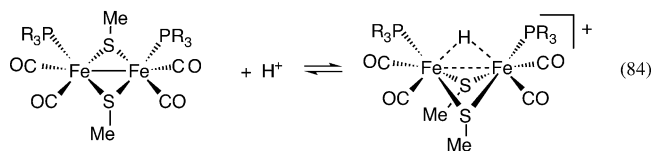
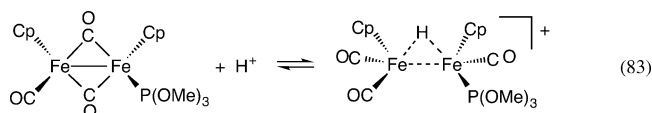
Scheme 21. Possible Mechanism for Hydrogenase H₂ ⇌ 2H⁺ + 2e⁻

reversible H₂ consumption/production on the Fe–Fe enzyme *C. Pasteurianum* that has been proposed by this author.^{6,277}

This mechanism is intended primarily to illustrate the basic principles of organometallic systems that can be applied to the function of the Fe–Fe active site here and possibly other H₂ases as well. Scheme 21 assumes that, as generally believed, one CN and one CO is coordinated to each Fe^{II} and a low spin d⁶ Fe^{II} octahedral configuration is present, which is well-known to favor H₂ binding. The transformations shown may involve participation of intermediate species not shown. Although there is yet no observable evidence for H₂ coordination in any form of the H₂ase enzymes, an H₂ complex of a rudimentary model for a H₂ase active site, [Ru₂(μ-H)(μ-S₂C₃H₆)₂(H₂)(CO)₃(PCy₃)₂]⁺, has been synthesized, albeit with Ru instead of Fe and with phosphine ligands that do not occur in enzymes.⁴⁰² The *J*(HD) value for the HD complex is 31 Hz, indicative of *d*_{HH} = 0.90 Å, i.e. a true H₂ complex. Solutions catalyze H₂/D₂ exchange, which is characteristic of H₂ases. In the mechanism for H₂ consumption in Scheme 21 (conversion to electrons and protons), an intermediate Fe–H₂ complex is produced by displacement of the H₂O ligand in the enzyme's "precursor" form observed crystallographically. The bound H₂ then heterolytically cleaves and transfers a proton to, for example, the basic amine functionality proposed to be present on the thiolate bridge in close proximity to the H₂.³⁸¹ Both Crabtree and Morris have demonstrated that such intramolecular heterolytic cleavage (and its microscopic reverse reaction) readily occurs in organometallic complexes, as exemplified by the equilibrium proton transfers shown in eqs 52 and 55. Transfer of a proton from η²-H₂ to the μ-thiolates in H₂ases is also possible as in eq 55. Calculations support such heterolysis, although it is endothermic by 15 kcal/mol.⁴⁰¹ Transfer of a proton to CN is nearly isoenergetic, but a high barrier is computed (38 kcal/mol, compared to 17 kcal/mol for transfer to sulfide). Oxidation to a cationic center could precede heterolysis and favor it, as in eq 82. The next steps for H₂ consumption involve movement of protons away from the active site to protein channels and synchronous or asynchronous electron transfer to the cubane cluster and away from the site via other Fe–S clusters. The electrons in the H–H bond could essentially flow through the Fe–Fe bond and, depending on whether one- or two-electron-transfer processes take place, one-electron Fe^{••}Fe bonds (2.9–3.1 Å)³⁸⁸ may be present in the intermediates (a two-electron-transfer step is shown in Scheme 21). The high flexibility of the M–M separation (2.6–3.2 Å, corresponding to 0, 1, or 2e M–M bonds) could facilitate electron/proton transfer

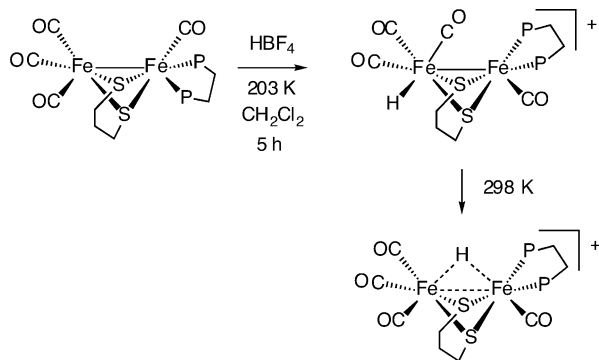
here and in the [NiFe] H₂ases. As will be discussed below, the M–M bond can easily be protonated to form a bridging hydride (and deprotonated) as part of the mechanism. Also, ligand shifts between bridging and terminal positions involving CO as well as hydride ligands are extremely facile in dinuclear organometallic complexes and are likely to occur here as well. Once the H₂ is converted to electrons and protons, or in the reverse reaction is eliminated, recoordination of an aqua ligand is unnecessary and would only slow the reaction rate. It is likely that the intermediate with the bridging hydride transfers the H away from the active site (as a proton) and another H₂ molecule immediately recoordinates to start another catalytic cycle.

The reverse reaction, formation of H₂ from 2H⁺ and 2e⁻, involves protonation of the 2Fe center to form a metal hydride. The most basic site for initial protonation in the enzyme active sites may be the electrons in the M–M bonds, which can readily be reversibly protonated to form hydride-bridged species.³⁹⁵ The Fe–Fe bonds in [CpFe(CO)(PR₃)₂](μ-CO)₂ are as basic as weak amines (p*K*_b around 6), and concomitant shift of μ-CO to terminal positions occurs on protonation (eq 83).⁴⁰³ Protonation of the Fe–Fe bond in [Fe(CO)₂(PR₃)₂](μ-SR')₂ occurs in preference to protonation of the sulfur ligands (eq 84).⁴⁰⁴ These are Fe^I centers, and

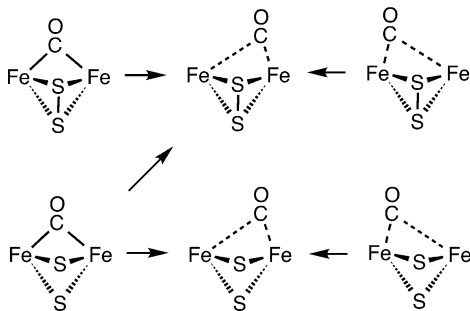


the Fe^I oxidation state has been proposed to occur in some forms of H₂ase metal cores. The basicities of M–M bonds such as in [CpRu(CO)₂]₂ are substantially higher than that of the metal sites in related 18e mononuclear complexes and are highly sensitive to the nature of the ancillary ligands.³⁹⁵ As discussed above, theoretical studies of [NiFe] hydrogenase mechanisms indicate that Fe(μ-H)Ni intermediates are energetically favorable and might also be expected to play a role in the [FeFe] H₂ases. Formation of a terminal hydride species is a possible intermediate in these M–M bond protonation processes. As shown in Scheme 21, hydride ligands could reversibly shift between bridging and terminal positions and be protonated to a readily dissociable H₂ ligand,

Scheme 22



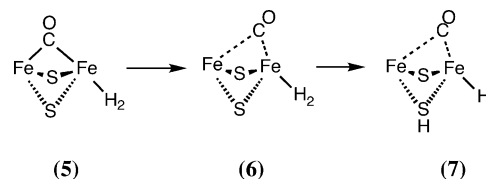
Scheme 23



leading to a cyclic process for either H_2 consumption or production. Indeed, the first examples of protonation of asymmetric iron hydrogenase active site mimics to form bridging hydride complexes via intermediacy of terminal hydrides and related studies were recently reported (Scheme 22).⁴⁰⁵ NMR evidence showed that protonation of the carbonyl–diphosphine complex at 203 K gave slow formation of a terminal hydride complex that isomerized to the μ -H complex on warming.^{405a} In the process, the diphosphine ($\text{Ph}_2\text{PC}_2\text{H}_4\text{PPh}_2$) shifted to a basal–basal position. Protonation at 233 K gave evidence for a species with hydride bound to the phosphine-containing iron as well. Protonation of a bis-carbene analogue also showed spectroscopic evidence for the initial presence of terminal hydrides.^{405b} A symmetric analogue of the complex in Scheme 22 with 2CO and PMe_3 on each Fe and containing an NR group in the middle of the bridge linking the sulfides instead of CH_2 showed that protonation at the metal bond to form a bridging hydride was thermodynamically more favorable than at the nitrogen (kinetically favored).^{405d} The synthesis of the diferrous terminal hydride complex $[\text{Fe}(\text{H})(\text{PMe}_3)_2(\mu\text{-CO})\{\mu\text{-S}(\text{CH}_2)_2\text{S}\}\text{-Fe}(\text{CO})(\text{PMe}_3)_2](\text{PF}_6)$ has been recently reported; its proton NMR spectrum exhibits a signal at -4.6 ppm, which has been assigned to the terminal hydrido ligand.^{405c} The corresponding μ -hydride compound $[\text{Fe}_2\{\mu\text{-S}(\text{CH}_2)_2\text{S}\}\{\mu\text{-H}\}(\text{CO})_2(\text{PMe}_3)_4](\text{PF}_6)$ displays a signal at -20.6 ppm, which has been attributed to the bridging hydride. The structures of both of these compounds were determined crystallographically.^{405c}

Such bridging/terminal shifts involving CO as well as H would be especially likely to occur in the [Fe] H_2 ase sites, which are attached to the protein only via the 4Fe–4S cluster, versus the [NiFe] sites, which are more tightly attached via cysteine groups that also bridge the metals. DFT calculations on $[(\text{MeS})(\text{CO})(\text{CN})\text{Fe}(\mu\text{-S})_2(\mu\text{-CO})\text{Fe}(\text{CO})(\text{CN})]^\pm$ ($z = 0$ to -2) models show that the μ -CO can easily shift like a gate, where the O atom moves little but the carbon swings left or right to form *semibridging* CO ligands that are well-known in organometallic chemistry. Also, the μ -S can join via S–S

bonds, a variable not even considered above (Scheme 23).⁴⁰⁶ Remarkably, the transformations between six different isomers at three possible redox levels are virtually barrierless. The active site possesses a relatively flat potential energy surface for geometrical changes at Fe, CO, S, and bound H, which is consistent with the extremely rapid rates of H_2 production in the enzymes. H_2 weakly binds to Fe in the position of the H_2O ligand in the enzyme as in the model (5), but calculations indicate the H_2 complex is stabilized by a CO gate shift to the right (6). In the reduced states of



these models, (5^{2-}) undergoes a mechanistically significant barrierless transfer of one H atom from $\text{Fe}-\text{H}_2$ to form SH (7^{2-}).

The above CO movements and overall coordination-sphere “rotations” about the iron centers were also examined theoretically by Darensbourg.^{248b} Both this author and Rauchfuss have recently structurally characterized mixed-valent Fe(II)Fe(I) dithiolate complexes that feature semibridging CO ligands, e.g., $(\mu\text{-pdt})[\text{Fe}(\text{CO})_2(\text{PMe}_3)][\text{Fe}(\text{CO})_2(\text{IMes})]^+$ (pdt = propanedithiolate; Imes = 1,3-bis(2,4,6-trimethylphenyl)imidazol-2-ylidene)^{407a} and $[\text{Fe}_2(\text{S}_2\text{C}_2\text{H}_4)(\text{CO})_3(\text{PMe}_3)(\text{dppv})][\text{BF}_4]$ (dppv = *cis*-1,2- $\text{C}_2\text{H}_2(\text{PPh}_2)_2$).^{407b} A protected open site with structural similarity to the active site of FeFe H_2 ases for possible H_2 binding and activation was found in these complexes, and the latter complex adds CO to this site with a concomitant shift of the semibridging CO to a normal bridging position.

8.2.12. Summary of the above Relationships

The important structure/bonding/reactivity relations between the active sites of H_2 ases and organometallic systems can be summarized as follows.

(1) Octahedral Fe(II) d^6 centers are favorable for reversible molecular H_2 binding and heterolytic cleavage. The binding strength of H_2 in organometallic systems can be competitive with that for aqua ligands, depending on the electrophilicity versus nucleophilicity of the metal center.

(2) The CO ligands are presumed to be present to increase the electrophilicity of the metal center to promote reversible H_2 binding rather than irreversible formation of catalytically inactive hydride complexes. Such electron-withdrawing ligands, especially when positioned trans to the H_2 ligand, are also known to favor heterolytic cleavage of H_2 . The CO ligands can easily shift between terminal, semibridging, and bridging positions, and it is thus crucial that the exact stereochemistry of a complex or an enzyme active site is known in order to understand H_2 activation. Electrophilic metal centers are also known to disfavor binding of atmospheric dinitrogen that could inhibit H_2 activation.

(3) Cyanide ligands may be present because of their very strong ligand-field strength that helps to maintain the metal centers in a low-spin (diamagnetic) state necessary to keep the CO ligands tightly bound. It is significant that cyanide can be formed biologically along with CO, unlike organophosphines or most other strong field ligands in inorganic and organometallic complexes.

(4) The M–M bonds in H_2 ases may be present to facilitate initial protonation of the active site. Such bonds can be fairly basic (perhaps more than the proposed amino groups in the

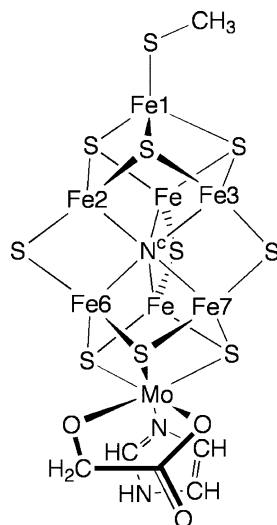


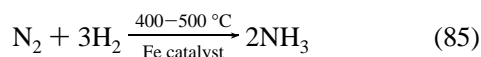
Figure 5. Model for the crystal structure of FeMo-co in *Azotobacter vinelandii*, as used in density functional calculations.⁴²¹

sulfido linker) and readily form hydride-bridged structures that are proposed to be a step in the mechanism of H₂ formation. They may also facilitate electron transfer from the site of H₂ heterolysis to the attached Fe–S cubane cluster in the Fe–Fe H₂ases.

Biomimetic production of hydrogen fuel is being intensely studied, and many of the above principles could be relevant to homogeneous catalytic cycles for formation of H₂ from protons and electrons. Splitting of water photochemically or otherwise on inexpensive first-row transition metals such as iron is ideally needed to avoid use of valuable hydrocarbons and precious metals.

9. Hydrogen Activation in Nitrogenases

Hydrogen conversion is again of prime importance in nitrogen fixation to ammonia by nitrogenase enzymes (N₂-ases)⁴⁰⁸ and can be at least partially understood in terms of inorganic chemistry. Massive research efforts^{241,242,244,336,409–425} have been directed at determination and modeling of the structure and function of N₂ase, rationalized partially on improving or providing alternate methods of ammonia production. Hundreds of million of tons of NH₃ are produced annually worldwide by the Haber process (eq 85).



Although the industrial catalyst is iron based, its chemistry is not comparable to that in biological systems, since it takes place under very high pressures and temperatures. Nitrogenase catalyzes this under much milder conditions, but the mechanism is still enigmatic. The structure of the iron–molybdenum cofactor (FeMo-co) that is the site of catalysis is a NFe₇MoS₉(homocitrate) cluster linked to the protein through a cysteine residue.⁴¹⁰ A model of the site simplified for computational analysis⁴²¹ is shown in Figure 5. There is an unusual central trigonal prism of six iron atoms (Fe2–Fe7) linked by three doubly bridging sulfur atoms and centered by a small atom, initially speculated to be nitrogen (Nc), although there is more recent spectroscopic evidence⁴¹⁹ that it is not nitrogen. Recent biochemical investigations have provided strong evidence that the Fe4 face of FeMo-co involving atoms Fe2, Fe3, Fe6, and Fe7 is where alkynes

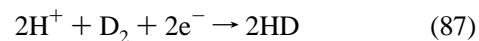
and alkenes are bound, implicating this as the site of dinitrogen activation.

Nitrogenase uses electrons and protons to hydrogenate N₂, requiring careful chemical control to direct electrons and protons toward difficult to reduce N₂ while avoiding the facile combination of electrons and protons to form H₂. There is always some diversion to form H₂ (obligatory hydrogen evolution), however. At least one H₂ is produced for every N₂ reduced, seemingly as a waste of reducing equivalents.



There are extensive studies concerning the hydrogen reactivity of nitrogenase, much of which was developed by Thorneley and Lowe from their detailed kinetic data.⁴²⁴ Their scheme involved eight stages of linked electron- and proton-transfer processes, and the earlier stages of reduction are the more intriguing, involving the accumulation of H atoms on FeMo-co, the evolution of H₂, and the initial binding of N₂. There are equilibria involving interchange of N₂ with H₂, reflecting the fact that H₂ is a competitive inhibitor of the reduction of N₂.

Insight into the nature of the intermediates comes from kinetic analysis of the HD formation reaction of nitrogenase, i.e., the N₂-dependent formation of HD in the presence of D₂.^{334,408a,423,424} When nitrogenase turns over under D₂, HD is formed, but only in the presence of N₂: other substrates such as acetylene do not enable the formation of HD. The HD formation is not catalyzed H₂/D₂ exchange but is a reduction, with the stoichiometry shown in eq 87.



Furthermore, during turnover under HD, D₂ is not formed, and when T₂ is used, there is negligible leakage of tritium label into the aqueous phase. This phenomenon implies that the H and D that form HD come from different sources that do not mix their hydrogen atoms, and that this reaction is facilitated only when N₂ is bound. This in turn implies that the displacement of H₂ by N₂ at a single active site must be an associative process. A reasonable explanation for this is that N₂ binds to a trihydride species, MH₃ or MH(H₂), with displacement of H₂. Subsequent loss of N₂ by reaction with protons toward NH₃ formation or by dissociation, followed by binding of D₂, would generate MHD₂, an obvious source of HD (section 8.2.8). The Lowe–Thorneley⁴²⁴ model of the nitrogenase mechanism is consistent with generation of a trihydride species by protons binding to the reduced site prior to N₂ binding. Some H₂ is released during this process, as in labile H₂ complexes that readily exchange N₂ and H₂. Hughes et al. propose a scheme for H₂ evolution, H₂ binding, and reduction at the Mo site of the enzyme wherein a Mo dihydride species eliminates H₂ on reaction with N₂.⁴²⁵ However, this model does not explain why, in the comparable experiment performed under HD, no D₂ forms, nor why substrates other than N₂ do not promote HD formation. Also, if H₂ can interact with the active site, why is a substrate of any kind needed to promote HD formation? Displacement of H₂ is not a necessity for binding N₂, but why does HD form only when N₂ is being reduced? One simple answer proposed by Hellen et al. is that HD formation and N₂ binding occur at different places.⁴²³ It is possible that different substrates bind to and are transformed at different parts of the large FeMo-co site of N₂ases (a separate P-cluster may also be involved). CO inhibits nitrogen fixation in N₂ases

but *not* H_2 evolution. A single site that binds H_2 and N_2 equivalently should be poisoned by CO for both H_2 and N_2 activity, and evidence increasingly points to multisite processes in the FeMoco cluster.

ENDOR spectroscopy showed that the cofactor covalently bound two chemically equivalent $H^{+/-}$, giving the first experimental insights into the structure of an intermediate formed during H_2 evolution catalyzed by N_2ase .⁴²⁰ A species with three hydrogenic species bound to one Fe was considered as a model for an intermediate state, and a dihydrogen-hydride structure was considered. However, the ENDOR patterns showed two 1H that appeared to be chemically equivalent, which would seem to be inconsistent with a $FeH-(H_2)$ structure (although this possibility was not precluded in other reaction steps or reduction states). At about the same time (2005), the coordination chemistry of H_2 on FeMo-co was examined computationally by Dance, who found that molecular H_2 coordination at iron is energetically and mechanistically reasonable. Key principles, some of which are summarized below, were derived for the coordination chemistry of hydrogenated FeMo-co modeled as in Figure 5.⁴²¹

(1) Both $Fe-H$ and molecular $Fe-H_2$ coordination can occur in exo- and endo-coordination positions at the central Fe atoms, with exo-coordination energetically better.

(2) FeMo-co has ample capacity to bind multiple H atoms and/or H_2 molecules. Two H_2 molecules can be bound to the one Fe atom if either the $Fe-N_c$ bond or the $Fe-(\mu_3-S)$ bond is severed.

(3) FeMo-co is able to distort substantially to accommodate binding of H and H_2 , but is subject to coordinative allosteric influences.

(4) $S-H$ to $Fe-H$ transfers have barriers of 9–16 kcal/mol.

(5) Association of H_2 at Fe is generally endergonic, but the presence of endo- Fe_6-H causes exo- Fe_6-H_2 association to be exergonic.

(6) Barriers for dissociation of $Fe-H_2$ are generally ca. 5 kcal/mol.

(7) One very favorable process for generation of H_2 is formation of exo- $Fe-H_2$ by transfer from proximal sulfides: the reaction is strongly exergonic, and the barrier is as low as ca. 3 kcal/mol.

(8) H atoms in endo and exo positions on the same Fe atom convert exergonically to H_2 in the exo position, with small (ca. 3 kcal/mol) barriers.

(9) Nondissociative atom exchange between H and H_2 can occur readily at one Fe site.

Clearly, the above features have similarities to established H_2 coordination chemistry, but whether H_2 ligation is important mechanistically remains open to debate.

10. Biomimetic Hydrogen Production

Production of H_2 fuel, e.g., from water via solar energy, is of high interest.^{263,426} Catalysis may involve H_2 complexes at least as intermediates, and, e.g., H_2 complexes have been implicated in solar energy conversion schemes based on photoreduction of water.²⁶⁴ Industrially important water gas shift and related H_2 -producing reactions undoubtedly proceed via transient H_2 complexes.¹⁴¹ DuBois and co-workers have found that dicationic nickel(II) complexes with two pendant amine ligands similar to that in Scheme 13 heterolyzed H_2 to form two protonated amines and were highly efficient electrocatalysts for both H_2 evolution and oxidation.²⁷⁰

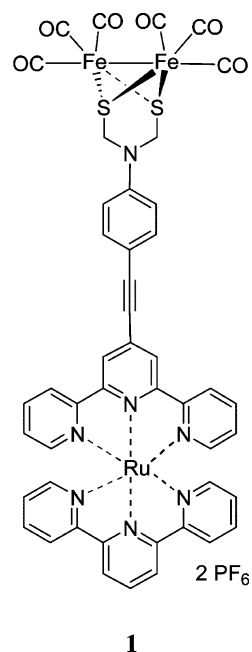


Figure 6. Schematic representation of light-driven proton reduction.²⁶²

Importantly, cooperative interactions of dihydrogen ligands with both the metal center and proton relays incorporated in the second coordination sphere contribute to the high activity observed for these Ni-based molecular catalysts that rival H_2ases in reaction rates. Electrochemical production of H_2 occurred at turnover frequencies as high as $350\ s^{-1}$, comparable to that of Ni-Fe H_2ases ($700\ s^{-1}$).^{270b}

Biomimetic H_2 production, particularly solar driven (photocatalysis), is taking cues from modeling of the active sites of hydrogenases coupled with models of Nature's photosystems.^{234,262,263,427} Here, the formation of $H-H$ bonds from protons and electrons, the microscopic reverse of H_2 heterolysis, will be crucial in leading to formation of H_2 and is very rapid at the Fe sites in H_2ases . Coupling model catalysts with photochemical water splitting will require fine-tuning of electrochemical potentials for tandem catalysis schemes. Homogeneous catalysts are advantageous, and studies are underway in this arena, e.g., by Sun et al. in their work on linking ruthenium photoreceptor complexes to dimetallic iron complexes modeling the H_2ase active site (Figure 6).^{262,427} Electrons photochemically generated from the Ru-bipy complex were designed to travel down a "molecular wire" linker to a di-iron center for combination with protons to form H_2 . However, the electrochemical potentials for the processes (photochemical production of electrons and proton reduction) must be compatible, which was a barrier to progress in the Sun system. A more promising alternative process has recently been described using a triad reaction system with a stronger reductant, $Ru(bipy)_3^+$.⁴²⁷

It should also be possible to study similar monometallic iron(II) complexes with octahedral geometry with CO, CN^- , and thiolate ligands as the site of H_2 production. One key to designing such functional catalysts for hydrogen formation via, for example, water splitting is having a proper electrochemical reduction/oxidation (redox) potential for the sequential electron addition steps. In the bimetallic model complexes for H_2ases , catalysis by the diiron units is quite sensitive to electronic effects; that is, the nature of the ligands controls the electrochemical potentials for oxidation/reduction (as in most metal complexes).²³⁵ This could partially explain

why cyanide (CN) ligands are used by hydrogenases. It is a strong electron donor (synthesizable biochemically) and would increase the electron-richness of the metal, which would facilitate protonation of the metal and lower the potential required for the electrochemical production of hydrogen. Interestingly, Sun's biomimetic dimetallic iron system did not have CN ligands and the potential was too high.²⁶² On the other hand, the metal center cannot be too electron-rich and must retain some electrophilic character; otherwise, release of hydrogen would not be facile, and a fine balance of electronics is needed. Thus, variation of ligands would be used to adjust the potentials, and there are well-defined parameters to predict this computationally by assuming a structure for the complex and applying an additive ligand parameter via the methodology developed by Lever.⁴²⁸ In addition to ligand effects, overall charge, i.e., cationic ([L-M-H₂]⁺) or anionic ([L-M-H₂]⁻), has a powerful effect on the binding and reactivity of H₂ ligands as well as electrochemistry. The nature of the metal is, of course, critical, and Rauchfuss found that platinum-group metal (e.g., Ru) mimics of the Fe-only hydrogenase active sites yield catalysts *less* effective for proton reduction, although many aspects of the associated reactivity are quite analogous.⁴²⁹ Thus, there are many factors and options for exploring homogeneous catalysts for biomimetic H₂ production (as can be seen in other articles in this thematic issue), and the work is still in its infancy.

11. H₂ Coordination Chemistry Relevant to Hydrogen Storage

11.1. Introduction

In addition to enzymatic hydrogen activation and biomimetic hydrogen production, the nature of dihydrogen coordination on metal complexes and other compounds is relevant to possible new materials for *hydrogen storage*. The reversible binding of H₂ to metal complexes and the low energies for hydrogen uptake and release as H₂ gas under near ambient conditions are ideal properties for hydrogen storage. Importantly, there would be little heat released on hydrogen uptake at a fueling station and little heat needed to release the weakly held H₂ molecules from the storage vessel. This may be the most important feature of utilizing molecular hydrogen binding for hydrogen storage. The binding energy of hydrogen molecules to stable transition metal complexes was determined to be 15–20 kcal/mol and may be as low as a few kcal/mol for the weakly bound systems under pressure. On the other hand, metal *hydrides* such as NaAlH₄ may have M–H bond energies as high as 60 kcal/mol, a potential waste of energy. However, intermediate interactions are also known in elongated H₂ complexes and in certain intermetallic rare-earth hydrides^{71–73} where d_{HH} is ~ 1.5 Å, indicating additional avenues may exist in the gray area between dihydrogen and dihydride complexes. Also, multimetallic hydrides (often clusters with μ -H) are known to dissociate and re-add H₂ reversibly.^{45c}

Materials that bind H₂ in the realm between physisorption and chemisorption are thus desirable, but there are severe challenges here. The main obstacle to overcome is the low gravimetric content of hydrogen (typically less than 1% in known complexes and 6% or greater is needed) because of the relatively high molecular weight of coligands. Only a few metal complexes are known to contain two H₂ ligands (none with more). The best known and most studied are

RuH₂(H₂)₂(PR₃)₂ [R = cyclohexyl (Cy) and cyclopentyl (Cyp)] and RuHX(H₂)₂(PCy₃)₂ (X = Cl, I), as shown in Figure 2 and eq 3.^{35,57,58,65} Up to ten hydrogens (including hydrogens from the phosphines) can be *reversibly* removed from the former (R = Cyp) under mild conditions.^{65b} Although this represents only 1.71% of the weight of the complex, this demonstrates that H₂ binding to transition metal centers could be useful in hydrogen storage materials, *particularly if the metals are incorporated into nanoporous materials*, as will be discussed below. Limiting the number of “heavy ligands” (e.g., phosphine) on the metal would obviously be beneficial. Computational studies reviewed by Heben in this thematic issue⁴³⁰ indicate that even complexes of the type M(H)_x(H₂)_n containing multiple H₂ ligands (up to $n = 6$) could be thermodynamically stable, even devoid of ligands other than hydrogen. Although multi-dihydrogen species with few or no ancillary ligands such as Cr(H₂)₆ and UH₄(H₂)₆ have been theoretically calculated to be stable,⁴³¹ they would undoubtedly be highly reactive. Such species might be stabilized when imbedded in nanoporous media, however. Although the uranium species would clearly not be a practical storage material, the calculation suggests that up to 16 H's could surround a single metal center. A buckyball can theoretically bind up to 12 metals on all of its faces (and thus up to 48 H₂),^{430,432} but again, synthesis of such species would be problematic. Nonetheless, design of such hydrogen-rich metal species is one area for exploration. As will be discussed below, unsaturated “naked” transition metal cations capable of binding multiple H₂ and/or hydride ligands may be able to be generated, since species such as [M(H₂)_n]⁺ are known in the gas phase with up to *ten* H₂ molecules “solvating” a first-row transition metal cation.^{433–449} Protonation of anionic metal polyhydrides is another possible pathway to such poly-H₂ complexes with few or no coligands, which may be stable under moderate H₂ pressures. As shown in Table 2, there are many metal–H₂ complexes with minimal or lightweight coligands. Although nearly all are unstable at room temperature, there may be means to stabilize such systems, as discussed below.

The binding of H₂ would be expected to be highly reversible in the above systems, which would be ideal for facile hydrogen storage. The above theoretically accessible multi-H₂ species would likely be unstable in the condensed phase, but they and complexes such as those in Table 2 could possibly be incorporated into nanoporous materials such as zeolites or fullerenes. As will be discussed below, metal organic framework compounds (MOFs) with very high surface areas are known to bind large numbers of H₂ primarily via physisorption within the open lattice (Figure 7). Inelastic neutron scattering spectroscopic measurements are valuable here to differentiate between the latter type of binding and coordination of H₂ to the metal, which will also be described below. Reversible binding of H₂ to main group compounds and nonmetal centers, e.g., oxides, will also be discussed. The structure and bonding properties of dihydrogen are important, and H₂ can behave as either a weak Lewis base or a weak Lewis acid toward main group compounds (Figure 8). This versatile, amphoteric-like behavior may be able to be exploited for facile reversible storage of hydrogen as molecular H₂ rather than chemical hydrides. The inability of main group compounds to backdonate electrons to H₂ σ^* (section 3.2) ensures that the H₂ is bound molecularly and reversibly rather than as a hydride, but as a result, the interaction is weak.

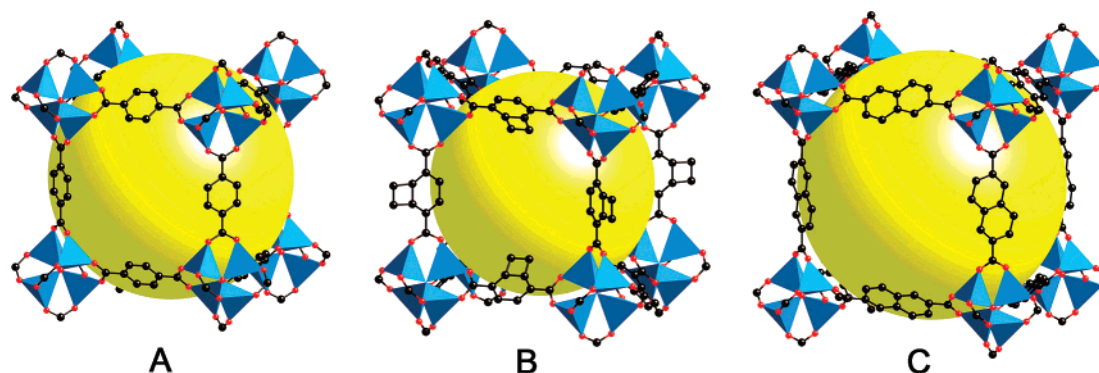


Figure 7. Single-crystal X-ray structures of MOF-5 (A), IRMOF-6 (B), and IRMOF-8 (C) illustrated for a single cube fragment of their respective cubic three-dimensional extended structures. On each of the corners is a cluster $[\text{OZn}_4(\text{CO}_2)_6]$ of an oxygen-centered Zn_4 tetrahedron that is bridged by six carboxylates of an organic linker. The large spheres represent the largest sphere that would fit in the cavities without touching the van der Waals atoms of the frameworks. Hydrogen atoms have been omitted. From ref 510 (<http://www.sciencemag.org>). Reprinted with permission from AAAS.

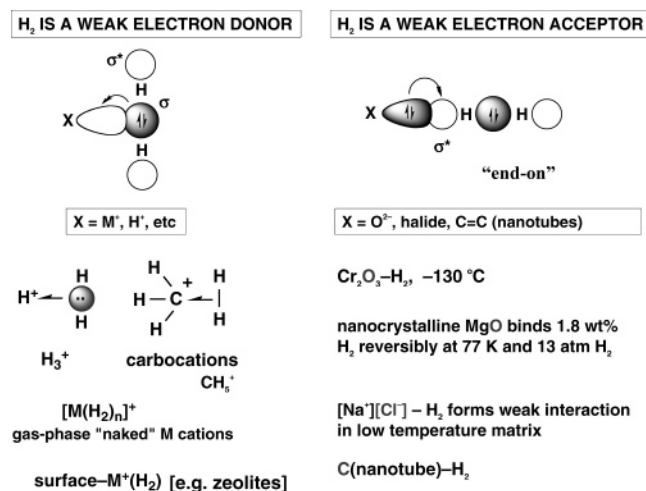


Figure 8. Examples of the ability of H_2 to behave either as a weak Lewis base or as a weak Lewis acid toward main group compounds.

Summarizing, the key advantages of molecular hydrogen binding for H_2 storage in vehicle tanks are as follows:

(1) The reversible binding of dihydrogen on a solid material would use only moderate pressure swings to fill the tank and release hydrogen. The H_2 could be added rapidly; that is, there is a small kinetic barrier for H_2 on/off and no need for catalysts or chemical conversions.

(2) Minimal heat is released on fueling the tank or is needed for hydrogen release from the tank.

(3) Inexpensive materials can be designed to bind hydrogen.

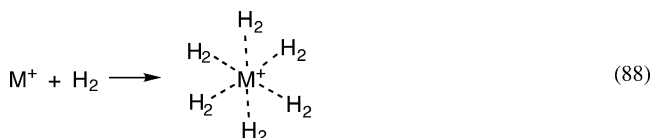
(4) The density of hydrogen bonded to solid materials may be greater than that of liquid hydrogen.

(5) Although pressure may be needed to fill the storage vessel, the pressure of the solid-bound hydrogen will not be anywhere near as high as that of liquid hydrogen or pure pressurized gas.

11.2. H_2 Binding to Naked Metal Ions

Significant theoretical and experimental investigations of molecular H_2 binding to metals have also been devoted to systems other than discrete transition metal complexes and rudimentary species such as $\text{Pd}-\text{H}_2$. A large class of "naked" metal cations, $[\text{M}(\text{H}_2)_n]^+$ (M = first row transition metal) studied by ion-beam and mass spectrometric techniques, give H_2 dissociation energies and are excellent systems for H_2

and alkane binding because of their high electrophilicity and reluctance to oxidatively add these molecules.⁴³³ These species are formed and studied, for example, by electron-impact ionization of organometallic precursors such as $\text{CpCo}(\text{CO})_2$, injection of the resulting Co^+ into a reaction cell containing H_2 , and mass spectrometric analysis. Alternately, "naked" metal ions can be produced by sputtering them off a metal cathode in a flow tube where H_2 molecules (or other small molecules) are added downstream in a guided ion-beam tandem mass spectrometer. These experiments are useful for determining $\text{M}-\text{H}_2$ binding energies on extremely electrophilic fragments. Neutral M on surfaces nearly always transfers electrons to approaching H_2 molecules to split the $\text{H}-\text{H}$ bond to give hydrides, analogous to excessive backdonation (BD) causing oxidative addition in metal complexes (Scheme 3). However, when H_2 approaches a bare M^+ , the BD bonding component is less energetically favorable because the second ionization potential of M^+ is quite high. Instead, the cation polarizes the H_2 and the M^+-H_2 bonding takes on a dipole character. Calculations indicate that M^+ can in essence be "solvated" sequentially by up to ten H_2 molecules, as in eq 88.⁴³³



Binding energies for all first-row clusters $[\text{M}(\text{H}_2)_n]^+$ ($n = 1-6$) and several small molecule analogues have been determined by temperature-dependent equilibrium measurements⁴³⁴⁻⁴⁴³ of mass-selected M^+ ions reacting with H_2 or by collision-induced dissociation (CID) in a guided ion-beam mass spectrometer (Table 5).⁴⁴⁴⁻⁴⁴⁶ Although noncovalent electrostatic interactions (charge-induced dipole and charge quadrupole) are present, they normally comprise a small fraction of the total bond strength because the purely electrostatic attraction in $[\text{Na}(\text{H}_2)_{1,2}]^+$ and $[\text{K}(\text{H}_2)_{1,2}]^+$ is only 1.3–2.5 kcal/mol.^{439,443} The presence of covalent forces in the bonding is shown by the strong influence of the nature of M^+ on both bonding energies and structures. The four covalent forces include the main interaction: electron donation from the H_2 σ orbital to M^+ that stabilizes the ion charge. Most of this donation is to the M 4s orbital with a minor amount to a 3d orbital of proper symmetry. Second, some BD to the H σ^* orbital still occurs in the later M^+

Table 5. Comparison of Experimental Binding Energies (± 0.4 – 1.4 kcal/mol) for $[M(L)_{n-1}]^+ + L \rightarrow [M(L)_n]^+$ for $L = H_2, CH_4,$ and N_2 up to $n = 4$

ion	L	binding energy			
		$n = 1$	$n = 2$	$n = 3$	$n = 4$
$[Ti(L)_n]^+$	H ₂	10.0	9.7	9.3	8.5
$[V(L)_n]^+$	H ₂	10.2	10.7	8.8	9.0
$[Cr(L)_n]^+$	H ₂	7.6	9.0	4.7	3.4
$[Mn(L)_n]^+$	H ₂	1.90	1.65	1.4	1.2
$[Fe(L)_n]^+$	H ₂	16.5	15.7	7.5	8.6
	CH ₄	13.6	23.2	23.7	17.7
	N ₂	12.9	19.8	10.8	13.6

Table 6. Interaction of H₂ with Neutral and Charged Fe

system	d_{FeH} , Å	d_{HH} , Å	binding energy, kcal/mol
$[FeH_2]^+$	1.92	0.73	−33.8
$[FeH_2]^0$	2.01	0.77	−5.0
$[FeH_2]^-$	2.25	0.86	−42.4

with filled 3d orbitals, despite the highly electron-deficient M here. In ions with half-filled 3d σ orbitals, a hybridization between the 3d_{z²} and the 4s orbital reduces on-axis Pauli repulsion. Last, minor contributions from hybridization with the 4p orbitals can occur, despite their significantly higher energy. The relative importance of these and the electrostatic factors depends strongly on the valence configuration of M⁺.

The observed binding energies for $[M(H_2)_n]^+$ as well as CH₄ analogues for comparison generally decrease with n , as shown in Table 5, which lists energies for $[M(L)_n]^+$ for $n = 1$ – 4 and L occupying octahedral sites. Computations show good agreement; that is, in $[Ti(H_2)_n]^+$, the bond energies at the DFT level are less than 1 kcal/mol lower than experimental values.⁴⁴⁰ In general, d_{HH} is near that in free H₂, 0.74–0.77 Å, for $n = 1$ – 6 , although in some cases the distance can approach the 0.82 Å value seen crystallographically in organometallic complexes. For Sc⁺, oxidative addition of H₂ to form two hydride ligands occurs for $n = 1$, followed by molecular H₂ binding to give $[ScH_2(H_2)_n]^+$.⁴³⁸ The bond strengths for $[M(H_2)_n]^+$ are greater for the later metals (Fe, Co, Ni) primarily because of greater BD and, secondarily, smaller ion size (much of the attraction is due to charge-induced dipole potential, which varies as $1/r^4$). The binding energies for Mn and Zn are by far the weakest because of repulsion between the singly occupied 4s orbital and the H₂ σ orbital.^{441–443} All other first-row metals, in contrast, have a 3d^{*n*} valence electron configuration for the $[M(H_2)_n]^+$ species.

CID measurements for CH₄ binding to $[Co(CH_4)_n]^+$ exhibit parallel behavior to that for $[Co(H_2)_n]^+$ (Table 5).^{447,448} Ab initio calculations show similar bond energies and predict that CH₄ binds in an η^2 -H,H fashion. The trend in bond energies is rationalized by electronic changes at M (e.g. s–d hybridization) on coordination of the third and successive molecules. The different trends for the Fe⁺ system for L binding are ascribed to changes in the electronic structure of M with sequential coordination of ligands of varying field strengths.⁴⁴⁹

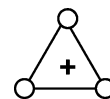
Calculations on the interaction of H₂ with Fe⁰, Fe⁺, and Fe[−] atoms show that positive charge on M favors η^2 -H₂ binding while negative charge promotes OA to dihydride (Table 6).⁴⁵⁰ This corresponds well with organometallic systems where positive charge favors η^2 -H₂ coordination. The H₂ binding energy for the positively charged molecule is much greater than that for the neutral species. An energy

barrier of 35 kcal/mol for H₂ OA on Fe⁰ is calculated, but excitation to a quintuplet 3d⁶4s¹4p¹ state leads to OA without a barrier, as is experimentally known. This large dependence on electronic state may relate to that for FeH₂(CO)₄, where H₂ is bound in dihydride form rather than as dihydrogen, which would have been expected because of the electron-poor metal center (section 3.2). Other calculations reiterate that metal cations bind H₂ with rather large binding energies while neutral metal atoms cleave H₂.^{451–454} For neutral atoms, the hydridic binding results from transfer of charge to the hydrogens that limits the number of H atoms that can subsequently be bound. However, in the cations, the binding is due to polarization of the H₂ molecule, and a large number of H₂ molecules can bind.

11.3. Interaction of H₂ with Metal Surfaces, Metal Oxides and Hydrides, and Non-transition-Metal Compounds

While the above ion species have been frequently observed spectroscopically, definitive observation of molecular binding of H₂ to metal surfaces and small metal clusters is both rare and nontrivial experimentally. Chemisorbed H₂ is observed on a stepped Ni(510) surface,¹⁹⁵ and calculations for H₂ on a Ni₁₃ cluster,⁴⁵⁵ triatomic NiH₂,⁴⁵⁵ and a Ni(100) surface⁴⁵⁶ indicate such molecularly bound states are possible, as well as hydride states. For H₂ on Ni₁₃, d_{HH} is 0.89 Å and $\nu(HH)$ is 2600 cm^{−1}, but no η^2 -H₂ state is found on Cu(100) because of differences in 3d orbital occupation. Evidence for Cu₂H₂-(H₂)_x ($x = 1, 2$) and Cu₃(H₂) in an Ar matrix exists however,⁴⁵⁷ and it should be noted that CuCl–H₂ is also known in an Ar matrix, as shown in Table 2. This table also lists other known low-temperature stable complexes with minimal or no coligands as well as surface-bound H₂ species.

Weak Lewis acid–base interactions of H₂ with main group compounds as shown in Figure 8 are known but are usually unstable and often studied only theoretically. Calculations predict H₂ binding to several types of Lewis acidic sites, including non-transition-metal cations and ionic solids such as BeO.^{458–467} The simplest such species is H₃⁺, a well-known but unstable species that is formed by protonation of H₂ and has a triangular structure with $d_{HH} = 0.87$ Å. Similar

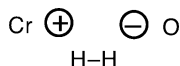


species are formed with M⁺ with all outer electrons removed and include Li(H₂)⁺ and Be(H₂)²⁺.⁴⁵⁸ Be(H₂)²⁺ is much more stable than the Li complex because Be²⁺ can accommodate two electrons in degenerate $n = 2$ empty orbitals, and the energy of these LUMOs (lowest unoccupied molecular orbitals) lies closer to the energy of the occupied σ_g H₂ orbital. This extends to neutral complexes involving light metal atoms such as OBe(H₂) and SBe(H₂)^{461–465} or F₂Mg-(H₂)⁴⁶⁶ and its dimer,⁴⁶⁷ where the “effective” positive charge on the M atom must be significant, e.g., metals with electronegative substituents such as O or F. Calculations⁴⁶⁵ show that monomeric BeO is a substantially stronger Lewis acid than AlCl₃ (BeO is actually a polymeric solid like alumina).

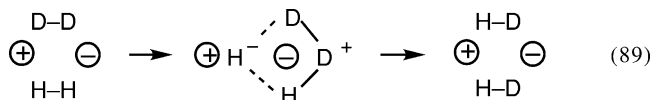
Transition metal oxides are vital heterogeneous catalysts and/or supports in many processes involving H₂ such as hydrotreatment of crude oils. Oxides studied theoretically include hematite (Fe₂O₃), modeled as a simple Fe(μ -O)₃Fe

cluster with H₂ binding to an apical Fe.⁴⁶⁸ The binding energy for (Fe₂O₃)(H₂) is calculated to be relatively high, 37.6 kcal/mol, with $d_{\text{HH}} = 0.80 \text{ \AA}$, but placing a negative charge on the cluster decreases it to -10.1 kcal/mol and d_{HH} to 0.75 \AA . This is unlike the situation for Fe atoms above (Table 6) because the negative charge on [(Fe₂O₃)(H₂)]⁻ resides mainly on oxygen, reducing the Lewis acidity of Fe without increasing the BD that activates H₂ toward OA on Fe atoms. DFT studies of the reaction surface of FeO⁺ + H₂ show η^2 -H₂ on Fe with $d_{\text{HH}} = 0.77\text{--}0.81 \text{ \AA}$ depending on Fe spin state.⁴⁶⁹

Experimental counterparts for the above computations are rare because the surface of metal oxides usually does not contain exposed unsaturated metal sites. Only very recently have coordinatively unsaturated sites (cus) been identified on an oxide surface: RuO₂(110) can be seen to bind CO to Ru cus by scanning tunneling microscopy.⁴⁷⁰ RuO₂(110) has recently been found to also bind H₂ nondissociatively at 85 K ($\nu_{\text{HH}} = 2960 \text{ cm}^{-1}$).^{471,472} Calculations indicate that $d_{\text{HH}} = 0.89 \text{ \AA}$ and that the H₂ is 1.8 \AA from the Ru^{cus} atoms (cf. 0.94 \AA and 1.81 \AA , respectively⁴⁷³ in *trans*-[RuH(H₂)(Ph₂PC₂H₄PPh₂)₂]⁺). These data suggest that, as for H₂ on Ni surfaces, the binding of H₂ to Ru^{cus} is similar to that in organometallics. Dehydroxylated chromia (Cr₂O₃) had much earlier been proposed by Burwell to contain cus in 1969, and the Cr³⁺(cus) and O²⁻(cus) ion pairs chemisorb H₂ nondissociatively below $-130 \text{ }^\circ\text{C}$.^{474,475} Pulses of D₂ at -196

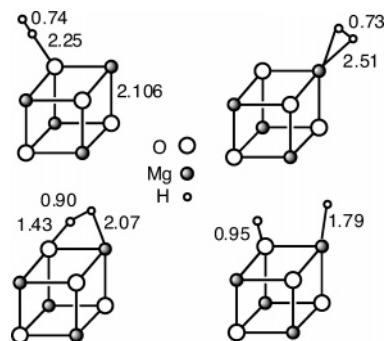


$^\circ\text{C}$ completely and rapidly displace adsorbed H₂ without formation of HD, although above $-163 \text{ }^\circ\text{C}$ substantial HD is formed. This is consistent with molecular binding of H₂ to the metal center at $-196 \text{ }^\circ\text{C}$, with *heterolytic H₂ splitting* taking place on Cr³⁺...O²⁻ sites at higher temperatures. A proposed mechanism for scrambling of H₂ + D₂ to HD involves a transient containing H⁻ associated with the Cr³⁺ and HD₂⁺ with O²⁻. A reverse situation in eq 89 with HD₂⁻



associated with Cr³⁺ and H⁺ with O²⁻ is also possible. Burwell points out that many other oxides adsorb and activate H₂ at low temperatures, including Co₃O₄, V₂O₅, MnO, and even main group oxides such as MgO.^{474,475} Calculations show that NiO weakly binds (3.7 kcal/mol) H₂ at the metal ($d_{\text{HH}} = 0.805 \text{ \AA}$)^{294,476} but ScO heterolytically cleaves H₂ to HScOH exothermically by 14 kcal/mol without forming an H₂ adduct as a local minimum on the potential energy surface.⁴⁷⁷ Computations also suggested that H₂ molecules adhere to the (111) surface of MgO with a much higher binding energy of 30 kcal/mol.⁴⁷⁸ Earlier ab initio studies of H₂ interaction and cleavage on a MgO surface using a cuboidal (MgO)₄ cluster as a model identified two types of interaction: η^1 -H₂ on the oxygen site and η^2 -H₂ at Mg.⁴⁷⁹ Because the calculated d_{HH} (0.73 \AA) in both cases is nearly the same as that for free H₂, the H₂ is most likely physisorbed. These weak complexes lead to a common transition state (TS) featuring a bridging H₂ unit with $d_{\text{HH}} = 0.90 \text{ \AA}$, followed by heterolytic cleavage of H₂ (Scheme 24). The estimated energies relative to the reactants are -2 , $+2$, and -21 kcal/mol for the physisorbed complexes, the TS, and the product. Similar results were found for an analogous

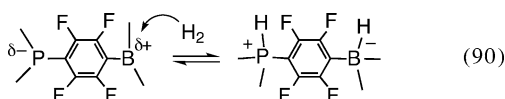
Scheme 24



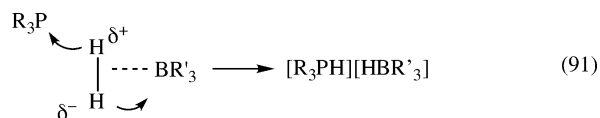
(ZnO)₄ system as a model for H₂ adsorption and heterolytic dissociation on Zn(II) zeolites.⁴⁸⁰ We have found experimentally that H₂ binds to commercial nanocrystalline MgO at 77 K and 13 atm up to 2% by weight,⁴⁸¹ although it mostly dissociates at room temperature. Using surface area = 600 m²/g and the theoretical monolayer hydrogen density of $1.3 \times 10^{-5} \text{ mol-H/m}^2$, the MgO adsorbs the equivalent of 2.5 H₂ monolayers. This indicates crevices store additional hydrogen. The enhancement storage factor of 2.5 is ~ 4 times smaller than that found in carbon, e.g., nanotubes studied by Heben and others.⁴³⁰ The light weight of MgO and similar main group oxides would make them attractive candidates for H₂ storage but probably only at low temperature.

In addition to binding of H₂ to naked metal cations, neutral hydrides can interact with H₂. Calculations show that H₂ weakly binds to a large variety of binary hydrides (MH_n),^{482,483} which have only rarely been observed, e.g., matrix-isolated CrH₂·(H₂).⁴⁸⁴ The binding energies for MH₂·(H₂) decrease with increasing atomic number for M = Ti, V, and Cr, and BD is the dominant reason. Comparisons of calculated and experimental⁴⁸⁴ vibrational frequencies support the existence of these species in matrices formed by cocondensation of M and H₂. Hydrogen exchange is calculated to occur on these systems via a trefoil-type M-H₃ transition state as in organometallic systems, which for alkali metal systems approximate ion pairs of M⁺ and H₃⁻.⁴⁸² The transition states for the exchange with group 3 transition metals have an energy of 8–10 kcal/mol relative to the reactants, which is lower than those for the alkali metal systems (16–22 kcal/mol) and group 4 metal hydrides (32–46 kcal/mol).

The metal-free aspect of most of the above systems for activation of H₂ is important because precious metals such as platinum are often used in catalysis and can be environmentally unfriendly as well as costly or in short supply. As discussed in section 8.2.5, H₂ can be cleaved at nonmetal centers, e.g., apparently on the bridging sulfides in Cp₂Mo₂S₄ that Rakowski DuBois found to react with H₂ to form SH ligands, perhaps via a four-center S₂H₂ transition state (eq 46). Metal-free hydrogenation of ketones on strong bases such as *t*-BuOK occurs under harsh conditions, apparently via base-assisted³⁵⁰ heterolysis of H₂.^{485,486} Thus, H₂ is a very weak acceptor (Lewis acid) via electron donation to its σ^* orbital and can thus interact with the O in alkoxide or metal oxides and can undergo heterolysis. Significantly, the first example of *reversible* splitting of H₂ on a *nonmetal center* has been found (eq 90).⁴⁸⁷ The phosphine borane has a strong



Lewis acidic center (boron) linked to a Lewis basic site (phosphorus). It is likely that H₂ heterolysis takes place at the electrophilic boron center where proton transfer from a transient R₃B•••H₂ complex to the basic phosphorus site occurs to form the phosphonium borate.^{487,488} Related formation of phosphonium borate salts [R₃PH][HBR'₃] from reaction of sterically demanding phosphines, boranes, and H₂ was also reported.⁴⁸⁸ Equation 91 shows a possible mechanism for the heterolyses. Theoretical and experimental



evidence indicates that H₂ can interact with a boron center. BH₃ exists computationally as a very weak Lewis acid–base complex H₂–BH₃ with a very low dissociation energy of 1–5 kcal/mol depending on methodology.^{489–491} Charge density analyses show that H₂ (and ethylene in C₂H₄–BH₃) are stronger donors than acceptors.⁴⁹¹ The barriers for hydrogen migration and rotation are very low, and the zero-point vibrational energy is similar to the binding energy so that H₂–BH₃ is barely a bound species. The dissociation energies for X₃B–H₂ (X = F, Cl) are even lower, 0.7–0.9 kcal/mol, indicative of van der Waals complexes.⁴⁹¹ Attempts to observe binding of H₂ to the latter in low-temperature matrices by Sweeney apparently led to heterolysis of H₂ to form B–H bonds.⁴⁹² A structure has been calculated for [H₃C]⁺[BH₂(H₂)⁻] and indicates interaction of H₂ with boron.⁴⁹³

Other weak interactions of H₂ with main group species shown in Table 7 help to define the Lewis acid–base strength of H₂ as a pure σ donor or acceptor. Significantly, complexes where H₂ can act only as a pure Lewis base are unstable, attesting to the vital role of BD from metal d orbitals in stabilizing σ -ligand binding. Hypervalent main group species such as CH₅⁺, CH₆²⁺, CH₇³⁺, SiH₃(H₂)₂⁺, and analogous B and Al series starting with BH₆⁺, AlH₄⁺, and AlH₆⁺ are rationalized theoretically as highly dynamic H₂ complexes of main group cations (see section 3.1). In regard to materials for hydrogen storage, some of the species in Table 7 have very high gravimetric percentages of hydrogen, e.g., LiH–(H₂)₂ (42%), but have been characterized only under low-temperature conditions and/or are unstable.

11.4. Inelastic Neutron Scattering (INS) Studies of H₂ Coordination and Rotation

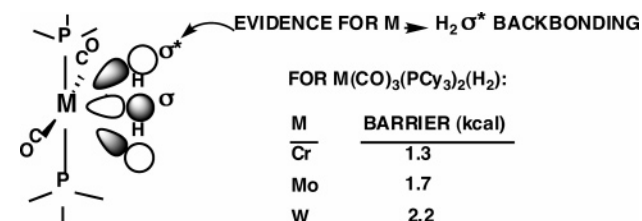
The H₂ ligand undergoes rapid two-dimensional hindered rotation about the M–H₂ axis; that is, it spins (librates) in propeller-like fashion with little or no wobbling. This phenomenon has been extensively studied by inelastic neutron scattering (INS) methods by Eckert because it *unequivocally distinguishes molecular H₂ binding from classical hydride binding*, where there is no such rotation.^{173,494–500} Furthermore, weak physisorption of H₂, e.g., van der Waals interaction with main group atoms, can be distinguished from the much stronger binding of H₂ to metal centers. This is particularly important in solid-state hydrogen-storage materials that cannot easily be studied by NMR or other conventional methods (see section 11.5). These discriminating features arise because there is always at least a small to moderate barrier to rotation, ΔE , in metal coordination brought about by M→H₂ σ^* backdonation (BD) (Scheme 3). The σ -donation from H₂ to M cannot give rise to a

Table 7. Weak Interactions of H₂ with Main Group Compounds

compound	evidence	ref
[PH ₂ (H ₂) ₂] ³⁺ , [AsH ₂ (H ₂) ₂] ³⁺	theory	<i>a</i>
[SiH ₃ (H ₂) ₂] ⁺	IR ($\nu_{\text{HH}} = 3866 \text{ cm}^{-1}$)	<i>b</i>
[SiH ₂ (H ₂) ₂] ⁺ ; [PH(H ₂) ₂] ⁺	theory	<i>c</i>
Na ⁺ /K ⁺ (H ₂) _{1,2}	surface ionization	439
MH(H ₂) ₂ (M = Li, Na, K)	solid hydrogen, theory	<i>d</i>
Al ⁺ (H ₂)	theory	451
AlH _x (H ₂); <i>x</i> = 1–3	argon matrix	<i>e, f</i>
[AlH _x] ^{<i>n</i>+} ; <i>x</i> = 4–8; <i>n</i> = 1–3	theory	<i>g</i>
AlH ₃ (H ₂)	theory	<i>h</i>
BH(H ₂)	solid argon	490
BH ₂ (H ₂)	esr, theory	<i>i, j</i>
BH ₃ (H ₂)	theory, solid argon	490 ^k
[BH ₄] ⁺ ; [BH ₇] ²⁺ ; [BH ₈] ³⁺	theory	<i>l</i>
[BH ₄ L] ⁺ ; L = NH ₃ , H ₂ O, etc.	theory	<i>m</i>
[BXH ₃] ⁺ ; [BX ₂ H ₄] ⁺ ; X = F, Cl	theory	<i>n</i>
Lewis base–H ₂	solid argon	<i>o</i>
halide–H ₂	argon matrix	<i>p–s</i>
[HnGe(H ₂) ₂] ⁺	<i>n</i> = 0, 1	theory, mass spec
[GeH ₃ (H ₂) ₂] ⁺	theory	<i>u, v</i>
BeO–H ₂ ; BeS–H ₂	theory	462–464
X ₃ B–H ₂	theory	491
MgO–H ₂	theory, experiment	478, 479, 481
C(nanotube)(η^1 -H ₂)	theory, experiment	430, 432 ^{w–z}
Li-ZSM-5-H ₂	IR ($\nu_{\text{HH}} = 4092 \text{ cm}^{-1}$)	<i>aa</i>

^a Rasul, G.; Prakash, G. K. S.; Olah, G. A. *J. Am. Chem. Soc.* **1997**, *119*, 12984. ^b Cao, Y.; Choi, J.-H.; Haas, B.-M.; Johnson, M. S.; Okumura, M. *J. Phys. Chem.* **1993**, *97*, 5215. ^c Gil, A.; Bertran, J.; Sodupe, M. *J. Am. Chem. Soc.* **2003**, *125*, 7561. ^d Wang, X.; Andrews, L. *J. Phys. Chem. A* **2007**, *111*, ASAP. ^e Chertihin, G. V.; Andrews, L. *J. Phys. Chem.* **1993**, *97*, 10295. ^f Pullumbi, P.; Bouteiller, Y.; Manceron, L. *J. Chem. Phys.* **1994**, *101*, 3610. ^g Olah, G. A.; Rasul, G. *Inorg. Chem.* **1998**, *37*, 2047. ^h Schreiner, P. R.; Schaefer, H. F., III; Schleyer, P. v. R. *J. Chem. Phys.* **1995**, *103*, 5565. ⁱ Saxon, R. P. *J. Phys. Chem.* **1993**, *97*, 9356. ^j Van Zee, R. J.; Williams, A. P.; Weltner, W., Jr. *J. Chem. Phys.* **1997**, *107*, 4756. ^k Schreiner, P. R.; Schaefer, H. F., III; Schleyer, P. v. R. *J. Chem. Phys.* **1994**, *101*, 7625. ^l Rasul, G.; Olah, G. A. *Inorg. Chem.* **1997**, *36*, 1278. ^m Rasul, G.; Prakash, G. K. S.; Olah, G. A. *Inorg. Chem.* **1999**, *38*, 1814. ⁿ Rasul, G.; Olah, G. A. *Inorg. Chem.* **2001**, *40*, 2453. ^o Moroz, A.; Sweeney, R. L.; Whittenburg, S. L. *J. Phys. Chem.* **1990**, *94*, 1352. ^p Ogden, J. S.; Rest, A. J.; Sweeney, R. L. *J. Phys. Chem.* **1995**, *99*, 8485. ^q Sweeney, R. L.; Ogden, J. S. *Inorg. Chem.* **1997**, *36*, 2523. ^r McKee, M. L.; Sweeney, R. L. *J. Phys. Chem. A* **2000**, *104*, 962. ^s Sweeney, R. L.; Vuong, L.; Bishara, J. *J. Phys. Chem. A* **2002**, *106*, 11440. ^t Jackson, P.; Sandig, N.; Diefenbach, M.; Schroder, D.; Schwarz, H.; Srinivas, R. *Chem.–Eur. J.* **2001**, *7*, 151. ^u Schreiner, P. R.; Schaefer, H. F., III; Schleyer, P. v. R. *J. Chem. Phys.* **1994**, *101*, 2141. ^v Archibong, E. F.; Leszczynski, J. *J. Phys. Chem.* **1994**, *98*, 10084. ^w Lee, E.-C.; Kim, Y.-S.; Jin, Y.-G.; Chang, K. *J. Phys. Rev. B* **2002**, *66*, 073415. ^x Han, S. S.; Lee, H. M. *Carbon* **2004**, *42*, 2169. ^y Yildirim, T.; Ciraci, S. *Phys. Rev. Lett.* **2005**, *94*, 175501. ^z Cheng, H.; Pez, G. P.; Cooper, A. C. *J. Am. Chem. Soc.* **2001**, *123*, 5845. ^{aa} Areat, C. O.; Manoilova, O. V.; Bonelli, B.; Rodriguez-Delgado, M.; Palomino, G. T. *Chem. Phys. Lett.* **2003**, *370*, 631.

Scheme 25



rotational barrier since it is completely isotropic about the M–H₂ bond. In M(CO)₃(PCy₃)₂(H₂), the barrier actually arises from the *disparity* in the BD energies from the d orbitals when H₂ is aligned parallel to P–M–P versus parallel to OC–M–CO, where BD is less (though not zero; Scheme 25).

ΔE varies with M, coligands, and other factors and can be analyzed in terms of the BD and other forces that lead to it, both computationally and by a series of experiments where

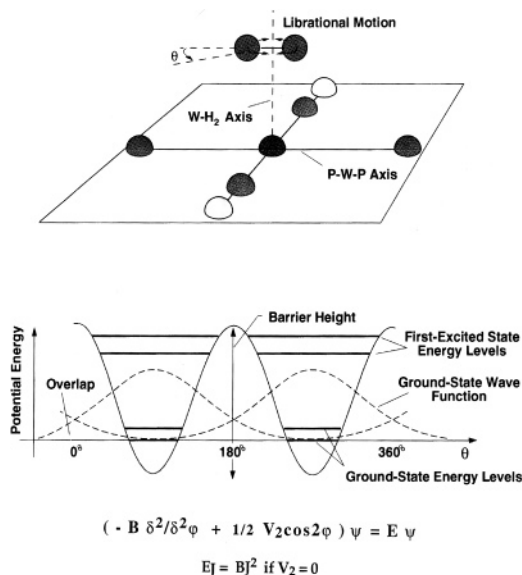


Figure 9. Model for the hindered rotation of the H₂ ligand in metal complexes. (top) Schematic of H₂ rotation in W(CO)₃(η²-H₂)P₂ about the axis from the W atom to the midpoint of the H-H bond. (bottom) Double-minimum potential V₂(φ). The transitions indicated are for W(CO)₃(H₂)(PCy₃)₂, where B is taken to be 49.5 cm⁻¹.

metal–ligand (M/L) sets are varied. In most “true” H₂ complexes with $d_{\text{HH}} < 0.9 \text{ \AA}$, the barrier is only a few kcal/mol and observable only by neutron scattering methods. It can be as low as 0.5 kcal/mol for symmetrical ligand sets, for example all cis L are the same, but has never been measured to be zero because minor geometrical distortions or crystal lattice-related effects are usually present. In the case of complexes with elongated H–H bonds or where rotation is sterically blocked as in [Cp'₂M(H₂)(L)]⁺ (M = Nb, Ta), much higher barriers of 3–12 kcal/mol are observed by INS or even solution NMR methods.^{222,501} Interactions of η²-H₂ with cis ligands can significantly lower the barriers, as was shown in section 6. The hindered rotation of η²-H₂ is thus governed by a variety of forces, which can be divided into bonded (electronic) and nonbonded interactions (“steric” effects). The direct electronic interaction between M and H₂ results from overlap of the appropriate molecular orbitals. Nonbonded interactions such as van der Waals forces between the η²-H₂ atoms and the other atoms on the molecule may vary as η²-H₂ rotates.

The geometry and height of the barrier can be derived by fitting the rotational transitions observed by INS techniques to a model for the barrier. The simplest possible model for the rotations of a dumbbell molecule is one of planar reorientation about an axis perpendicular to the midpoint of the H–H bond in a potential of twofold symmetry (Figure 9). Application of a barrier to rotation rapidly decreases the separation between the lowest two rotational levels, which may then be viewed as a split librational ground state. Transitions within this ground state as well as those to the excited librational state (often called torsions) may be observed by INS. The former occur by way of rotational tunneling,⁵⁰² since the wave functions for the H₂ in the two wells 180° apart overlap. This rotational tunneling transition has an approximately exponential dependence on the barrier height, and is therefore extremely sensitive to the latter and thus to even very minor changes in H₂ environment (e.g., crystal packing forces). It is this property that is exploited to gain information on the origin of the barrier and to easily

distinguish even small variations in H₂ binding sites in materials (section 11.5).

Both the rotational tunneling transition and the transitions to the first excited librational state can readily be observed by INS techniques.^{173,494–500,502} Neutrons are extremely well suited as probes for molecular rotations when the motion involves mainly H atoms. The INS studies allow observation of low-lying transitions within the ground librational state of the η²-H₂ (tunnel splitting), which corresponds to the para ($I = 0, J = 0$) to ortho ($I = 1, J = 1$) transition for free H₂ (120 cm⁻¹ in liquid hydrogen). INS measurements are typically carried out at ~5 K using ~1 g of polycrystalline H₂ complex sealed under inert atmosphere in aluminum or quartz sample holders. This measurement can be performed without regard to other hydrogen-containing ligands, which do not have observable excitations at low temperatures in the energy range of those of the H₂. In most cases, the ground-state rotational tunneling, as well as the two transitions to the split excited librational state, are observed. Because the tunnel splittings (typically 1–10 cm⁻¹) can be measured with much better accuracy than the librational transitions, the value for the barrier height V₂ is usually extracted from the former. Prior to the discovery of H₂ complexes, the only systems known containing hydrogen molecules were H₂ gas or H₂ that was barely affected by its surroundings (as in physisorbed H₂). The smallest splittings between the ortho and para H₂ states that had previously been observed were 4.8–10.5 cm⁻¹ for H₂ in K-intercalated graphite⁵⁰³ and 30.6 cm⁻¹ for H₂ in Co ion-exchanged NaA zeolite.⁵⁰⁴ In both of these cases, H₂ is in all likelihood physisorbed as no indication of H–H bond activation could be found. However, for the M(η²-H₂) ground librational state, splittings as high as 17 and 0.6 cm⁻¹ are observed at temperatures as high as 200 K. The signals shift to lower energy and broaden but remain visible into the quasielastic scattering region. Observation of rotational tunneling, which is a *quantum mechanical* phenomenon, at such a high temperature is extraordinary: in all previous studies of this type involving CH₃ or [NH₄]⁺, the transition to classical behavior occurs well below 100 K.

Considerable molecular level detail on the interaction and binding of H₂ with both metal centers and nonmetal substances can be obtained by inelastic neutron scattering from the hindered rotor states of the bound molecule. The transition energies between these quantum mechanical rotational states for an adsorbed hydrogen molecule are very sensitive to the shape and height of the barrier to rotation, which in turn is a reasonably direct measure of the guest–host interactions. For low to medium barrier heights (as in, for example, the MOF hydrogen storage materials discussed below), the transition between the lowest two states (rotational tunneling transition) decreases approximately exponentially with an increase of the barrier to rotation from the molecule’s chemical environment. Moreover, the very large inelastic scattering cross section of ¹H compared to that of any other atoms present in such systems makes rotational tunneling spectroscopy by INS a highly specific method to characterize the interaction between H₂ and its host.

In addition to studies of H₂ rotational motion, the low-frequency to midfrequency (200–1000 cm⁻¹) region of the neutron vibrational spectrum can be probed to investigate the nature of dihydrogen bonding. This measurement is only possible by use of a differential technique⁵⁰⁵ involving subtraction of the spectrum observed for a sample with a

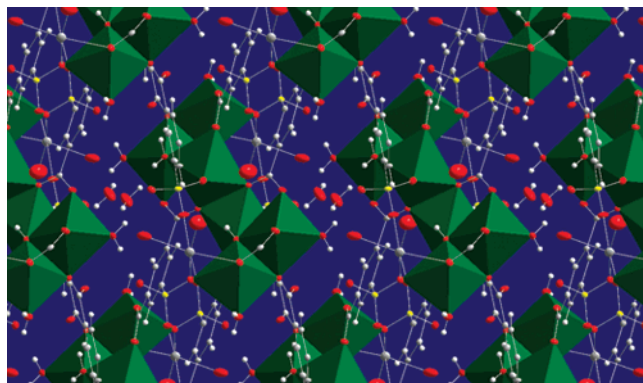


Figure 10. Crystal structure for hydrated $\text{NaNi}_3(\text{SIPA})_2(\text{OH})-(\text{H}_2\text{O})_5\cdot\text{H}_2\text{O}$, viewed in the ab plane. NiO_6 octahedra are illustrated as green polygons. Sodium, sulfur, carbon, oxygen, and hydrogen atoms are shown as blue, yellow, gray, red, and white spheres, respectively.

D_2 -ligand (or another suitable “blank”) from that of an identical sample with the H_2 ligand, which leaves only the vibrational modes for the $\text{M}-(\text{H}_2)$ fragment. For example, deformational modes in $\text{W}(\text{CO})_3(\text{PCy}_3)_2(\text{H}_2)$ have been identified by this technique (section 5). It is also useful for studies of almost any low-energy vibration involving hydrogen in the solid phase, e.g., in ammonia–borane, $\text{NH}_3\text{-BH}_3$. The latter has received a great deal of interest recently as a solid-state hydrogen storage material (“chemical hydrogen storage”), since it was discovered to release hydrogen under mild thermal conditions in the presence of acids or transition metal catalysts.⁵⁰⁶ The unique “dihydrogen bonding” interactions (see eq 53) between the adjacent protic NH and hydridic BH groups in NH_3BH_3 are important in both the dynamics of hydrogen motion and the reaction chemistry here.

11.5. Binding of H₂ to Highly Porous Solids and INS Studies

Nonmetal highly porous compounds such as carbon-based substances, e.g., fullerenes, and metal organic framework (MOF) materials have been intensely studied as possible lightweight materials for H_2 storage.^{507–523} This subject has been reviewed in this thematic issue by Heben and will not be discussed in detail except for relevance to the structure/bonding principles and methods developed for studying metal– H_2 complexes, such as neutron scattering. Techniques such as inelastic neutron scattering discussed above provide a unique tool for investigating the structure, dynamics, and chemical environment of hydrogen in potential hydrogen storage materials. This method as well as other neutron spectroscopy methods (powder and single-crystal neutron diffraction) has been applied to H_2 adsorption at low temperatures (typically 77 K) in porous carbons,⁵⁰⁷ zeolites,^{504,508,518} nickel phosphates,⁵⁰⁹ Prussian blue analogues,⁵¹⁴ and hybrid inorganic–organic compounds (e.g., MOFs),^{510–513,515–517,519–523} These methods have been described in more detail in a study of hybrid materials that will be discussed below.⁵¹⁶ IR spectroscopy has also been used, and the presence of a doublet at 4029 and 4008 cm^{-1} has been ascribed to H_2 adsorbed on available surface Zn^{2+} ions on MOF-5.⁵²⁰

An excellent recent example of the value of INS studies on H_2 –MOF interaction that will be discussed in detail is H_2 adsorbed in $\text{NaNi}_3(\text{SIPA})_2(\text{OH})(\text{H}_2\text{O})_5\cdot\text{H}_2\text{O}$, a MOF synthesized by Cheetham shown in Figure 10.⁵¹⁶ The organic linker here is 5-sulfoisophthalate (SIPA). At the lowest

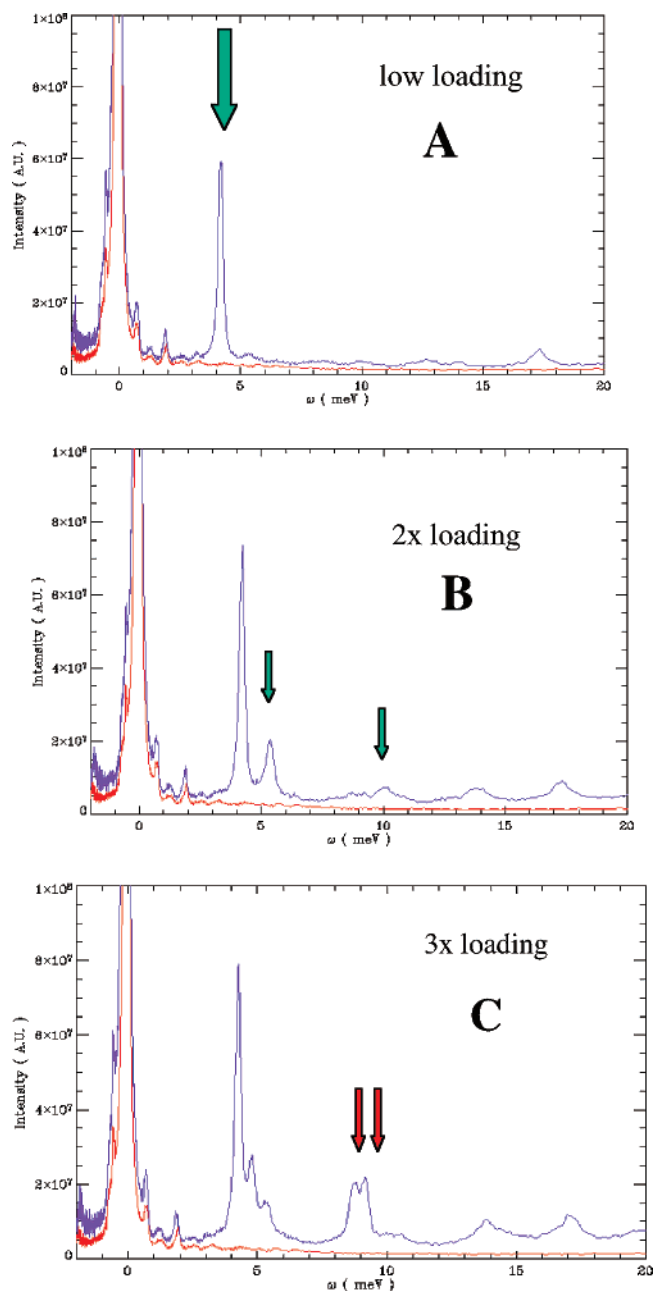


Figure 11. Inelastic neutron spectra of H_2 in $\text{NaNi}_3(\text{SIPA})_2(\text{OH})-(\text{H}_2\text{O})_5\cdot\text{H}_2\text{O}$ for different loading levels. Various loadings are shown in purple, with an unloaded measurement in red for comparison. The intensity is expressed in arbitrary units (A.U.). Several well-defined binding sites with strong guest–host interactions (much greater than carbons) or MOF-5 sites with planar rotation (green arrows in A and B) indicate peaks for chemisorbed H_2 at unsaturated Ni sites. 3-D rotation (physisorbed H_2) is seen in part C (two new peaks shown by red arrows).

loading of H_2 , a strong peak is observed in rotational tunneling spectra (Figure 11) at 4.2 meV along with a weaker peak at 17.3 meV from hindered rotational transitions of the bound H_2 molecule. This value of 4.2 meV for the energy of the lowest rotational transition (or the rotational tunnel splitting) may be compared with about 1.5 meV for H_2 in VSB-5,⁵⁰⁹ where it must be kept in mind that a lower energy indicates a larger barrier to rotation. A larger barrier to rotation may not necessarily be equivalent to stronger binding of the sorbed hydrogen, but in a general sense this seems to be the case and these results provide a good confirmation of this general trend. The INS spectra of H_2 in NiSIPA appear

to strongly suggest that binding of molecular hydrogen first occurs by molecular chemisorption at the unsaturated Ni(II) binding sites created by dehydration (Figure 11), as the series of transitions at 4.1 and 17.3 and 22 meV (not shown) cannot be assigned on the basis of a model for physisorbed H₂ (i.e., double-minimum with *two* rotational degrees of freedom) but can be fitted to the model used for coordinated dihydrogen (*planar* rotation in a double-minimum potential) with a barrier height $V/B = 3.1$, where the rotational constant B for H₂ is 7.35 meV. A second site becomes occupied when the H₂ loading is increased to twice the initial loading (Figure 11) with a set of transitions at 5.4 meV and about 10 meV that again fit to the model for planar rotation ($V/B = 1.7$) indicative of molecular chemisorption. Two additional binding sites for H₂ become evident at three times the lowest loading, another strong binding site characterized by peaks at 4.8 and 13.8 meV and a second one characterized by a doublet at 8.5 and 9.2 meV. This latter set of transitions, however, corresponds to that for a physisorbed molecule (two-dimensional reorientation) and a barrier of $3.4B$. Another site for physisorbed H₂ becomes progressively occupied at four and five times the original loading with transitions at 10.8 meV and 7 meV and a shoulder at approximately 17.2 meV, which correspond to a barrier of $2.2B$. At the highest loading ($5\times$, not shown), a peak is also observed close to the free rotor value (14.7 meV) that would suggest some agglomeration of hydrogen molecules into bulk solid particles.

The above data suggest that several accessible, coordinatively unsaturated Ni(II) sites exist in NaNi₃(SIPA)₂(OH)-(H₂O)₅·H₂O when it is dehydrated at sufficiently high temperature to remove aqua ligands from the Ni octahedra. Additional sites in the structure, where H₂ is thought to be physisorbed, bind the molecule much more strongly than do carbon supports. Remarkably detailed information has also been obtained on the primary binding sites of H₂ in a series of metal-organic frameworks composed of Zn₄O(O₂C⁻)₆ secondary building units (Figure 7) with the use of INS.^{510,511b} Five primary binding sites had been identified for gases in IRMOF-1, including three on the inorganic cluster and two solely on the phenylene link.⁵¹⁷ Each (CO₂)₃ site is surrounded trigonally by (ZnO)₂ sites at 4 Å, and so each cluster can accommodate at most 16 adsorbed molecules per formula unit. In the INS spectra, two unique 0–1 transitions for these sites, in a 1:3 intensity ratio, were expected, saturating at approximately 16 H₂ per formula unit. Aside from variance in peak positions, and possible overlap in the case of IRMOF-8, this is what was observed, and it was concluded that sites I and II for H₂ adsorption are (CO₂)₃ and (ZnO)₂. Despite their chemical similarities, the variation in INS peak positions associated with sites I and II of each MOF is significant and clearly indicates that the organic links play an active role in defining the nature of the adsorption sites for H₂. This is reasonable given the variety of links employed in these materials, which strongly affect the local structure of the Zn₄O(O₂C⁻)₆ clusters and thus the charge transfer between the Zn²⁺ and the aryl carboxylates. In contrast, features assigned to H₂ bound to primarily organic sites cover a more narrow energy range and show low barriers to rotation consistent with the weaker binding on those sites. These sites show much larger increases in INS intensity with higher H₂ loading, as their capacity for adsorption at the low temperature of these experiments is significantly higher.

Direct evidence for strong side-on H₂ binding to metal centers as in organometallic dihydrogen complexes (so-called Kubas complexes) has been obtained. Binding to exposed

Cu coordination sites has been seen by neutron diffraction and INS methods in a Cu-exchanged zeolite ZSM-5⁵¹⁸ and in the Prussian blue analogue, Cu₃[Co(CN)₆]₂.⁵¹⁴ The INS study on Cu-ZSM-5 showed H₂ rotational barriers of 1.8 and 2.1 kcal/mol, similar to those seen in metal-dihydrogen complexes, indicating side-on bonding of H₂ to Cu. This is in marked contrast to what has been observed for open Cu binding sites in MOFs or partially Cu²⁺ exchanged zeolite A.⁵²⁴

The development of such highly porous solids for reversible molecular H₂ binding in the above Ni, Cu, Zn, and other systems is a major challenge in materials science. The difficulty arises because a sufficiently strong affinity toward H₂ for room-temperature storage applications is needed, but the interaction cannot be so strong that it leads to irreversible dissociative binding, slows kinetics, or results in large energy losses associated with cycling. The MOFs and other highly porous materials containing coordinatively unsaturated metal sites are a realistic and promising means of achieving this goal. In order to bind molecular H₂, it is necessary to design compounds with high surface areas or mimic the nanotube structures of carbon fullerenes, but using much less expensive materials. There is a great opportunity for design of, for example, supramolecular cage-like structures of light main group elements such as boron, oxygen, nitrogen, lithium, etc. that would help trap molecular hydrogen. As discussed above, H₂ molecules have the ability to bind to a large variety of materials as either a Lewis acid or a Lewis base, albeit weakly, and this is the key feature to be explored for new hydrogen storage methods.

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13. References

- (1) Kubas, G. J.; Ryan, R. R.; Swanson, B. I.; Vergamini, P. J.; Wasserman, H. J. *J. Am. Chem. Soc.* **1984**, *106*, 451.
- (2) Kubas, G. J.; Unkefer, C. J.; Swanson, B. I.; Fukushima, E. *J. Am. Chem. Soc.* **1986**, *108*, 7000.
- (3) Kubas, G. J. *Acc. Chem. Res.* **1988**, *21*, 120.
- (4) Bender, B. R.; Kubas, G. J.; Jones, L. H.; Swanson, B. I.; Eckert, J.; Capps, K. B.; Hoff, C. D. *J. Am. Chem. Soc.* **1997**, *119*, 9179.
- (5) Kubas, G. J. *J. Organomet. Chem.* **2001**, *635*, 37.
- (6) Kubas, G. J. *Metal Dihydrogen and σ -Bond Complexes*; Kluwer Academic/Plenum Publishers: New York, 2001.
- (7) Kubas, G. J.; Ryan, R. R. *Polyhedron* **1986**, *5*, 473.
- (8) Baik, M.-H.; Friesner, R. A.; Parkin, G. *Polyhedron* **2004**, *23*, 2879.
- (9) Crabtree, R. H. *Angew. Chem., Int. Ed. Engl.* **1993**, *32*, 789.
- (10) James, B. R. *Homogeneous Hydrogenation*; John Wiley and Sons: New York, 1973.
- (11) Halpern, J. *Adv. Catal.* **1959**, *11*, 301.
- (12) Kubas, G. J. *Chem. Commun.* **1980**, 61.
- (13) Wasserman, H. J.; Kubas, G. J.; Ryan, R. R. *J. Am. Chem. Soc.* **1986**, *108*, 2294.
- (14) Brookhart, M.; Green, M. L. H.; Wong, L.-L. *Prog. Inorg. Chem.* **1988**, *36*, 1.
- (15) Allen, A. D.; Senoff, C. V. *J. Chem. Soc., Chem. Commun.* **1965**, 621.
- (16) Olah, G. A. *Angew. Chem., Int. Ed. Engl.* **1993**, *32*, 767.
- (17) Perutz, R. N.; Turner, J. J. *J. Am. Chem. Soc.* **1975**, *97*, 4791.
- (18) Sweany, R. L. *J. Am. Chem. Soc.* **1985**, *107*, 2374.
- (19) (a) Upmacis, R. K.; Gadd, G. E.; Poliakoff, M.; Simpson, M. B.; Turner, J. J.; Whyman, R.; Simpson, A. F. *J. Chem. Soc., Chem. Commun.* **1985**, 27. (b) Upmacis, R. K.; Poliakoff, M.; Turner, J. J. *J. Am. Chem. Soc.* **1986**, *108*, 3645.
- (20) Church, S. P.; Grevels, F.-W.; Hermann, H.; Shaffner, K. *J. Chem. Soc., Chem. Commun.* **1985**, 30.
- (21) Matthews, S. L.; Pons, V.; Heinekey, D. M. *J. Am. Chem. Soc.* **2005**, *127*, 850.

- (22) Matthews, S. L.; Heinekey, D. M. *J. Am. Chem. Soc.* **2006**, *128*, 2615.
- (23) Saillard, J.-Y.; Hoffmann, R. *J. Am. Chem. Soc.* **1984**, *106*, 2006.
- (24) Lin, Z.; Hall, M. B. *Coord. Chem. Rev.* **1994**, *135/136*, 845.
- (25) Maseras, F.; Lledós, A.; Clot, E.; Eisenstein, O. *Chem. Rev.* **2000**, *100*, 601.
- (26) Andrews, M. A.; Kirtley, S. W.; Kaesz, H. D. *Adv. Chem. Ser.* **1978**, *167*, 229.
- (27) Schubert, U. *Adv. Organomet. Chem.* **1990**, *30*, 151.
- (28) Corey, J. Y.; Braddock-Wilking, J. *Chem. Rev.* **1999**, *99*, 175.
- (29) (a) Nikonov, G. I. *Adv. Organomet. Chem.* **2005**, *217*. (b) Lin, Z. *Chem. Soc. Rev.* **2002**, *31*, 239.
- (30) Jessop, P. G.; Morris, R. H. *Coord. Chem. Rev.* **1992**, *121*, 155.
- (31) Morris, R. H. *Can. J. Chem.* **1996**, *74*, 1907.
- (32) Crabtree, R. H. *Acc. Chem. Res.* **1990**, *23*, 95.
- (33) Heinekey, D. M.; Oldham, W. J., Jr. *Chem. Rev.* **1993**, *93*, 913.
- (34) Heinekey, D. M.; Lledós, A.; Lluch, J. M. *Chem. Soc. Rev.* **2004**, *33*, 175.
- (35) Sabo-Etienne, S.; Chaudret, B. *Coord. Chem. Rev.* **1998**, *178–180*, 381.
- (36) Crabtree, R. H.; Hamilton, D. G. *J. Am. Chem. Soc.* **1986**, *108*, 3124.
- (37) Knoth, W. H. *J. Am. Chem. Soc.* **1968**, *90*, 7172.
- (38) Ashworth, T. V.; Singleton, E. *J. Chem. Soc., Chem. Commun.* **1976**, 705.
- (39) Gusev, D. G.; Vymenits, A. B.; Bakhmutov, V. I. *Inorg. Chim. Acta* **1991**, *179*, 195. In 1993 Zilm obtained solid-state ¹H NMR evidence for H₂ coordination ($d_{\text{HH}} = 0.93 \text{ \AA}$) on a sample we prepared (Wisniewski, L.; Zilm, K. W.; Kubas, G. J.; Van der Sluys, L. Unpublished results).
- (40) Esteruelas, M. A.; Oro, L. A. *Chem. Rev.* **1998**, *98*, 577.
- (41) Perutz, R. N.; Sabo-Etienne, S. *Angew. Chem., Int. Ed.* **2007**, *46*, 460.
- (42) Jia, G.; Lau, C.-P. *Coord. Chem. Rev.* **1999**, *190–192*, 83.
- (43) Esteruelas, M. A.; Oro, L. A. *Adv. Organomet. Chem.* **2001**, *47*, 1.
- (44) Kuhlman, R. *Coord. Chem. Rev.* **1997**, *167*, 205.
- (45) (a) McGrady, G. S.; Guilera, G. *Chem. Soc. Rev.* **2003**, *32*, 383. (b) Schneider, J. J. *Adv. Eng. Chem., Int. Ed. Engl.* **1996**, *35*, 1068. (c) Weller, A. S.; McIndoe, J. S. *Eur. J. Inorg. Chem.*, submitted.
- (46) Kubas, G. J. *Adv. Inorg. Chem.* **2004**, *56*, 127.
- (47) Kubas, G. J. *Catal. Lett.* **2005**, *104*, 79.
- (48) (a) *Transition Metal Hydrides*; Dedieu, A., Ed.; VCH Publishers, Inc.: New York, 1992. (b) *Recent Advances in Hydride Chemistry*; Peruzzini, M., Poli, R., Eds.; Elsevier Science B. V.: Amsterdam, 2001.
- (49) (a) Kristjansdottir, S. S.; Norton, J. R. In *Transition Metal Hydrides*; Dedieu, A., Ed.; VCH Publishers, Inc.: New York, 1992; pp 309–359. (b) Papish, E. T.; Magee, M. P.; Norton, J. R. In *Recent Advances in Hydride Chemistry*; Peruzzini, M., Poli, R., Eds.; Elsevier Science B. V.: Amsterdam, 2001; pp 39–74.
- (50) Kubas, G. J. The Extraordinary Dynamic Behavior and Reactivity of Dihydrogen and Hydride in the Coordination Sphere of Transition Metals. In *Handbook of Hydrogen Transfer*; Schowen, R. L., Ed.; Vol. 1: *Physical and Chemical Aspects of Hydrogen Transfer*; Hynes, J. T., Limbach, H. H., Eds.; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, 2006; p 603.
- (51) Kubas, G. J. Dihydrogen and Other Sigma Bond Complexes. In *Comprehensive Organometallic Chemistry III*; Crabtree, R. H., Mingos, D. M. P., Eds.; Vol. 1: *Special Topics*; Elsevier: Oxford, 2006; pp 671–698.
- (52) Bakhmutov, V. I. *Magn. Reson. Chem.* **2004**, *42*, 66.
- (53) Andrews, L. *Chem. Soc. Rev.* **2004**, *33*, 123.
- (54) Clapham, S. E.; Hadzovic, A.; Morris, R. H. *Coord. Chem. Rev.* **2004**, *248*, 2201.
- (55) Bakhmutov, V. I. *Eur. J. Inorg. Chem.* **2005**, 245.
- (56) Nolan, S. P.; Marks, T. J. *J. Am. Chem. Soc.* **1989**, *111*, 8538.
- (57) Chaudret, B.; Chung, G.; Eisenstein, O.; Jackson, S. A.; Lahoz, F. J.; Lopez, J. A. *J. Am. Chem. Soc.* **1991**, *113*, 2314.
- (58) Christ, M. L.; Sabo-Etienne, S.; Chaudret, B. *Organometallics* **1994**, *13*, 3800.
- (59) (a) Bart, S. C.; Lobkovsky, E.; Chirik, P. J. *J. Am. Chem. Soc.* **2004**, *126*, 13794. (b) Baya, M.; Houghton, J.; Daran, J.-C.; Poli, R.; Male, L.; Albinati, A.; Gutman, M. *Chem.—Eur. J.* **2007**, *13*, 5347.
- (60) Hasegawa, T.; Li, Z.; Parkin, S.; Hope, H.; McMullan, R. K.; Koetzle, T. F.; Taube, H. *J. Am. Chem. Soc.* **1994**, *116*, 4352.
- (61) (a) Aebischer, N.; Frey, U.; Merbach, A. E. *Chem. Commun.* **1998**, 2303. (b) Grundler, P. V.; Yazzev, O. V.; Aebischer, N.; Helm, L.; Laurency, G.; Merbach, A. E. *Inorg. Chim. Acta* **2006**, *359*, 1795.
- (62) Jia, G.; Lau, C. P. *J. Organomet. Chem.* **1998**, *565*, 37.
- (63) Vogt, M.; Pons, V.; Heinekey, D. M. *Organometallics* **2005**, *24*, 1832.
- (64) (a) Bacskay, G. B.; Bytheway, I.; Hush, N. S. *J. Am. Chem. Soc.* **1996**, *118*, 3753. (b) Hush, N. S. *J. Am. Chem. Soc.* **1997**, *119*, 1717. (c) Maltby, P. A.; Schlaf, M.; Steinbeck, M.; Lough, A. J.; Morris, R. H.; Klooster, W. T.; Koetzle, T. F.; Srivastava, R. C. *J. Am. Chem. Soc.* **1996**, *118*, 5396. (d) Luther, T. A.; Heinekey, D. M. *Inorg. Chem.* **1998**, *37*, 127. (e) Grundemann, S.; Limbach, H.-H.; Buntkowsky, G.; Sabo-Etienne, S.; Chaudret, B. *J. Phys. Chem. A* **1999**, *103*, 4752. (f) Gelabert, R.; Moreno, M.; Lluch, J. M.; Lledós, A.; Pons, V.; Heinekey, D. M. *J. Am. Chem. Soc.* **2004**, *126*, 8813. (g) Mort, B. C.; Autschbach, J. *J. Am. Chem. Soc.* **2006**, *128*, 10060.
- (65) (a) Grellier, M.; Vendier, L.; Chaudret, B.; Albinati, A.; Rizzato, S.; Mason, S.; Sabo-Etienne, S. *J. Am. Chem. Soc.* **2005**, *127*, 17592. (b) Grellier, M.; Vendier, L.; Sabo-Etienne, S. *Angew. Chem., Int. Ed.* **2007**, *46*, 2613.
- (66) Ingleson, M. J.; Brayshaw, S. K.; Mahon, M. F.; Ruggiero, G. D.; Weller, A. S. *Inorg. Chem.* **2005**, *44*, 3162.
- (67) Eckert, J.; Albinati, A.; Bucher, U. E.; Venanzi, L. M. *Inorg. Chem.* **1996**, *35*, 1292.
- (68) Heinekey, D. M.; Liegeois, A.; van Roon, M. *J. Am. Chem. Soc.* **1994**, *116*, 8388.
- (69) Gelabert, R.; Moreno, M.; Lluch, J. M. *Chem.—Eur. J.* **2005**, *11*, 6315.
- (70) Yousufuddin, M.; Wen, T. B.; Mason, S. A.; McIntyre, G. J.; Jia, G.; Bau, R. *Angew. Chem., Int. Ed.* **2005**, *44*, 7227.
- (71) Ghoshray, K.; Bandyopadhyay, B.; Sen, M.; Ghoshray, A.; Chatterjee, N. *Phys. Rev. B* **1993**, *47*, 8277.
- (72) Sen, M.; Ghoshray, A.; Ghoshray, K.; Sil, S.; Chatterjee, N. *Phys. Rev. B* **1996**, *53*, 14345.
- (73) Halet, J.-F.; Saillard, J.-Y.; Koudou, C.; Minot, C.; Nomikou, Z.; Hoffmann, R.; Demangeat, C. *Chem. Mater.* **1992**, *4*, 153 and references therein.
- (74) Huhmann-Vincent, J.; Scott, B. L.; Kubas, G. J. *J. Am. Chem. Soc.* **1998**, *120*, 6808.
- (75) (a) Belkova, N. V.; Collange, E.; Dub, P.; Epstein, L. M.; Lemenovskii, D. A.; Lledós, A.; Maresca, O.; Maseras, F.; Poli, R.; Revlin, P. O.; Shubina, E. S.; Vorontsov, E. V. *Chem.—Eur. J.* **2005**, *11*, 873. (b) Belkova, N. V.; Dub, P. A.; Baya, M.; Houghton, J. *Inorg. Chim. Acta* **2007**, *360*, 149.
- (76) Jia, G.; Ng, W. S.; Lau, C. P. *Organometallics* **1998**, *17*, 4538.
- (77) Gemel, C.; Huffman, J. C.; Caulton, K. G.; Mauthner, K.; Kirchner, K. *J. Organomet. Chem.* **2000**, *593–594*, 342.
- (78) (a) Mediat, M.; Tachibana, G. N.; Jensen, C. M. *Inorg. Chem.* **1992**, *31*, 1827. (b) Le-Husebo, T.; Jensen, C. M. *Inorg. Chem.* **1993**, *32*, 3797.
- (79) Zidan, R. A.; Rocheleau, R. E. *J. Mater. Res.* **1999**, *14*, 286.
- (80) Heinekey, D. M.; Radzewicz, C. E.; Voges, M. H.; Schomber, B. M. *J. Am. Chem. Soc.* **1997**, *119*, 4172.
- (81) Butts, M. D.; Kubas, G. J.; Luo, X.-L.; Bryan, J. C. *Inorg. Chem.* **1997**, *36*, 3341.
- (82) Kubas, G. J.; Nelson, J. E.; Bryan, J. C.; Eckert, J.; Wisniewski, L.; Zilm, K. *Inorg. Chem.* **1994**, *33*, 2954.
- (83) Gonzalez, A. A.; Mukerjee, S. L.; Chou, S.-L.; Zhang, K.; Hoff, C. D. *J. Am. Chem. Soc.* **1988**, *110*, 4419.
- (84) Lee, D. W.; Jensen, C. M. *J. Am. Chem. Soc.* **1996**, *118*, 8749.
- (85) Eckert, J.; Jensen, C. M.; Koetzle, T. F.; Le-Husebo, T.; Nicol, J.; Wu, P. *J. Am. Chem. Soc.* **1995**, *117*, 7271.
- (86) Hamilton, R. J.; Leong, C. G.; Bigam, G.; Miskolzie, M.; Bergens, S. H. *J. Am. Chem. Soc.* **2005**, *127*, 4152.
- (87) Based on use of the Sackur–Tetrode equation for translational entropy (see: Stull, D. R.; Westrum, E. F., Jr.; Sinke, G. C. *The Chemical Thermodynamics of Organic Compounds*; Wiley: New York, 1969). For approximate application to reactions in solution, see: Page M. I. *Angew. Chem., Int. Ed. Engl.* **1977**, *16*, 449.
- (88) (a) Kubas, G. J.; Ryan, R. R.; Wroblewski, D. J. *J. Am. Chem. Soc.* **1986**, *108*, 1339. (b) Kubas, G. J.; Ryan, R. R.; Unkefer, C. J. *J. Am. Chem. Soc.* **1987**, *109*, 8113. (c) Kubas, G. J.; Burns, C. J.; Eckert, J.; Johnson, S.; Larson, A. C.; Vergamini, P. J.; Unkefer, C. J.; Khalsa, G. R. K.; Jackson, S. A.; Eisenstein, O. *J. Am. Chem. Soc.* **1993**, *115*, 569.
- (89) Luo, X.-L.; Kubas, G. J.; Burns, C. J.; Eckert, J. *Inorg. Chem.* **1994**, *33*, 5219.
- (90) Luo, X.-L.; Kubas, G. J.; Bryan, J. C.; Burns, C. J.; Unkefer, C. J. *J. Am. Chem. Soc.* **1994**, *116*, 10312.
- (91) Huhmann-Vincent, J.; Scott, B. L.; Butcher, R.; Luo, S.; Unkefer, C. J.; Kubas, G. J.; Lledós, A.; Maseras, F.; Tomas, J. *Organometallics* **2003**, *22*, 5307.
- (92) Crabtree, R. H.; Lavin, M. *J. Chem. Soc., Chem. Commun.* **1985**, 794.
- (93) Crabtree, R. H.; Lavin, M.; Bonneviot, L. *J. Am. Chem. Soc.* **1986**, *108*, 4032.
- (94) Bianchini, C.; Linn, K.; Masi, D.; Peruzzini, M.; Polo, A.; Vacca, A.; Zanolini, F. *Inorg. Chem.* **1993**, *32*, 2366.
- (95) Gonzalez, A. A.; Hoff, C. D. *Inorg. Chem.* **1989**, *28*, 4295.
- (96) Sweany, R. L.; Moroz, A. J. *J. Am. Chem. Soc.* **1989**, *111*, 3577.
- (97) King, W. A.; Luo, X.-L.; Scott, B. L.; Kubas, G. J.; Zilm, K. W. *J. Am. Chem. Soc.* **1996**, *118*, 6782.

- (98) King, W. A.; Scott, B. L.; Eckert, J.; Kubas, G. J. *Inorg. Chem.* **1999**, *38*, 1069.
- (99) Toupadakis, A.; Kubas, G. J.; King, W. A.; Scott, B. L.; Huhmann-Vincent, J. *Organometallics* **1998**, *17*, 5315.
- (100) Fang, X.; Huhmann-Vincent, J.; Scott, B. L.; Kubas, G. J. *J. Organomet. Chem.* **2000**, *609*, 95.
- (101) Butts, M. D.; Kubas, G. J.; Scott, B. L. *J. Am. Chem. Soc.* **1996**, *118*, 11831.
- (102) Amendola, P.; Antonutti, S.; Albertin, G.; Bordignon, E. *Inorg. Chem.* **1990**, *29*, 318.
- (103) Chinn, M. S.; Heinekey, D. M.; Payne, N. G.; Sofield, C. D. *Organometallics* **1989**, *8*, 1824.
- (104) Bautista, M. T.; Cappellani, E. P.; Drouin, S. D.; Morris, R. H.; Schweitzer, C. T.; Sella, A.; Zubkowski, J. *J. Am. Chem. Soc.* **1991**, *113*, 4876.
- (105) Cappellani, E. P.; Maltby, P. A.; Morris, R. H.; Schweitzer, C. T.; Steele, M. R. *Inorg. Chem.* **1989**, *28*, 4437.
- (106) Chin, B.; Lough, A. J.; Morris, R. H.; Schweitzer, C.; D'Agostino, C. *Inorg. Chem.* **1994**, *33*, 6278.
- (107) Amrhein, P. I.; Drouin, S. D.; Forde, C. E.; Lough, A. J.; Morris, R. H. *J. Chem. Soc., Chem. Commun.* **1996**, 1665.
- (108) Rocchini, E.; Rigo, P.; Mezzetti, A.; Stephan, T.; Morris, R. H.; Lough, A. J.; Forde, C. E.; Fong, T. P.; Drouin, S. D. *J. Chem. Soc., Dalton Trans.* **2000**, 3591.
- (109) Mezzetti, A.; Del Zotto, A.; Rigo, P.; Farnetti, E. *J. Chem. Soc., Dalton Trans.* **1991**, 1525.
- (110) Esteruelas, M. A.; Garcia, M. P.; Lopez, A. M.; Oro, L. A.; Ruiz, N.; Schlunken, C.; Valero, C.; Werner, H. *Inorg. Chem.* **1992**, *31*, 5580.
- (111) Heinekey, D. M.; Voges, M. H.; Barnhart, D. M. *J. Am. Chem. Soc.* **1996**, *118*, 10792.
- (112) Hlatky, G. G.; Crabtree, R. H. *Coord. Chem. Rev.* **1985**, *65*, 1.
- (113) Hamilton, D. G.; Crabtree, R. H. *J. Am. Chem. Soc.* **1988**, *110*, 4126.
- (114) Bianchini, C.; Peruzzini, M.; Zanolini, F. *J. Organomet. Chem.* **1988**, *354*, C19.
- (115) Van Der Sluys, L. S.; Eckert, J.; Eisenstein, O.; Hall, J. H.; Huffman, J. C.; Jackson, S. A.; Koetzle, T. F.; Kubas, G. J.; Vergamini, P. J.; Caulton, K. G. *J. Am. Chem. Soc.* **1990**, *112*, 4831.
- (116) Jia, G.; Meek, D. W. *J. Am. Chem. Soc.* **1989**, *111*, 757.
- (117) Jia, G.; Meek, D. W.; Gallucci, J. C. *Inorg. Chem.* **1991**, *30*, 403.
- (118) Bucher, U. E.; Lengweiler, T.; Nanz, D.; von Philipsborn, W.; Venanzi, L. M. *Angew. Chem., Int. Ed. Engl.* **1990**, *29*, 548.
- (119) Cariati, F.; Ugo, R.; Bonati, F. *Inorg. Chem.* **1966**, *5*, 1128.
- (120) Cotton, F. A.; Luck, R. L. *J. Chem. Soc., Chem. Commun.* **1988**, 1277.
- (121) Harman, W. D.; Taube, H. *J. Am. Chem. Soc.* **1990**, *112*, 2261.
- (122) Nunes, F. S.; Taube, H. *Inorg. Chem.* **1994**, *33*, 3111, 3116.
- (123) Li, Z.-W.; Taube, H. *J. Am. Chem. Soc.* **1994**, *116*, 11584.
- (124) Li, Z.-W.; Taube, H. *J. Am. Chem. Soc.* **1991**, *113*, 8946.
- (125) Lin, P.; Hasegawa, T.; Parkin, S.; Taube, H. *J. Am. Chem. Soc.* **1992**, *114*, 2712.
- (126) Li, Z.-W.; Yeh, A.; Taube, H. *Inorg. Chem.* **1994**, *33*, 2874.
- (127) Abdur-Rashid, K.; Gusev, D. G.; Lough, A. J.; Morris, R. H. *Organometallics* **2000**, *19*, 1652.
- (128) (a) Werner, H.; Esteruelas, M. A.; Meyer, U.; Wrackmeyer, B. *Chem. Ber.* **1987**, *120*, 11. (b) Esteruelas, M. A.; Lahoz, F. J.; Lopez, J. A.; Oro, L. A.; Schlunken, C.; Valero, C.; Werner, H. *Organometallics* **1992**, *11*, 2034.
- (129) Gusev, D. G.; Kuhlman, R. L.; Renkema, K. B.; Eisenstein, O.; Caulton, K. G. *Inorg. Chem.* **1996**, *35*, 6775.
- (130) Bianchini, C.; Moneti, S.; Peruzzini, M.; Vizza, F. *Inorg. Chem.* **1997**, *36*, 5818.
- (131) Bakhmutov, V. I.; Bianchini, C.; Peruzzini, M.; Vizza, F.; Vorontsov, E. V. *Inorg. Chem.* **2000**, *39*, 1655.
- (132) Sweany, R. L. In *Transition Metal Hydrides*; Dedieu, A., Ed.; VCH Publishers, Inc.: New York, 1992; pp 65–101.
- (133) Darr, J. A.; Poliakov, M. *Chem. Rev.* **1999**, *99*, 495.
- (134) Goff, S. E. J.; Nolan, T. F.; George, M. W.; Poliakov, M. *Organometallics* **1998**, *17*, 2730.
- (135) (a) Banister, J. A.; Lee, P. D.; Poliakov, M. *Organometallics* **1995**, *14*, 3876. (b) Lee, P. D.; King, J. L.; Seebald, S.; Poliakov, M. *Organometallics* **1998**, *17*, 524.
- (136) Tspis, C. A. *Coord. Chem. Rev.* **1991**, *108*, 163.
- (137) van Leeuwen, P. W. N. M.; Morokuma, K.; van Lenthe, J. H., Eds. *Theoretical Aspects of Homogeneous Catalysis*; Kluwer Academic Publishers: Boston, 1995.
- (138) Musae, D. G.; Morokuma, K. *Adv. Chem. Phys.* **1996**, *95*, 61.
- (139) Dedieu, A. *Chem. Rev.* **2000**, *100*, 543.
- (140) (a) Frenking, G.; Fröhlich, N. *Chem. Rev.* **2000**, *100*, 717. (b) Niu, S.; Hall, M. B. *Chem. Rev.* **2000**, *100*, 353.
- (141) Torrent, M.; Solà, M.; Frenking, G. *Chem. Rev.* **2000**, *100*, 439.
- (142) Marx, D.; Parrinello, M. *Nature* **1995**, *375*, 216.
- (143) Thompson, K. C.; Crittenden, D. L.; Jordan, M. J. T. *J. Am. Chem. Soc.* **2005**, *127*, 4954.
- (144) Hay, P. J. *Chem. Phys. Lett.* **1984**, *103*, 466.
- (145) Dewar, M. J. S. *Bull. Soc. Chim. Fr.* **1951**, *18*, C79.
- (146) Chatt, J.; Duncanson, L. A. *J. Chem. Soc.* **1953**, 2929.
- (147) Gritsenko, O. V.; Bagatur'yants, A. A.; Moiseev, I. I.; Kazanskii, V. B.; Kalechits, I. V. *Kinet. Katal.* **1980**, *21*, 632; **1981**, *22*, 354.
- (148) Nakatsujii, H.; Hada, M. *Croat. Chem. Acta* **1984**, *57*, 1371.
- (149) (a) Blomberg, M. R. A.; Brandemark, U. B.; Petterson, L. G. M.; Siegbahn, P. E. M. *Int. J. Quantum Chem.* **1983**, *23*, 855. (b) Brandemark, U. B.; Blomberg, M. R. A.; Petterson, L. G. M.; Siegbahn, P. E. M. *J. Phys. Chem.* **1984**, *88*, 4617.
- (150) Jarque, C.; Novaro, O.; Ruiz, M. E.; Garcia-Prieto, J. *J. Am. Chem. Soc.* **1986**, *108*, 3507.
- (151) (a) Low, J. J.; Goddard, W. A., III. *J. Am. Chem. Soc.* **1984**, *106*, 8321. (b) Low, J. J.; Goddard, W. A., III. *Organometallics* **1986**, *5*, 609. (c) Nakatsujii, H.; Hada, M.; Yonezawa, T. *J. Am. Chem. Soc.* **1987**, *109*, 1902. (d) Balasubramanian, K.; Feng, P. Y.; Liao, D. W. *J. Chem. Phys.* **1988**, *88*, 6955.
- (152) (a) Ozin, G. A.; Garcia-Prieto, J. *J. Am. Chem. Soc.* **1986**, *108*, 3099. (b) Andrews, L.; Manceron, L.; Alikhani, M. E.; Wang, X. *J. Am. Chem. Soc.* **2000**, *122*, 11011. (c) Andrews, L.; Wang, X.; Alikhani, M. E.; Manceron, L. *J. Phys. Chem. A* **2001**, *105*, 3052.
- (153) Crabtree, R. H. *The Organometallic Chemistry of the Transition Metals*; Wiley & Sons: New York, 1988.
- (154) Li, J.; Ziegler, T. *Organometallics* **1996**, *15*, 3844.
- (155) Dapprich, S.; Frenking, G. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 354.
- (156) Dapprich, S.; Frenking, G. *Z. Anorg. Allg. Chem.* **1998**, *624*, 583.
- (157) (a) Dapprich, S.; Frenking, G. *Organometallics* **1996**, *15*, 4547. (b) Frenking, G.; Pidum, U. *J. Chem. Soc., Dalton Trans.* **1997**, 1653.
- (158) Bickelhaupt, F. M.; Baerends, E. J.; Ravenek, W. *Inorg. Chem.* **1990**, *29*, 350.
- (159) Li, J.; Dickson, R. M.; Ziegler, T. *J. Am. Chem. Soc.* **1995**, *117*, 11482 and references therein.
- (160) Maseras, F.; Li, X.-K.; Koga, N.; Morokuma, K. *J. Am. Chem. Soc.* **1993**, *115*, 10974.
- (161) Radius, U.; Bickelhaupt, F. M.; Ehlers, A. W.; Goldberg, N.; Hoffmann, R. *Inorg. Chem.* **1998**, *37*, 1080.
- (162) Kubas, G. J. *Comments Inorg. Chem.* **1988**, *7*, 17.
- (163) Heinekey, D. M.; Law, J. K.; Schultz, S. M. *J. Am. Chem. Soc.* **2001**, *123*, 12728.
- (164) Ishida, T.; Mizobe, Y.; Tanase, T.; Hidai, M. *J. Organomet. Chem.* **1991**, *409*, 355.
- (165) Brammer, L.; Howard, J. A.; Johnson, O.; Koetzle, T. F.; Spencer, J. L.; Stringer, A. M. *J. Chem. Soc., Chem. Commun.* **1991**, 241.
- (166) Albinati, A.; Bakhmutov, V. I.; Caulton, K. G.; Clot, E.; Eckert, J.; Eisenstein, O.; Gusev, D. G.; Grushin, V. V.; Hauger, B. E.; Klooster, W. T.; Koetzle, T. F.; McMullan, R. K.; O'Loughlin, T. J.; Pelissier, M.; Ricci, J. S.; Sigalas, M. P.; Vymenits, A. B. *J. Am. Chem. Soc.* **1993**, *115*, 7300.
- (167) Gusev, D. G. *J. Am. Chem. Soc.* **2004**, *126*, 14249.
- (168) Appleton, T. G.; Clark, H. C.; Manzer, L. E. *Coord. Chem. Rev.* **1973**, *10*, 335.
- (169) Dutta, S.; Jagirdar, B. R. *Inorg. Chem.* **2006**, *45*, 7047.
- (170) (a) Drouin, B. J.; Kukolich, S. G. *J. Am. Chem. Soc.* **1998**, *120*, 6774. (b) Lavaty, T. G.; Wikrent, P.; Drouin, B. J.; Kukolich, S. G. *J. Chem. Phys.* **1998**, *109*, 9473. (c) Wang, W.; Weitz, E. *J. Phys. Chem. A* **1997**, *101*, 2358. (d) Wang, W.; Narducci, A. A.; House, P. G.; Weitz, E. *J. Am. Chem. Soc.* **1996**, *118*, 8654. (e) Ziegler, T.; Tschinke, V.; Fan, L.; Becke, A. D. *J. Am. Chem. Soc.* **1989**, *111*, 9177.
- (171) Lesnard, H.; Demachy, I.; Jean, Y.; Lledós, A. *Chem. Commun.* **2003**, 850.
- (172) (a) Law, J. K.; Mellows, H.; Heinekey, D. M. *J. Am. Chem. Soc.* **2002**, *124*, 1024. (b) Gelabert, R.; Moreno, M.; Lluch, J. M.; Lledós, A.; Heinekey, D. M. *J. Am. Chem. Soc.* **2005**, *127*, 5632.
- (173) Eckert, J.; Kubas, G. J. *J. Chem. Phys.* **1993**, *97*, 2378.
- (174) Gross, C. L.; Girolami, G. S. *Organometallics* **2007**, *26*, 1658 and references therein.
- (175) Esteruelas, M. A.; Hernandez, Y. A.; Lopez, A. M.; Oliván, M.; Onate, E. *Organometallics* **2007**, *26*, 2193 and references therein.
- (176) (a) Bianchini, C.; Meli, A.; Peruzzini, M.; Frediani, P.; Bohanna, C.; Esteruelas, M. A.; Oro, L. A. *Organometallics* **1992**, *11*, 138. (b) Bianchini, C.; Meli, A.; Peruzzini, M.; Vizza, F.; Zanolini, F.; Frediani, P. *Organometallics* **1989**, *8*, 2080.
- (177) Bianchini, C.; Bohanna, C.; Esteruelas, M. A.; Frediani, P.; Meli, A.; Oro, L. A.; Peruzzini, M. *Organometallics* **1992**, *11*, 3837.
- (178) Bianchini, C.; Masi, D.; Peruzzini, M.; Casarin, M.; Maccato, C.; Rizzi, G. A. *Inorg. Chem.* **1997**, *36*, 1061.
- (179) Cotton, F. A.; Luck, R. L. *Inorg. Chem.* **1989**, *28*, 2181.
- (180) Bianchini, C.; Laschi, F.; Peruzzini, M.; Ottaviani, F. M.; Vacca, A.; Zanello, P. *Inorg. Chem.* **1990**, *29*, 3394.

- (181) Bruns, W.; Kaim, W.; Waldhor, E.; Krejčík, M. *Inorg. Chem.* **1995**, *34*, 663.
- (182) Howard, J. A. K.; Johnson, O.; Koetzle, T. F.; Spencer, J. L. *Inorg. Chem.* **1987**, *26*, 2930.
- (183) Johnson, T. J.; Albinati, A.; Koetzle, T. F.; Ricci, J.; Eisenstein, O.; Huffman, J. C.; Caulton, K. G. *Inorg. Chem.* **1994**, *33*, 4966.
- (184) Morris, R. H.; Earl, K. A.; Luck, R. L.; Lazarowich, N. J.; Sella, A. *Inorg. Chem.* **1987**, *26*, 2674.
- (185) Johnson, T. J.; Huffman, J. C.; Caulton, K. G.; Jackson, S. A.; Eisenstein, O. *Organometallics* **1989**, *8*, 2073.
- (186) Zilm, K. W.; Millar, J. M. *Adv. Magn. Opt. Reson.* **1990**, *15*, 163.
- (187) Zilm, K. W.; Merrill, R. A.; Kummer, M. W.; Kubas, G. J. *J. Am. Chem. Soc.* **1986**, *108*, 7837.
- (188) Nemcsok, D. S.; Kovacs, A.; Rayon, V. M.; Frenking, G. *Organometallics* **2002**, *21*, 5803.
- (189) Desrosiers, P. J.; Cai, L.; Lin, Z.; Richards, R.; Halpern, J. *J. Am. Chem. Soc.* **1991**, *113*, 4173.
- (190) (a) Morris, R. H.; Wittebort, R. J. *Magn. Reson. Chem.* **1997**, *35*, 243. (b) Bautista, M. T.; Earl, K. A.; Maltby, P. A.; Morris, R. H.; Schweitzer, C. T.; Sella, A. *J. Am. Chem. Soc.* **1988**, *110*, 7031.
- (191) Gottker-Schnetmann, I.; Heinekey, D. M.; Brookhart, M. *J. Am. Chem. Soc.* **2006**, *128*, 17114.
- (192) Eckert, J.; Webster, C. E.; Hall, M. B.; Albinati, A.; Venanzi, L. M. *Inorg. Chim. Acta* **2002**, *330*, 240.
- (193) Chopra, M.; Wong, K. F.; Jia, G.; Yu, N.-T. *J. Mol. Struct.* **1996**, *379*, 93.
- (194) Torres, L.; Gelabert, R.; Moreno, M.; Lluch, J. M. *J. Phys. Chem. A* **2000**, *104*, 7898.
- (195) (a) Martensson, A.-S.; Nyberg, C.; Andersson, S. *Phys. Rev. Lett.* **1986**, *57*, 2045. (b) Martensson, A.-S.; Nyberg, C.; Andersson, S. *Surf. Sci.* **1988**, *205*, 12.
- (196) *Transition Metal Hydrides*; Muetterties, E. L., Eds; Dekker: New York, 1971.
- (197) Meakin, P.; Guggenberger, L. J.; Peet, W. G.; Muetterties, E. L.; Jesson, J. P. *J. Am. Chem. Soc.* **1973**, *95*, 1467 and references therein.
- (198) Jesson, J. P.; Meakin, P. *Acc. Chem. Res.* **1973**, *6*, 269.
- (199) Gusev, D. G.; Berke, H. *Chem. Ber.* **1996**, *129*, 1143.
- (200) Bergamo, M.; Beringhelli, T.; D'Alfonso, G.; Mercandelli, P.; Sironi, A. *J. Am. Chem. Soc.* **2002**, *124*, 5117.
- (201) Egbert, J. D.; Bullock, R. M.; Heinekey, D. M. *Organometallics* **2007**, *26*, 2291 and references therein.
- (202) (a) Morris, R. H.; Sawyer, J. F.; Shiralian, M.; Zubkowski, J. D. *J. Am. Chem. Soc.* **1985**, *107*, 5581. (b) Baker, M. V.; Field, L. D.; Young, D. J. *J. Chem. Soc., Chem. Commun.* **1988**, 546.
- (203) Bayse, C. A.; Hall, M. B.; Pleune, B.; Poli, R. *Organometallics* **1998**, *17*, 4309.
- (204) (a) Luo, X.-L.; Crabtree, R. H. *J. Am. Chem. Soc.* **1990**, *112*, 6912. (b) Luo, X.-L.; Michos, D.; Crabtree, R. H. *Organometallics* **1992**, *11*, 237. (c) Gusev, D. G.; Nietlispach, D.; Eremenko, I. L.; Berke, H. *Inorg. Chem.* **1993**, *32*, 3628.
- (205) Heinekey, D. M.; Mellows, H.; Pratum, T. *J. Am. Chem. Soc.* **2000**, *122*, 6498.
- (206) Oldham, W. J., Jr.; Hinkle, A. S.; Heinekey, D. M. *J. Am. Chem. Soc.* **1997**, *119*, 11028.
- (207) (a) Maseras, F.; Duran, M.; Lledos, A.; Bertran, J. *J. Am. Chem. Soc.* **1992**, *114*, 2922. (b) Lin, Z.; Hall, M. B. *J. Am. Chem. Soc.* **1994**, *116*, 4446.
- (208) Jackson, S. A.; Eisenstein, O. *J. Am. Chem. Soc.* **1990**, *112*, 7203.
- (209) Riehl, J.-F.; Pelissier, M.; Eisenstein, O. *Inorg. Chem.* **1992**, *31*, 3344.
- (210) Maseras, F.; Duran, M.; Lledos, A.; Bertran, J. *J. Am. Chem. Soc.* **1991**, *113*, 2879.
- (211) Rodriguez, V.; Sabo-Etienne, S.; Chaudret, B.; Thoburn, J.; Ulrich, S.; Limbach, H.-H.; Eckert, J.; Barthelat, J.-C.; Hussein, K.; Marsden, C. *J. Inorg. Chem.* **1998**, *37*, 3475.
- (212) (a) Soubra, C.; Chan, F.; Albright, T. A. *Inorg. Chim. Acta* **1998**, *272*, 95. (b) Borowski, A. F.; Donnadieu, B.; Daran, J.-C.; Sabo-Etienne, S.; Chaudret, B. *Chem. Commun.* **2000**, 543.
- (213) Thompson, M. E.; Baxter, S. M.; Bulls, A. R.; Burger, B. J.; Nolan, M. C.; Santarsiero, B. D.; Schaefer, W. P.; Bercaw, J. E. *J. Am. Chem. Soc.* **1987**, *109*, 203.
- (214) Jacobsen, H. J. *Phys. Chem. A* **2002**, *106*, 6189.
- (215) Perutz, R. N.; Sabo-Etienne, S. *Angew. Chem., Int. Ed.* **2007**, *46*, 460.
- (216) Brintzinger, H. H. *J. Organomet. Chem.* **1979**, *171*, 337.
- (217) Eckert, J.; Jensen, C. M.; Jones, G.; Clot, E.; Eisenstein, O. *J. Am. Chem. Soc.* **1993**, *115*, 11056.
- (218) Wisniewski, L. L.; Mediat, M.; Jensen, C. M.; Zilm, K. W. *J. Am. Chem. Soc.* **1993**, *115*, 7533.
- (219) Li, S.; Hall, M. B.; Eckert, J.; Jensen, C. M.; Albinati, A. *J. Am. Chem. Soc.* **2000**, *122*, 2903.
- (220) Pons, V.; Conway, S. L. J.; Green, M. L. H.; Green, J. C.; Herbert, B. J.; Heinekey, D. M. *Inorg. Chem.* **2004**, *43*, 3475.
- (221) Janak, K. E.; Shin, J. H.; Parkin, G. *J. Am. Chem. Soc.* **2004**, *126*, 13054.
- (222) Sabo-Etienne, S.; Rodriguez, V.; Donnadieu, B.; Chaudret, B.; el Makarim, H. A.; Barthelat, J.-C.; Ulrich, S.; Limbach, H.-H.; Moïse, C. *New J. Chem.* **2001**, *25*, 55.
- (223) Lundquist, E. G.; Foltling, K.; Streib, W. E.; Huffman, J. C.; Eisenstein, O.; Caulton, K. G. *J. Am. Chem. Soc.* **1990**, *112*, 855.
- (224) Moreno, B.; Sabo-Etienne, S.; Chaudret, B.; Rodriguez, A.; Jalon, F.; Trofimenko, S. *J. Am. Chem. Soc.* **1995**, *117*, 7441.
- (225) (a) Klooster, W. T.; Koetzle, T. F.; Jia, G.; Fong, T. P.; Morris, R. H.; Albinati, A. *J. Am. Chem. Soc.* **1994**, *116*, 7677. (b) Earl, K. A.; Jia, G.; Maltby, P. A.; Morris, R. H. *J. Am. Chem. Soc.* **1991**, *113*, 3027.
- (226) (a) Gonzalez, A. A.; Zhang, K.; Nolan, S. P.; de la Vega, R. L.; Mukerjee, S. L.; Hoff, C. D.; Kubas, G. J. *Organometallics* **1988**, *7*, 2429. (b) Gonzalez, A. A.; Zhang, K.; Mukerjee, S. L.; Hoff, C. D.; Khalsa, G. R. K.; Kubas, G. J. *ACS Symp. Ser.* **1990**, *428*, 133. (c) Kubas, G. J.; Burns, C. J.; Khalsa, G. R. K.; Van Der Sluys, L. S.; Kiss, G.; Hoff, C. D. *Organometallics* **1992**, *11*, 3390. (d) Zhang, K.; Gonzalez, A. A.; Hoff, C. D. *J. Am. Chem. Soc.* **1989**, *111*, 3627.
- (227) Muckerman, J. T.; Fujita, E.; Hoff, C. D.; Kubas, G. J. *J. Phys. Chem. B* **2007**, *111*, 6815.
- (228) Grills, D. C.; van Eldik, R.; Muckerman, J. T.; Fujita, E. *J. Am. Chem. Soc.* **2006**, *128*, 15728.
- (229) Armstrong, F. A. *Curr. Opin. Chem. Biol.* **2004**, *8*, 133.
- (230) Volbeda, A.; Fonticella-Camps, J. C. *Coord. Chem. Rev.* **2005**, 1609.
- (231) Liu, X.; Ibrahim, S. K.; Tard, C.; Pickett, C. J. *Coord. Chem. Rev.* **2005**, 1641.
- (232) (a) Darensbourg, M. Y.; Lyon, E. J.; Zhao, Z.; Georgakaki, I. P. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 3683. (b) Darensbourg, M. Y.; Lyon, E. J.; Smees, J. J. *Coord. Chem. Rev.* **2000**, *206–207*, 533.
- (233) Peters, J. W.; Lanzilotta, W. N.; Lemon, B. J.; Seefeldt, L. C. *Science* **1998**, *282*, 1853.
- (234) Artero, V.; Fontecave, M. *Coord. Chem. Rev.* **2005**, 1518.
- (235) Capon, J.-F.; Gloaguen, F.; Schollhammer, P.; Talarmin, J. *Coord. Chem. Rev.* **2005**, 1664.
- (236) Frey, M. *Struct. Bonding (Berlin)* **1998**, *90*, 97.
- (237) Cammack, R. *Nature* **1995**, *373*, 556.
- (238) Fonticella-Camps, J. C.; Ragsdale, S. W. *Adv. Inorg. Chem.* **1999**, *47*, 283.
- (239) Adams, M. W. W.; Stiefel, E. I. *Curr. Opin. Struct. Biol.* **2000**, *4*, 214.
- (240) Cammack, R. *Nature* **1999**, *397*, 214.
- (241) Sellmann, D.; Sutter, J. *Acc. Chem. Res.* **1997**, *30*, 460.
- (242) Holm, R. H.; Kennepohl, P.; Solomon, E. I. *Chem. Rev.* **1996**, *96*, 2239.
- (243) (a) Thauer, R. K.; Klein, A. R.; Hartmann, G. C. *Chem. Rev.* **1996**, *96*, 3031. (b) Berkessel, A. *Curr. Opin. Chem. Biol.* **2001**, *5*, 486.
- (244) Henderson, R. A. *J. Chem. Soc., Dalton Trans.* **1995**, 503.
- (245) Albracht, S. P. J. *Biochim. Biophys. Acta* **1994**, *1188*, 167.
- (246) Montet, Y.; Amara, P.; Volbeda, A.; Vernede, X.; Hatchikian, E. C.; Field, M. J.; Frey, M.; Fonticella-Camps, J. C. *Nat. Struct. Biol.* **1997**, *4*, 523.
- (247) Nicolet, Y.; Piras, C.; Legrand, P.; Hatchikian, C. E.; Fonticella-Camps, J. C. *Structure* **1999**, *7*, 13.
- (248) (a) Nicolet, Y.; de Lacey, A. L.; Vernede, X.; Fernandez, V. M.; Legrand, P.; Hatchikian, C. E.; Fonticella-Camps, J. C. *J. Am. Chem. Soc.* **2001**, *123*, 1596. (b) Lyon, E. J.; Georgakaki, I. P.; Reibenspies, J. H.; Darensbourg, M. Y. *J. Am. Chem. Soc.* **2001**, *123*, 3268. (c) Fan, H.-J.; Hall, M. B. *J. Am. Chem. Soc.* **2001**, *123*, 3828.
- (249) Pereira, A. S.; Tavares, P.; Moura, I.; Moura, J. J. G.; Huynh, B. H. *J. Am. Chem. Soc.* **2001**, *123*, 2771.
- (250) Lemon, B. J.; Peters, J. W. *Biochemistry* **1999**, *38*, 12969; *J. Am. Chem. Soc.* **2000**, *122*, 3793.
- (251) (a) Volbeda, A.; Charon, M. H.; Piras, C.; Hatchikian, E. C.; Frey, M.; Fonticella-Camps, J. C. *Nature* **1995**, *373*, 580. (b) Maroney, M. J.; Bryngelson, P. A. *J. Biol. Inorg. Chem.* **2001**, *6*, 453. (c) Brecht, M.; van Gastel, M.; Buhre, T.; Friedrich, B.; Lubitz, W. *J. Am. Chem. Soc.* **2003**, *125*, 13075. (d) Burgdorf, T.; Loscher, S.; Liebisch, P.; Van der Linden, E.; Galander, M.; Lendzian, F.; Meyer-Klaucke, W.; Albracht, S. P. J.; Friedrich, B.; Dau, H.; Haumann, M. *J. Am. Chem. Soc.* **2005**, *127*, 576. (e) Ogata, H.; Mizoguchi, Y.; Mizuno, N.; Miki, K.; Adachi, S.; Yasuoka, N.; Yagi, T.; Yamauchi, O.; Hirota, S.; Higuchi, Y. *J. Am. Chem. Soc.* **2005**, *127*, 11628.
- (252) Bagley, K. A.; Duin, E. C.; Rosebloom, W.; Albracht, S. P. J.; Woodruff, W. H. *Biochemistry* **1995**, *34*, 5527.
- (253) Bagley, K. A.; Van Garderen, C. J.; Chen, M.; Duin, E. C.; Albracht, S. P. J.; Woodruff, W. H. *Biochemistry* **1994**, *33*, 9229.
- (254) Happe, R. P.; Rosebloom, W.; Pierek, A. J.; Albracht, S. P. J.; Bagley, K. A. *Nature* **1997**, *385*, 126. Pierek, A. J.; Rosebloom, W.; Happe, R. P.; Bagley, K. A.; Albracht, S. P. J. *J. Biol. Chem.* **1999**, *274*, 3331.

- (255) Garcin, E.; Vernede, X.; Volbeda, A.; Hatchikian, E. C.; Frey, M.; Fontecilla-Camps, J. C. *Structure* **1999**, 5, 557.
- (256) Cioslowski, J.; Boche, G. *Angew. Chem., Int. Ed. Engl.* **1997**, 36, 107.
- (257) Teles, J. H.; Brode, S.; Berkessel, A. *J. Am. Chem. Soc.* **1998**, 120, 1345.
- (258) (a) Olah, G. A.; Hartz, N.; Rasul, G.; Prakash, G. K. S. *J. Am. Chem. Soc.* **1995**, 117, 1336. (b) Olah, G. A. *Angew. Chem., Int. Ed. Engl.* **1995**, 34, 1393.
- (259) (a) Shima, S.; Lyon, E. J.; Thauer, R. K.; Mienert, B.; Bill, E. *J. Am. Chem. Soc.* **2005**, 127, 10430. (b) Korbas, M.; Vogt, S.; Meyer-Klaucke, W.; Bill, E.; Lyon, E. J.; Thauer, R. K.; Shima, S. *J. Biol. Chem.* **2006**, 281, 30804. (c) Lyon, E. J.; Shima, S.; Buurman, G.; Chowdhuri, S.; Batschauer, A.; Steinbach, K.; Thauer, R. K. *Eur. J. Biochem.* **2004**, 271, 195. (d) Pilak, O.; Mamat, B.; Vogt, S.; Hagemeyer, C. H.; Thauer, R. K. *J. Mol. Biol.* **2006**, 358, 798. (e) Thauer, R. K. The 8th International Hydrogenase Conference, Breckenridge, CO, August 5–10, 2007; Abstract L34.
- (260) Frey, G. D.; Lavallo, V.; Donnadieu, B.; Scholler, W. W.; Bertrand, G. *Science* **2007**, 316, 439.
- (261) McEvoy, J. P.; Brudvig, G. W. *Chem. Rev.* **2006**, 106, 4455.
- (262) Sun, L.; Akermark, B.; Ott, S. *Coord. Chem. Rev.* **2005**, 1653.
- (263) Lewis, N. S.; Nocera, D. G. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, 103, 15729.
- (264) Sutín, N.; Creutz, C.; Fujita, E. *Comments Inorg. Chem.* **1997**, 19, 67.
- (265) Crabtree, R. H. *Inorg. Chim. Acta* **1986**, 125, L7.
- (266) Forde, C. E.; Landau, S. E.; Morris, R. H. *J. Chem. Soc., Dalton Trans.* **1997**, 1663.
- (267) Brothers, P. J. *Prog. Inorg. Chem.* **1981**, 28, 1.
- (268) (a) Morris, R. H. In *Recent Advances in Hydride Chemistry*; Peruzzini, M.; Poli, R., Eds.; Elsevier Science B. V.: Amsterdam, 2001; pp 1–38. (b) Abdur-Rashid, K.; Fong, T. P.; Greaves, B.; Gusev, D. G.; Hinman, J. G.; Landau, S. E.; Lough, A. J.; Morris, R. H. *J. Am. Chem. Soc.* **2000**, 122, 9155 and references therein.
- (269) Calvin, M. *Trans. Faraday Soc.* **1938**, 34, 1181.
- (270) (a) Wilson, A. D.; Newell, R. H.; McNeven, M. J.; Muckerman, J. T.; Rakowski DuBois, M.; DuBois, D. L. *J. Am. Chem. Soc.* **2006**, 128, 358. (b) Wilson, A. D.; Shoemaker, A.; Miedaner, J. T.; Muckerman, J. T.; DuBois, D. L.; Rakowski DuBois, M. *Proc. Natl. Acad. Sci. U.S.A.* **2007**, 104, 6951. (c) Curtis, C. J.; Miedaner, A.; Ciancanelli, R.; Ellis, W. W.; Noll, B. C.; Rakowski DuBois, M.; DuBois, D. L. *Inorg. Chem.* **2003**, 42, 216.
- (271) Chinn, M. S.; Heinekey, D. M. *J. Am. Chem. Soc.* **1987**, 109, 5865.
- (272) Huhmann-Vincent, J.; Scott, B. L.; Kubas, G. J. *Inorg. Chem.* **1999**, 38, 115.
- (273) Perdoncin, G.; Scorrano, G. *J. Am. Chem. Soc.* **1977**, 99, 6983.
- (274) Fong, T. P.; Lough, A. J.; Morris, R. H.; Mezzetti, A.; Rocchini, E.; Rigo, P. *J. Chem. Soc., Dalton Trans.* **1998**, 2111.
- (275) Fong, T. P.; Forde, C. E.; Lough, A. J.; Morris, R. H.; Rigo, P.; Rocchini, E.; Stephan, T. *J. Chem. Soc., Dalton Trans.* **1999**, 4475.
- (276) Ontko, A. C.; Houllis, J. F.; Schnabel, R. C.; Roddick, D. M.; Fong, T. P.; Lough, A. J.; Morris, R. H. *Organometallics* **1998**, 17, 5467.
- (277) Huhmann-Vincent, J.; Scott, B. L.; Kubas, G. J. *Inorg. Chim. Acta* **1999**, 294, 240.
- (278) Cheng, T. Y.; Bullock, R. M. *Organometallics* **1995**, 14, 4031.
- (279) Beck, W.; Schweiger, M. Z. *Anorg. Allg. Chem.* **1991**, 595, 203.
- (280) Brewer, S. T.; Buggey, L. A.; Holloway, J. H.; Hope, E. G. *J. Chem. Soc., Dalton Trans.* **1995**, 2941.
- (281) Fernandez, J. M.; Gladysz, J. A. *Organometallics* **1989**, 8, 207.
- (282) Colman, M. R.; Newbound, T. D.; Marshall, L. J.; Noirot, M. D.; Miller, M. M.; Wulfberg, G. P.; Frye, J. S.; Anderson, O. P.; Strauss, S. H. *J. Am. Chem. Soc.* **1990**, 112, 2349.
- (283) Armdtsen, B. A.; Bergman, R. G. *Science* **1995**, 270, 1970.
- (284) Tellers, D. M.; Bergman, R. G. *J. Am. Chem. Soc.* **2001**, 123, 11508.
- (285) Forniés, J.; Martínez, F.; Navarro, R.; Urriolabeitia, E. P. *Organometallics* **1996**, 15, 1813.
- (286) Huang, D.; Huffman, J. C.; Bollinger, J. C.; Eisenstein, O.; Caulton, K. G. *J. Am. Chem. Soc.* **1997**, 119, 7398.
- (287) Basallote, M. G.; Besora, M.; Castillo, C. E.; Fernandez-Trujillo, J.; Lledós, A.; Maseras, F.; Manez, M. A. *J. Am. Chem. Soc.* **2007**, 129, 6608.
- (288) (a) Bullock, R. M.; Rappoli, B. J. *J. Chem. Soc., Chem. Commun.* **1989**, 1447. (b) Bullock, R. M.; Song, J.-S.; Szalda, D. J. *Organometallics* **1996**, 15, 2504.
- (289) Bullock, R. M.; Voges, M. H. *J. Am. Chem. Soc.* **2000**, 122, 12594.
- (290) (a) Voges, M. H.; Bullock, R. M. *J. Chem. Soc., Dalton Trans.* **2002**, 759. (b) Schlaf, M.; Ghosh, P.; Fagan, P. J.; Hauptman Bullock, R. M. *Angew. Chem., Int. Ed.* **2001**, 40, 3887. (c) Magee, M. P.; Norton, J. R. *J. Am. Chem. Soc.* **2001**, 123, 1778.
- (291) Dioumaev, V. K.; Bullock, R. M. *Nature* **2000**, 424, 530.
- (292) (a) Curtis, C. J.; Miedaner, A.; Raebiger, J. W.; DuBois, D. L. *Organometallics* **2004**, 23, 511. (b) Curtis, C. J.; Miedaner, A.; Ellis, W. W.; DuBois, D. L. *J. Am. Chem. Soc.* **2002**, 124, 1918.
- (293) (a) Breyse, M.; Furimsky, E.; Kasztelan, S.; Lacroix, M.; Perot, G. *Catal. Rev.* **2002**, 44, 651. (b) Neurock, M.; van Santen, R. A. *J. Am. Chem. Soc.* **1994**, 116, 4427.
- (294) Hwang, D.-Y.; Mebel, A. M. *J. Phys. Chem. A* **2002**, 106, 520.
- (295) Rakowski DuBois, M. *Chem. Rev.* **1989**, 89, 1 and references therein.
- (296) Sweeney, Z. K.; Polse, J. L.; Andersen, R. A.; Bergman, R. G.; Kubinec, M. G. *J. Am. Chem. Soc.* **1997**, 119, 4543.
- (297) Bianchini, C.; Mealli, C.; Meli, A.; Sabat, M. *Inorg. Chem.* **1986**, 25, 4617.
- (298) Rakowski DuBois, M.; Jagirdar, B.; Noll, B.; Dietz, S. In *Transition Metal Sulfur Chemistry*; Stiefel, E. I., Matsumoto, K., Eds.; ACS Symposium Series No. 653; American Chemical Society: Washington, DC, 1996; pp 269–281.
- (299) Pan, W.-H.; Harmer, M. A.; Halbert, T. R.; Stiefel, E. I. *J. Am. Chem. Soc.* **1984**, 106, 459.
- (300) Shibahara, T. *Coord. Chem. Rev.* **1993**, 123, 73 and references therein.
- (301) Kubas, G. J.; Ryan, R. R. *J. Am. Chem. Soc.* **1985**, 107, 6138.
- (302) Grushin, V. V. *Acc. Chem. Res.* **1993**, 26, 279.
- (303) (a) Gilbertson, J. D.; Szymczak, N. K.; Tyler, D. R. *Inorg. Chem.* **2004**, 43, 3341. (b) Szymczak, N. K.; Zakharov, L. N.; Tyler, D. R. *J. Am. Chem. Soc.* **2006**, 128, 15830. (c) Gilbertson, J. D.; Szymczak, N. K.; Crossland, J. L.; Miller, W. K.; Lyon, D. K.; Foxman, B. M.; Davis, J.; Tyler, D. R. *Inorg. Chem.* **2007**, 46, 1205.
- (304) Bianchini, C.; Barbaro, P.; Scapacci, G.; Zanobini, F. *Organometallics* **2000**, 19, 2450.
- (305) Lee, D.-H.; Patel, B. P.; Clot, E.; Eisenstein, O.; Crabtree, R. H. *Chem. Commun.* **1999**, 297.
- (306) Gruet, K.; Clot, E.; Eisenstein, O.; Lee, D. H.; Patel, B. P.; Macchioni, A.; Crabtree, R. H. *New J. Chem.* **2003**, 27, 80.
- (307) Conner, D.; Jajaprakash, K. N.; Cundari, T. R.; Gunnoe, T. B. *Organometallics* **2004**, 23, 272460.
- (308) (a) Fryzuk, M. D.; MacNeil, P. A. *Organometallics* **1983**, 2, 682. (b) Fryzuk, M. D.; MacNeil, P. A.; Rettig, S. J. *J. Am. Chem. Soc.* **1987**, 109, 2803.
- (309) (a) Lough, A. J.; Park, S.; Ramachandran, R.; Morris, R. H. *J. Am. Chem. Soc.* **1994**, 116, 8356. (b) Park, S.; Lough, A. J.; Morris, R. H. *Inorg. Chem.* **1996**, 35, 3001.
- (310) Lee, J. C., Jr.; Peris, E.; Rheingold, A. L.; Crabtree, R. H. *J. Am. Chem. Soc.* **1994**, 116, 11014.
- (311) Crabtree, R. H.; Siegbahn, P. E. M.; Eisenstein, O.; Rheingold, A. L.; Koetzle, T. F. *Acc. Chem. Res.* **1996**, 29, 348.
- (312) Crabtree, R. H. *Science* **1998**, 282, 2000.
- (313) Custelcean, R.; Jackson, J. E. *Chem. Rev.* **2001**, 101, 1963.
- (314) Epstein, L. M.; Shubina, E. S. *Coord. Chem. Rev.* **2002**, 231, 165.
- (315) Liu, Q.; Hoffmann, R. *J. Am. Chem. Soc.* **1995**, 117, 10108.
- (316) Braga, D.; Grepioni, F. *Coord. Chem. Rev.* **1999**, 183, 19.
- (317) Braga, D.; Grepioni, F.; Desiraju, G. R. *Chem. Rev.* **1998**, 98, 1375.
- (318) Jalon, F. A.; Manzano, B. R.; Caballero, A.; Carrion, M. C.; Santos, L.; Espino, G.; Moreno, M. *J. Am. Chem. Soc.* **2005**, 127, 15364.
- (319) Ogo, S.; Kabe, R.; Uehara, K.; Kure, B.; Nishimura, T.; Menon, S. C.; Harada, R.; Fukuzumi, S.; Higuchi, Y.; Ohhara, T.; Tamada, T.; Kuroki, R. *Science* **2007**, 316, 585. See also: Rauchfuss, T. B. *Science* **2007**, 316, 553.
- (320) Schlaf, M.; Lough, A. J.; Morris, R. H. *Organometallics* **1996**, 15, 4423.
- (321) Ienco, A.; Calhorda, M. J.; Reinhold, J.; Reineri, F.; Bianchini, C.; Meli, A.; Peruzzini, M.; Vizza, F.; Mealli, C. *J. Am. Chem. Soc.* **2004**, 126, 11954.
- (322) Kato, H.; Seino, H.; Mizobe, Y.; Hidai, M. *J. Chem. Soc., Dalton Trans.* **2002**, 1494.
- (323) Sweeney, Z. K.; Polse, J. L.; Andersen, R. A.; Bergman, R. G. *Organometallics* **1999**, 18, 5502.
- (324) Linck, R. C.; Pafford, R. J.; Rauchfuss, T. B. *J. Am. Chem. Soc.* **2001**, 123, 8856.
- (325) Kuwata, S.; Hidai, M. *Coord. Chem. Rev.* **2001**, 213, 211.
- (326) Sellmann, D.; Kappler, J.; Moll, M. *J. Am. Chem. Soc.* **1993**, 115, 1830.
- (327) Sellmann, D.; Rackelmann, G. H.; Heinemann, F. W. *Chem.—Eur. J.* **1997**, 3, 2071.
- (328) Sellmann, D.; Gottschalk-Gaudig, T.; Heinemann, F. W. *Inorg. Chem.* **1998**, 37, 3982.
- (329) Sellmann, D.; Geipel, F.; Moll, M. *Angew. Chem., Int. Ed.* **2000**, 39, 561.
- (330) Jessop, P. G.; Morris, R. H. *Inorg. Chem.* **1993**, 32, 2236.
- (331) Morris, R. H. *NATO ASI Ser., Ser. 3* **1998**, 60 (Transition Metal Sulfides), 57.
- (332) Stiefel, E. I. In *Transition Metal Sulfur Chemistry*; Matsumoto, K., Eds.; ACS Symposium Series No. 653; American Chemical Society: Washington, DC, 1996.

- (333) Bayon, J. C.; Claver, C.; Masdeu-Bulto, A. M. *Coord. Chem. Rev.* **1999**, 193–195, 73.
- (334) Sellmann, D.; Fursattel, A.; Sutter, J. *Coord. Chem. Rev.* **2000**, 200–202, 545 and references therein.
- (335) Jia, G.; Morris, R. H.; Schweitzer, C. T. *Inorg. Chem.* **1991**, 30, 594.
- (336) Nishibayashi, Y.; Takemoto, S.; Iwai, S.; Hidai, M. *Inorg. Chem.* **2000**, 39, 5946.
- (337) Fryzuk, M. D.; Love, J. B.; Rettig, S. J.; Young, V. G. *Science* **1997**, 275, 1445.
- (338) Basch, H.; Muaeve, D. G.; Morokuma, K.; Fryzuk, M. D.; Love, J. B.; Seidel, W. W.; Albinati, A.; Koetzle, T. F.; Klooster, W. T.; Mason, S. A.; Eckert, J. *J. Am. Chem. Soc.* **1999**, 121, 523.
- (339) Pool, J. A.; Lobkovsky, E.; Chirik, P. J. *Nature* **2004**, 427, 527.
- (340) Bernskoetter, W. H.; Olmos, A. V.; Lobkovsky, E.; Chirik, P. J. *Organometallics* **2006**, 25, 1021.
- (341) Brown, S. D.; Mehn, M. P.; Peters, J. C. *J. Am. Chem. Soc.* **2005**, 127, 13146.
- (342) Gilbertson, J. D.; Szymczak, N. K.; Tyler, D. R. *J. Am. Chem. Soc.* **2005**, 127, 10184.
- (343) Noyori, R.; Koizumi, M.; Ishii, D.; Ohkuma, T. *Pure Appl. Chem.* **2001**, 73, 227.
- (344) Ohkuma, T.; Noyori, R. *J. Am. Chem. Soc.* **2003**, 125, 13490 and references therein.
- (345) Noyori, R. *Angew. Chem., Int. Ed.* **2002**, 41, 2008.
- (346) Abdur-Rashid, K.; Clapham, S. E.; Hadzovic, A.; Harvey, J. N.; Lough, A. J.; Morris, R. H. *J. Am. Chem. Soc.* **2002**, 124, 15104.
- (347) Hartmann, R.; Chen, P. *Angew. Chem., Int. Ed.* **2001**, 40, 3581.
- (348) (a) Hedberg, C.; Kallstrom, K.; Arvidsson, P. I.; Brandt, P.; Andersson, P. G. *J. Am. Chem. Soc.* **2005**, 127, 15083. (b) Casey, C. P.; Johnson, J. B.; Singer, S. W.; Cui, Q. *J. Am. Chem. Soc.* **2005**, 127, 3100. (c) Dahlenburg, L.; Gotz, R. *Inorg. Chem. Commun.* **2003**, 6, 443. (d) Muniz, K. *Angew. Chem., Int. Ed.* **2005**, 44, 6622.
- (349) Hutschka, F.; Dedieu, A. *J. Chem. Soc., Dalton Trans.* **1997**, 1899.
- (350) Llamazares, A.; Schmalte, H. W.; Berke, H. *Organometallics* **2001**, 20, 5277.
- (351) Nagaraja, C. M.; Parameswaran, P.; Jemmis, E. D.; Jagirdar, B. R. *J. Am. Chem. Soc.* **2007**, 129, 5587.
- (352) Chan, W.-C.; Lau, C.-P.; Chen, Y.; Fang, Y.-Q.; Ng, S.-M.; Jia, G. *Organometallics* **1997**, 16, 34.
- (353) Szymczak, N. K.; Zakharov, L. N.; Tyler, D. R. *J. Am. Chem. Soc.* **2006**, 128, 15830.
- (354) Chatt, J.; Dilworth, J. R.; Richards, R. L. *Chem. Rev.* **1978**, 78, 589.
- (355) Bancroft, G. M.; Garrod, R. E.; Maddock, A. G.; Mays, M. J.; Prater, B. E. *J. Am. Chem. Soc.* **1972**, 94, 647.
- (356) Morris, R. H.; Schlaf, M. *Inorg. Chem.* **1994**, 33, 1725.
- (357) Kaltsoyannis, N.; Scott, P. *J. Chem. Soc., Chem. Commun.* **1998**, 1665.
- (358) Rosi, M.; Sgamellotti, A.; Tarantelli, F.; Floriani, C.; Cederbaum, L. S. *J. Chem. Soc., Dalton Trans.* **1989**, 33.
- (359) (a) Krasna, A. I.; Rittenberg, D. J. *J. Am. Chem. Soc.* **1954**, 76, 3015. (b) Adams, M. W. W.; Mortenson, L. E.; Chen, J.-S. *Biochim. Biophys. Acta* **1981**, 594, 105. (c) Lespinat, P. A.; Berlier, Y.; Faque, G.; Czechowski, M.; Dimon, B.; LeGall, J. *Biochimie* **1986**, 68, 55. (d) Vignais, P. M. *Coord. Chem. Rev.* **2005**, 249, 1677. (e) Evans, D. J.; Pickett, C. J. *Chem. Soc. Rev.* **2003**, 32, 268. (f) Henrici-Olivé, G.; Olivé, S. *J. Mol. Catal.* **1975/76**, 1, 121. (g) Collman, J. P.; Wagenknecht, P. S.; Hembre, R. H.; Lewis, N. S. *J. Am. Chem. Soc.* **1990**, 112, 1269. (h) Collman, J. P.; Wagenknecht, P. S.; Hutchison, J. E.; Lewis, N. S.; Lopez, M. A.; Guillard, R.; L'Her, M. *J. Am. Chem. Soc.* **1992**, 114, 5654. (i) Zimmer, M.; Schulte, G.; Luo, X.-L.; Crabtree, R. H. *Angew. Chem., Int. Ed. Engl.* **1991**, 30, 193. (j) Tye, J. W.; Hall, M. B.; Georgakaki, I. P.; Darenbourg, M. Y. *Adv. Inorg. Chem.* **2004**, 56, 1. (k) Zhao, X.; Georgakaki, I. P.; Miller, M. L.; Yarbrough, J. C.; Darenbourg, M. Y. *J. Am. Chem. Soc.* **2001**, 123, 9710. (l) Zhao, X.; Georgakaki, I. P.; Miller, M. L.; Mejia-Rodriguez, R.; Chiang, C.; Darenbourg, M. Y. *Inorg. Chem.* **2002**, 41, 3917. (m) Carriker, J. L.; Wagenknecht, P. S.; Hosseini, M. A.; Fleming, P. E. *J. Mol. Catal. A* **2007**, 267, 218. (n) Kovacs, G.; Nadasdi, L.; Laurenczy, G.; Joo, F. *Green Chem.* **2003**, 5, 213. (o) Leung, C. W.; Zheng, W.; Wang, D.; Ng, S. M.; Yeung, C. H.; Zhou, Z.; Lin, Z.; Lau, C. P. *Organometallics* **2007**, 26, 1924.
- (360) Albeniz, A. C.; Heinekey, D. M.; Crabtree, R. H. *Inorg. Chem.* **1991**, 30, 3632.
- (361) Heinekey, D. M.; Schomber, B. M.; Radzewich, C. E. *J. Am. Chem. Soc.* **1994**, 116, 4515.
- (362) Schlaf, M.; Lough, A. J.; Maltby, P. A.; Morris, R. H. *Organometallics* **1996**, 15, 2270.
- (363) Reid, S. M.; Neuner, B.; Schrock, R. R.; Davis, W. M. *Organometallics* **1998**, 17, 4077.
- (364) Cappellani, E. P.; Drouin, S. D.; Jia, G.; Maltby, P. A.; Morris, R. H.; Schweitzer, C. T. *J. Am. Chem. Soc.* **1994**, 116, 3375.
- (365) Lau, C.-P.; Cheng, L. *J. Mol. Catal.* **1993**, 84, 39.
- (366) Hembre, R. T.; McQueen, S. J. *Am. Chem. Soc.* **1994**, 116, 2141.
- (367) Van Der Sluys, L. S.; Miller, M. M.; Kubas, G. J.; Caulton, K. G. *J. Am. Chem. Soc.* **1991**, 113, 2513.
- (368) Sola, E.; Bakhmutov, V. I.; Torres, F.; Elduque, A.; Lopez, J. A.; Lahoz, F. J.; Werner, H.; Oro, L. A. *Organometallics* **1998**, 17, 683.
- (369) *Spin Crossover in Transition Metal Compounds I. Topics in Current Chemistry*, 235; Gütllich, P., Goodwin, H. A., Eds.; Springer-Verlag: Berlin, Heidelberg, 2004.
- (370) Poli, R. *Chem. Rev.* **1996**, 96, 2135.
- (371) Alberding, N.; et al. *Biophys. J.* **1978**, 24, 319.
- (372) McMahon, B. H.; Stojkovic, B. P.; Hay, P. J.; Martin, R. L.; Garcia, A. E. *J. Chem. Phys.* **2000**, 113, 6831.
- (373) Thompson, D. W.; Kretzer, R. M.; Lebeau, E. L.; Scaltrito, D. V.; Ghilardi, R. A.; Lam, K.-C.; Rheingold, A. L.; Karlin, K. D.; Meyer, G. J. *Inorg. Chem.* **2003**, 42, 5211.
- (374) Harvey, J. N. *J. Am. Chem. Soc.* **2000**, 122, 12401.
- (375) Calderazzo, F.; Fachinetti, G.; Floriani, C. *J. Am. Chem. Soc.* **1974**, 96, 3695.
- (376) Wong, K. L. T.; Brintzinger, H. H. *J. Am. Chem. Soc.* **1975**, 97, 5143.
- (377) Hardman, N. J.; Fang, X.; Wright, R. J.; Scott, B. L.; Martin, R. L.; Kubas, G. J. *Inorg. Chem.* **2005**, 44, 8306.
- (378) (a) Fang, X.; Scott, B. L.; Watkin, J. G.; Kubas, G. J. *Organometallics* **2000**, 19, 4193. (379) Fang, X.; Watkin, J. G.; Scott, B. L.; Kubas, G. J.; *Organometallics* **2001**, 20, 3351.
- (379) (a) Henry, R. M.; Shoemaker, R. K.; Newell, R. H.; Jacobsen, G. M.; DuBois, D. L.; Rakowski DuBois, M. *Organometallics* **2005**, 24, 2481. (b) Henry, R. M.; Shoemaker, R. K.; DuBois, D. L.; Rakowski DuBois, M. *J. Am. Chem. Soc.* **2006**, 128, 3002.
- (380) (a) Carreon-Macedo, J.-L.; Harvey, J. N. *J. Am. Chem. Soc.* **2004**, 126, 5789. (b) Poli, R. N.; Harvey, J. N. *Chem. Soc. Rev.* **2003**, 32, 1.
- (381) Greco, C.; Bruschi, M.; de Gioia, L.; Ryde, U. *Inorg. Chem.* **2007**, 46, 594.
- (382) Niu, S.; Thomson, L. M.; Hall, M. B. *J. Am. Chem. Soc.* **1999**, 121, 4000.
- (383) Volbeda, A.; Fontecilla-Camps, J. C. *Dalton Trans.* **2003**, 4030.
- (384) Reissmann, S.; Hochleitner, E.; Wang, H.; Paschos, A.; Lottspeich, F.; Glass, R. S.; Bock, A. *Science* **2003**, 299, 1067.
- (385) Glass, R. S.; Paschos, A.; Reissmann, S.; Singh, M.; Wang, H.; Bock, A. *Phosphorus, Sulfur Silicon, Relat. Elem.* **2005**, 180, 1183.
- (386) Rauchfuss, T. B.; Contakes, S. M.; Hsu, S. C. N.; Reynolds, M. A.; Wilson, S. R. *J. Am. Chem. Soc.* **2001**, 123, 6933.
- (387) Dismukes, C. G. *Chem. Rev.* **1996**, 96, 2909.
- (388) Connelly, N. G.; Dahl, L. F. *J. Am. Chem. Soc.* **1970**, 92, 7472.
- (389) Vergamini, P. J.; Kubas, G. J. *Prog. Inorg. Chem.* **1976**, 21, 261.
- (390) Beinert, H.; Holm, R. H.; Münck, E. *Science* **1997**, 277, 653.
- (391) Siegbahn, P. E. M. *Inorg. Chem.* **1999**, 38, 2880.
- (392) Han, J.; Beck, K.; Ockwig, N.; Coucouvanis, D. *J. Am. Chem. Soc.* **1999**, 121, 10488.
- (393) Peters, J. W.; Stowell, M. H. B.; Soltis, S. M.; Finnegan, M. G.; Johnson, M. K.; Rees, D. C. *Biochemistry* **1997**, 36, 1181.
- (394) Deng, H.; Hoffmann, R. *Angew. Chem., Int. Ed. Engl.* **1993**, 32, 1062.
- (395) Nataro, C.; Angelici, R. J. *Inorg. Chem.* **1998**, 37, 2975.
- (396) (a) Siegbahn, P. E. M. *Chem. Rev.*, this thematic issue. (b) Pavlov, M.; Siegbahn, P. E. M.; Blomberg, M. R. A.; Crabtree, R. H. *J. Am. Chem. Soc.* **1998**, 120, 548. (c) Siegbahn, P. E. M.; Blomberg, M. R. A. *Chem. Rev.* **2000**, 100, 421.
- (397) Niu, S.; Hall, M. B. *Inorg. Chem.* **2001**, 39, 6201.
- (398) Pardo, A.; De Lacey, A. L.; Fernandez, V. M.; Fan, H.-J.; Fan, Y.; Hall, M. B. *J. Biol. Inorg. Chem.* **2006**, 11, 286.
- (399) Fan, C.; Teixeira, M.; Moura, I.; Huynh, B.-H.; LeGall, J.; Peck, H. D., Jr.; Hoffman, B. M. *J. Am. Chem. Soc.* **1991**, 113, 20.
- (400) Volbeda, A.; Garcin, E.; Piras, C.; de Lacey, A. L.; Fernandez, V. M.; Hatchikian, E. C.; Frey, M.; Fontecilla-Camps, J. C. *J. Am. Chem. Soc.* **1996**, 118, 12989.
- (401) Cao, Z.; Hall, M. B. *J. Am. Chem. Soc.* **2001**, 123, 3734.
- (402) Justice, A. K.; Linck, R. C.; Rauchfuss, T. B.; Wilson, S. R. *J. Am. Chem. Soc.* **2004**, 126, 13214.
- (403) Harris, D. C.; Gray, H. B. *Inorg. Chem.* **1975**, 14, 1215.
- (404) (a) Fauvel, K.; Mathieu, R.; Poilblanc, R. *Inorg. Chem.* **1976**, 15, 976. (b) Arabi, M. S.; Mathieu, R.; Poilblanc, R. *J. Organomet. Chem.* **1979**, 177, 199.
- (405) (a) Ezzaher, S.; Capon, J.-F.; Gloaguen, F.; Pétilion, F. Y.; Schollhammer, P.; Talarmin, J.; Pichon, R.; Kervarec, N. *Inorg. Chem.* **2007**, 46, 3426. (b) Morvan, D.; Capon, J.-F.; Gloaguen, F.; Le Goff, A.; Marchivie, M.; Michaud, F.; Schollhammer, P.; Talarmin, J.; Yaouanc, J.-J. *Organometallics* **2007**, 26, 2042. (c) Van Der Vlugt, J. I.; Rauchfuss, T. B.; Whaley, C. M.; Wilson, S. R. *J. Am. Chem. Soc.* **2005**, 127, 16012. (d) Eilers, G.; Schwartz, L.; Stein, M.; Zampella, G.; de Gioia, L.; Ott, S.; Lomoth, R. *Chem.—Eur. J.* **2007**, 13, 7075.

- (406) Dance, I. *Chem. Commun.* **1999**, 1655.
- (407) (a) Liu, T.; Darensbourg, M. Y. *J. Am. Chem. Soc.* **2007**, *129*, 7008. (b) Justice, A. K.; Rauchfuss, T. B.; Wilson, S. R. *Angew. Chem., Int. Ed.* **2007**, *46*, 6152.
- (408) (a) Burgess, B. K.; Lowe, D. J. *Chem. Rev.* **1996**, *96*, 2983. (b) Eady, R. R. *Chem. Rev.* **1996**, *96*, 3013.
- (409) Coucouvanis, D. *Adv. Inorg. Chem.* **1998**, *45*, 1.
- (410) Einsle, O.; Tezcan, F. A.; Andrade, S. L. A.; Schmid, B.; Yoshida, M.; Howard, J. B.; Rees, D. C. *Science* **2002**, *297*, 1696.
- (411) Smith, B. E. *Adv. Inorg. Chem.* **1999**, *47*, 159.
- (412) Hidai, M.; Mizobe, Y. *Chem. Rev.* **1995**, *95*, 1115.
- (413) Fryzuk, M. D.; Johnson, S. A. *Coord. Chem. Rev.* **2000**, *200–202*, 379.
- (414) Shilov, A. E. *Metal Complexes in Biomimetic Chemical Reactions*; CRC Press: Boca Raton, FL, 1997.
- (415) Hidai, M. *Coord. Chem. Rev.* **1999**, *185–186*, 99.
- (416) Nishibayashi, Y.; Iwai, S.; Hidai, M. *Science* **1998**, *279*, 540.
- (417) Barriere, F. *Coord. Chem. Rev.* **2003**, *236*, 71.
- (418) Sellmann, D.; Utz, J.; Blum, N.; Heinemann, F. W. *Coord. Chem. Rev.* **1999**, *190–192*, 607.
- (419) Yang, T.-C.; Maeser, N. K.; Laryukhin, M.; Lee, H.-I.; Dean, D. R.; Seefeldt, L. C.; Hoffman, B. M. *J. Am. Chem. Soc.* **2005**, *127*, 12804.
- (420) Igarashi, R. Y.; Laryukhin, M.; Dos Santos, P. C.; Lee, H.-I.; Dean, D. R.; Seefeldt, L. C.; Hoffman, B. M. *J. Am. Chem. Soc.* **2005**, *127*, 6231.
- (421) Dance, I. *J. Am. Chem. Soc.* **2005**, *127*, 10925; **2007**, *129*, 1076.
- (422) Le Gall, T.; Ibrahim, S. K.; Gormal, C. A.; Smith, B. E.; Pickett, C. J. *Chem. Commun.* **1999**, 773.
- (423) Hellere, C. A.; Henderson, R. A.; Leigh, G. J. *J. Chem. Soc., Dalton Trans.* **1999**, 1213.
- (424) Thorneley, R. N. F.; Lowe, D. J. In *Molybdenum Enzymes*; Spiro, T. G., Ed.; Wiley-Interscience: New York, 1985.
- (425) Hughes, D. L.; Ibrahim, S. K.; Pickett, C. J.; Querne, G.; Lauoenan, A.; Talarmin, J.; Queiros, A.; Fonseca, A. *Polyhedron* **1994**, *13*, 3341.
- (426) McCusker, J. K. *Science* **2001**, *293*, 1599.
- (427) Na, Y.; Pan, J. P.; Wang, M.; Sun, L. *Inorg. Chem.* **2007**, *46*, 3813.
- (428) Lever, A. B. P. *Inorg. Chem.* **1990**, *29*, 1271.
- (429) Justice, A. K.; Linck, R. C.; Rauchfuss, T. B. *Inorg. Chem.* **2006**, *45*, 2406.
- (430) Heben, M. J. *Chem. Rev.*, this thematic issue.
- (431) (a) Gagliardi, L.; Pyykko, P. *J. Am. Chem. Soc.* **2004**, *126*, 15014. (b) Raab, J.; Lindh, R. H.; Wang, X.; Andrews, L.; Gagliardi, L. *J. Phys. Chem. A* **2007**, *111*, 6383.
- (432) Zhao, Y.; Kim, Y.-H.; Dillon, A. C.; Heben, M. J.; Zhang, S. B. *Phys. Rev. Lett.* **2005**, *94*, 155504.
- (433) (a) Weisshaar, J. C. *Acc. Chem. Res.* **1993**, *26*, 213. (b) Armentrout, P. B. *Acc. Chem. Res.* **1995**, *28*, 430.
- (434) Kemper, P. R.; Bushnell, J. E.; von Helden, G.; Bowers, M. T. *J. Phys. Chem.* **1993**, *97*, 52.
- (435) Bushnell, J. E.; Kemper, P. R.; Bowers, M. T. *J. Phys. Chem.* **1995**, *99*, 15602.
- (436) Bushnell, J. E.; Kemper, P. R.; Bowers, M. T. *J. Phys. Chem.* **1993**, *97*, 11628.
- (437) Kemper, P. R.; Bushnell, J. E.; van Koppen, P.; Bowers, M. T. *J. Phys. Chem.* **1993**, *97*, 1810.
- (438) Bushnell, J. E.; Kemper, P. R.; Maitre, P.; Bowers, M. T. *J. Am. Chem. Soc.* **1994**, *116*, 9710.
- (439) Bushnell, J. E.; Kemper, P. R.; Bowers, M. T. *J. Phys. Chem.* **1994**, *98*, 2044.
- (440) Bushnell, J. E.; Maitre, P.; Kemper, P. R.; Bowers, M. T. *J. Chem. Phys.* **1997**, *106*, 10153.
- (441) Weis, P.; Kemper, P. R.; Bowers, M. T. *J. Phys. Chem. A* **1997**, *101*, 2809.
- (442) Kemper, P. R.; Weis, P.; Bowers, M. T. *Chem. Phys. Lett.* **1998**, *293*, 503.
- (443) Kemper, P. R.; Weis, P.; Bowers, M. T.; Maitre, P. *J. Am. Chem. Soc.* **1998**, *120*, 13494.
- (444) Haynes, C. L.; Armentrout, P. B. *Chem. Phys. Lett.* **1996**, *249*, 64.
- (445) Tjelta, B. L.; Armentrout, P. B. *J. Phys. Chem. A* **1997**, *101*, 2064.
- (446) Sievers, M. R.; Jarvis, L. M.; Armentrout, P. B. *J. Am. Chem. Soc.* **1998**, *120*, 1891.
- (447) Haynes, C. L.; Armentrout, P. B.; Perry, J. K.; Goddard, W. A., III. *J. Phys. Chem.* **1995**, *99*, 6340.
- (448) Haynes, C. L.; Chen, Y.-M.; Armentrout, P. B. *J. Phys. Chem.* **1995**, *99*, 9110.
- (449) Schultz, R. H.; Haynes, C. L. *J. Phys. Chem.* **1993**, *97*, 596.
- (450) Sanchez, M.; Ruetter, F.; Hernandez, A. J. *J. Phys. Chem.* **1992**, *96*, 823.
- (451) Niu, J.; Rao, B. K.; Jena, P.; Manninen, M. *Phys. Rev. B* **1995**, *51*, 4475.
- (452) Niu, J.; Rao, B. K.; Jena, P. *Phys. Rev. Lett.* **1992**, *68*, 2277.
- (453) Bauschlicher, C. W., Jr.; Maitre, P. *J. Phys. Chem.* **1995**, *99*, 5238.
- (454) Perry, J. K.; Ohanessian, G.; Goddard, W. A., III. *J. Phys. Chem.* **1993**, *97*, 5238.
- (455) Siegbahn, P.; Blomberg, M.; Panas, I.; Wahlgren, U. *Theor. Chim. Acta* **1989**, *75*, 143.
- (456) Li, J.; Schiott, B.; Hoffmann, R.; Proserpio, D. M. *J. Phys. Chem.* **1990**, *94*, 1554.
- (457) Hauge, R. H.; Margrave, J. L.; Kafafi, Z. H. *NATO ASI Ser., Ser. B* **1987**, *158* (Phys. Chem. Small Clusters), 787.
- (458) Nicolaides, C. A.; Simandiras, E. D. *Comments Inorg. Chem.* **1996**, *18*, 65 and references therein.
- (459) Blickensderfer, R. P.; Jordan, K. D.; Adams, N.; Breckenridge, W. H. *J. Phys. Chem.* **1982**, *86*, 1930.
- (460) Curtiss, L. A.; Pople, J. A. *J. Phys. Chem.* **1988**, *92*, 894.
- (461) Nicolaides, C. A.; Valtazanos, P. *Chem. Phys. Lett.* **1990**, *174*, 489; **1991**, *176*, 239.
- (462) Valtazanos, P.; Nicolaides, C. A. *J. Chem. Phys.* **1993**, *98*, 549.
- (463) Hwang, D.-Y.; Mebel, A. M. *Chem. Phys. Lett.* **2000**, *321*, 95.
- (464) Hwang, D.-Y.; Mebel, A. M. *J. Am. Chem. Soc.* **2000**, *122*, 11406.
- (465) Frenking, G.; Dapprich, S.; Kohler, K. F.; Koch, W.; Collins, J. R. *Mol. Phys.* **1996**, *89*, 1245.
- (466) Nicolaides, C. A.; Simandiras, E. D. *Chem. Phys. Lett.* **1992**, *196*, 213.
- (467) Simandiras, E. D.; Nicolaides, C. A. *Chem. Phys. Lett.* **1994**, *223*, 233.
- (468) Rodriguez, L. J.; Ruetter, F.; Rosa-Brussin, M. *J. Mol. Catal.* **1990**, *62*, 199.
- (469) Fiedler, A.; Schroder, D.; Shaik, S.; Schwarz, H. *J. Am. Chem. Soc.* **1994**, *116*, 10734.
- (470) Over, H.; Kim, Y. D.; Seitsonen, A. P.; Wendt, S.; Lundgren, E.; Schmid, M.; Varga, P.; Morgante, A.; Ertl, G. *Science* **2000**, *287*, 1474.
- (471) Wang, J.; Fan, C. Y.; Sun, Q.; Reuter, K.; Jacobi, K.; Scheffler, M.; Ertl, G. *Angew. Chem., Int. Ed.* **2003**, *42*, 2151.
- (472) Sun, Q.; Reuter, K.; Scheffler, M. *Phys. Rev. B* **2004**, *70*, 235402.
- (473) Albinati, A.; Klooster, W. T.; Koetzle, T. F.; Fortin, J. B.; Ricci, J. S.; Eckert, J.; Fong, T. P.; Lough, A. J.; Morris, R. H.; Golombek, A. P. *Inorg. Chim. Acta* **1997**, *259*, 351.
- (474) Burwell, R. L., Jr.; Haller, G. L.; Taylor, K. C.; Read, J. F. *Adv. Catal.* **1969**, *20*, 1.
- (475) Burwell, R. L., Jr.; Stec, K. S. *J. Colloid Interface Sci.* **1977**, *58*, 54.
- (476) Hwang, D.-Y.; Mebel, A. M. *J. Phys. Chem. A* **2002**, *106*, 520.
- (477) Hwang, D.-Y.; Mebel, A. M. *Chem. Phys. Lett.* **2001**, *341*, 393.
- (478) Hermansson, K.; Baudin, M.; Ensing, B.; Alfreðsson, M.; Wojcik, M. *J. Chem. Phys.* **1998**, *109*, 7515.
- (479) Sawabe, K.; Koga, N.; Morokuma, K.; Iwasawa, Y. *J. Chem. Phys.* **1992**, *97*, 6871.
- (480) Barbosa, L. A. M. M.; Zhidomirov, G. M.; van Santen, R. A. *Catal. Lett.* **2001**, *77*, 55.
- (481) Schwarz, R.; Kubas, G. J. Unpublished results.
- (482) Neuhaus, A. H.; Glendening, E. D.; Streitwieser, A. *Organometallics* **1996**, *15*, 3688.
- (483) Ma, B.; Collins, C. L.; Schaefer, H. F., III. *J. Am. Chem. Soc.* **1996**, *118*, 870.
- (484) Xiao, Z. L.; Hauge, R. H.; Margrave, J. L. *J. Phys. Chem.* **1992**, *96*, 636.
- (485) Berkessel, A.; Schubert, T. J. S.; Muller, T. N. *J. Am. Chem. Soc.* **2002**, *124*, 8693.
- (486) Chan, B.; Radom, L. *J. Am. Chem. Soc.* **2005**, *127*, 2443.
- (487) Welch, G. C.; San Juan, R. R.; Masuda, J. D.; Stephan, D. W. *Science* **2006**, *314*, 1124. See also commentary: Kubas, G. J. *Science* **2006**, *314*, 1096.
- (488) Welch, G. C.; Stephan, D. W. *J. Am. Chem. Soc.* **2007**, *129*, 1880.
- (489) Watts, J. D.; Bartlett, R. J. *J. Am. Chem. Soc.* **1995**, *117*, 825.
- (490) Tague, T. J., Jr.; Andrews, L. *J. Am. Chem. Soc.* **1994**, *116*, 4970.
- (491) Fau, S.; Frenking, G. *Mol. Phys.* **1999**, *96*, 519.
- (492) Moroz, A.; Sweany, R. L. *Inorg. Chem.* **1992**, *31*, 5236.
- (493) Macchi, P.; Donghi, D.; Sironi, A. *J. Am. Chem. Soc.* **2005**, *127*, 16494.
- (494) Eckert, J.; Kubas, G. J.; Dianoux, A. J. *J. Chem. Phys.* **1988**, *88*, 466.
- (495) Eckert, J.; Blank, H.; Bautista, M. T.; Morris, R. H. *Inorg. Chem.* **1990**, *29*, 747.
- (496) Eckert, J.; Kubas, G. J.; Hall, J. H.; Hay, P. J.; Boyle, C. M. *J. Am. Chem. Soc.* **1990**, *112*, 2324.
- (497) Eckert, J. *Spectrochim. Acta* **1992**, *48A*, 363.
- (498) Eckert, J. *Trans. Am. Crystallogr. Assoc.* **1997**, *31*, 45.
- (499) Clot, E.; Eckert, J. *J. Am. Chem. Soc.* **1999**, *121*, 8855.
- (500) Webster, C. E.; Gross, C. L.; Young, D. M.; Girolami, G. S.; Schultz, A. J.; Hall, M. B.; Eckert, J. *J. Am. Chem. Soc.* **2005**, *127*, 15091.
- (501) (a) Antinolo, A.; Carrillo-Hermosilla, F.; Fajardo, M.; Garcia-Yuste, S.; Otero, A.; Camanyes, S.; Maseras, F.; Moreno, M.; Lledos, A.; Lluch, J. M. *J. Am. Chem. Soc.* **1997**, *119*, 6107. (b) Jalon, F. A.; Otero, A.; Manzano, B. R.; Villaseñor, E.; Chaudret, B. *J. Am. Chem.*

- Soc.* **1995**, *117*, 10123. (c) Sabo-Etienne, S.; Chaudret, B.; Abou el Makarim, H.; Barthelet, J.-C.; Daudey, J.-C.; Ulrich, S.; Limbach, H.-H.; Moise, C. *J. Am. Chem. Soc.* **1995**, *117*, 11602.
- (502) Prager, M.; Heidemann, A. *Chem. Rev.* **1997**, *97*, 2933.
- (503) Beaufile, J. P.; Crowley, T.; Rayment, R. K.; Thomas, R. K.; White, J. W. *Mol. Phys.* **1981**, *44*, 1257.
- (504) Nicol, J. M.; Eckert, J.; Howard, J. *J. Phys. Chem.* **1988**, *92*, 7117.
- (505) Eckert, J. *Physica* **1986**, *136B*, 150.
- (506) (a) Stephens, F. H.; Baker, R. T.; Matus, M. H.; Grant, D. J.; Dixon, D. A. *Angew. Chem., Int. Ed.* **2006**, *45*, 1. (b) Keaton, R. J.; Blacquiere, J. M.; Baker, R. T. *J. Am. Chem. Soc.* **2007**, *129*, 1844. (c) Jaska, C. A.; Temple, K.; Lough, A. J.; Manners, I. *J. Am. Chem. Soc.* **2003**, *125*, 9424. (d) Denney, M. C.; Pons, V.; Hebden, T. J.; Heinekey, D. M.; Goldberg, K. I. *J. Am. Chem. Soc.* **2006**, *128*, 12048.
- (507) Brown, C. M.; Yildirim, T.; Neumann, D. A.; Heben, M. J.; Gennett, T.; Dillon, A. C.; Alleman, J. L.; Fischer, J. E. *Chem. Phys. Lett.* **2000**, *329*, 311.
- (508) MacKinnon, J. A.; Eckert, J.; Coker, D. F.; Bug, A. L. *J. Chem. Phys.* **2001**, *114*, 10137.
- (509) Forster, P. M.; Eckert, J.; Chang, J.-S.; Park, S.-E.; Férey, G.; Cheetham, A. K. *J. Am. Chem. Soc.* **2003**, *125*, 1309.
- (510) Rosi, N. L.; Eckert, J.; Eddaoudi, M.; Vodak, D. T.; Kim, J.; O'Keeffe, M.; Yaghi, O. M. *Science* **2003**, *300*, 1127.
- (511) (a) Rowsell, J. L. C.; Yaghi, O. M. *Angew. Chem., Int. Ed.* **2005**, *44*, 4670. (b) Rowsell, J. L. C.; Eckert, J.; Yaghi, O. M. *J. Am. Chem. Soc.* **2005**, *127*, 14904.
- (512) Liu, Y.; Eubank, J. F.; Cairns, A. J.; Eckert, J.; Kravtsov, V. C.; Luebke, R.; Eddaoudi, M. *Angew. Chem., Int. Ed.* **2007**, *46*, 3278.
- (513) Spencer, E. C.; Howard, J. A. K.; McIntyre, G. J.; Rowsell, J. L. C.; Yaghi, O. M. *Chem. Commun.* **2006**, 278.
- (514) Hartman, M. R.; Peterson, V. K.; Liu, Y.; Kaye, S. S.; Long, J. R. *Chem. Mater.* **2006**, *18*, 3221.
- (515) Kubota, Y.; Takata, M.; Matsuda, R.; Kitaura, R.; Kitagawa, S.; Kato, K.; Sakata, M.; Kobayashi, T. C. *Angew. Chem., Int. Ed.* **2005**, *44*, 920.
- (516) Forster, P. M.; Eckert, J.; Heiken, B. D.; Parise, J. B.; Yoon, J. W.; Jhung, S. H.; Chang, J.-S.; Cheetham, A. K. *J. Am. Chem. Soc.* **2006**, *128*, 16846.
- (517) Rowsell, J. L. C.; Spencer, E. C.; Eckert, J.; Howard, J. A. K.; Yaghi, O. M. *Science* **2005**, *309*, 1350.
- (518) Georgiev, P. A.; Albinati, A.; Mojet, B. L.; Ollivier, J.; Eckert, J. *J. Am. Chem. Soc.* **2007**, *129*, 8086.
- (519) Xiao, B.; Wheatley, P. S.; Zhao, X.; Fletcher, A. J.; Fox, S.; Rossi, A. G.; Megson, I. L.; Bordiga, S.; Regli, L.; Thomas, K. M.; Morris, R. E. *J. Am. Chem. Soc.* **2007**, *129*, 1203.
- (520) Bordiga, S.; Vitillo, J. G.; Ricchiardi, G.; Regli, L.; Cocina, D.; Zecchina, A.; Arstad, B.; Bjorgen, M.; Hafizovic, J.; Lillerud, K. P. *J. Phys. Chem. B* **2005**, *109*, 18237.
- (521) Peterson, V. K.; Liu, Y.; Brown, C. M.; Kepert, C. J. *J. Am. Chem. Soc.* **2006**, *128*, 15578.
- (522) Dinca, M.; Dailly, A.; Liu, Y.; Brown, C. M.; Neumann, D. A.; Long, J. R. *J. Am. Chem. Soc.* **2006**, *128*, 16876.
- (523) *Chem. Eng. News* **2007**, *January 1*, 11.
- (524) Eckert, J.; et al. In preparation.

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